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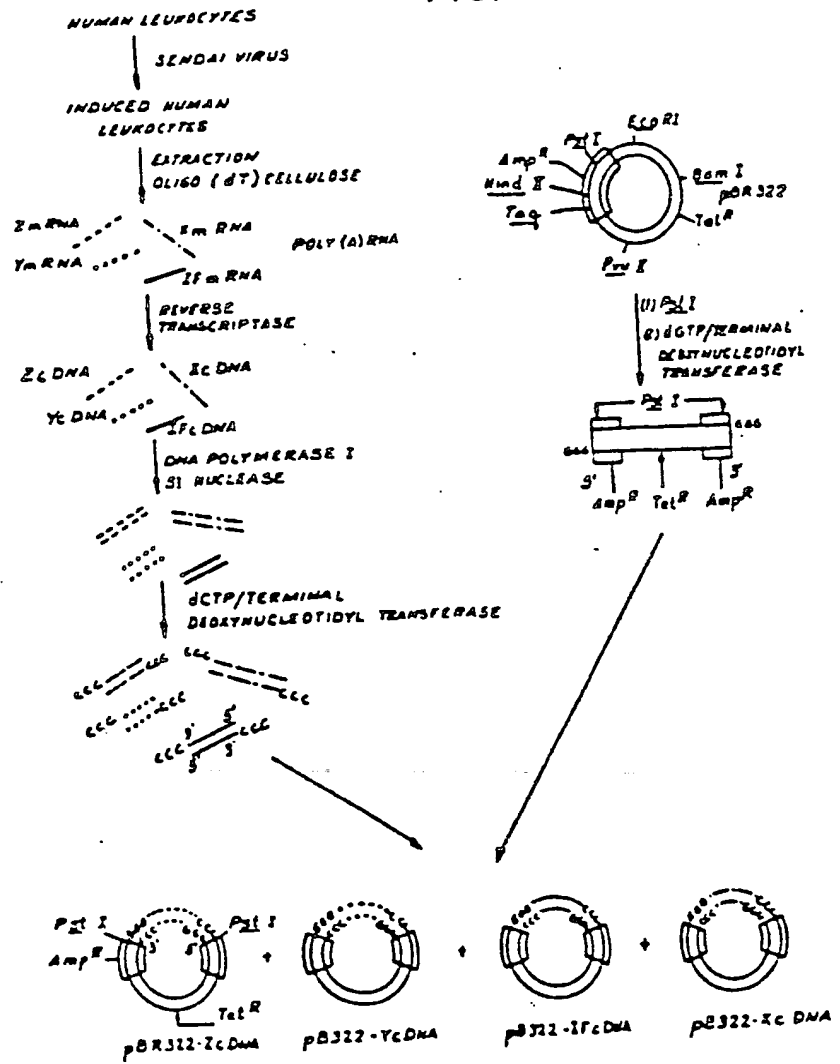
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(54) DNA sequences, recombinant DNA molecules and processes for producing human interferon-like polypeptides:

(57) DNA sequences, recombinant DNA molecules and hosts transformed with them which produce polypeptides displaying a biological or immunological activity of human interferon, the genes coding for these polypeptides and methods of making and using these molecules, hosts, genes and polypeptides. The DNA sequences are characterized in that they code for a polypeptide displaying a biological or immunological activity of human interferon. In appropriate hosts these DNA sequences and recombinant DNA molecules permit the production and identification of genes and polypeptides displaying a biological or immunological activity of human interferon and their use in antiviral and antitumor or anticancer agents.

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FIG. 1



DNA SEQUENCES, RECOMBINANT DNA MOLECULES AND PROCESSES
FOR PRODUCING HUMAN INTERFERON - LIKE POLYPEPTIDES

TECHNICAL FIELD OF INVENTION

This invention relates to DNA sequences, recombinant DNA molecules and processes for producing interferon and interferon-like polypeptides. More particularly, the invention relates to DNA sequences expressed in appropriate host organisms. The recombinant DNA molecules disclosed herein are characterized by DNA sequences that code for polypeptides having an immunological or biological activity of human leukocyte interferon. As will be appreciated from the disclosure to follow, the DNA sequences, recombinant DNA molecules and processes of this invention may be used in the production of polypeptides useful in antiviral and antitumor or anticancer agents and methods.

BACKGROUND ART

In this application the interferon nomenclature announced in Nature, 286, p. 2421 (July 10, 1980) will be used. This nomenclature replaces that used in our earlier applications from which this application claims priority. E.g., IF is now designated IFN and leukocyte interferon is now designated IFN- α .

Two classes of interferons ("IFN") are known to exist. Interferons of Class I are small, acid stable (glyco)-proteins that render cells resistant to viral

infection (A. Isaacs and J. Lindenmann, "Virus Interference I. The Interferon", Proc. Royal Soc. Ser. B., 147, pp. 258-67 (1957) and W. E. Stewart, II, The Interferon System, Springer-Verlag (1979) (hereinafter "The Interferon System")). Although to some extent cell specific (The Interferon System, pp. 135-45), IFNs are not virus specific. Instead IFNs protect cells against a wide spectrum of viruses.

Human interferons ("HuIFN") have been classified into three groups α , β and γ . HuIFN- α or leukocyte interferon is produced in human leukocyte cells and together with minor amounts of HuIFN- β (fibroblast interferon) in lymphoblastoid cells. HuIFN- α has been purified to homogeneity and characterized (e.g. M. Rubenstein et al., "Human Leukocyte Interferon: Production, Purification To Homogeneity And Initial Characterization" Proc. Natl. Acad. Sci. USA, 76, pp. 640-44 (1979)). It is heterogeneous in regard to size presumably because of the carbohydrate moiety. Two components have been described, one of 21000 to 22000 and the other of 15000-18000 molecular weight. The component of lower molecular weight has been reported to represent a non-glycosylated form. The smaller form of HuIFN- α has also been reported to retain most or all of its HuIFN- α activity (W. E. Stewart, II et al., "Effect Of Glycosylation Inhibitors On The Production And Properties Of Human Leukocyte Interferon", Virology, 97, pp. 473-76 (1979)). A portion of the amino acid sequence of HuIFN- α from lymphoblastoid cells and its amino acid composition have been reported (K.C. Zoon et al., "Amino Terminal Sequence Of The Major Component Of Human Lymphoblastoid Interferon", Science, 207, pp. 527-28 (1980) and M. Hunkapiller and L. Hood, personal communication (1980))

HuIFN- α has also been reported to exist in several different forms, e.g. British patent application 2,037,296A. These forms appear to differ from each other structurally and physiologically. No accepted nomenclature has been adopted for these forms of HuIFN- α . Therefore,

in this application each form will be referred to by a number after the general HuIFN- α designation, i.e., HuIFN- α 1 or HuIFN- α 3.

HuIFN- α may, like many human proteins, also be polymorphic. Therefore, cells of particular individuals may produce HuIFN- α species within the more general HuIFN- α group or forms within that group which are physiologically similar but structurally slightly different than the group or form of which it is a part. Therefore, while the protein structure of an HuIFN- α may be generally well-defined, particular individuals may produce a HuIFN- α that is a slight variation thereof, this allelic variation probably being less severe than the difference between the various forms of HuIFN- α .

HuIFN is usually not detectable in normal or healthy cells (The Interferon System, pp. 55-57). Instead, the protein is produced as a result of the cell's exposure to an IFN inducer. IFN inducers are usually viruses but may also be non-viral in character, such as natural or synthetic double-stranded RNA, intracellular microbes, microbial products and various chemical agents. Numerous attempts have been made to take advantage of these non-viral inducers to render human cells resistant to viral infection (S. Baron and F. Dianzani (eds.), Texas Reports On Biology And Medicine, 35 ("Texas Reports"), pp. 528-40 (1977)). These attempts have not been very successful. Instead, use of exogenous HuIFN itself is now preferred.

Interferon therapy against viruses and tumors or cancers has been conducted at varying dosage regimes and under several modes of administration (The Interferon System, pp. 305-321). For example, interferon has been effectively administered orally, by inoculation -- intravenous, intramuscular, intranasal, intradermal and subcutaneous --, and in the form of eye drops, ointments and sprays. It is usually administered one to three times daily in dosages of 10^4 to 10^7 units. The extent of the therapy depends on the patient and the condition being treated. For example, virus infections are usually

treated by daily or twice daily doses over several days to two weeks and tumors and cancers are usually treated by daily or multiple daily doses over several months or years. The most effective therapy for a given patient must of course be determined by the attending physician who will consider such well known factors as the course of the disease, previous therapy, and the patient's response to interferon in selecting a mode of administration and dosage regime.

As an antiviral agent, HuIFN has been used to treat the following: respiratory infections (Texas Reports, pp. 486-96); herpes simplex keratitis (Texas Reports, pp. 497-500); acute hemorrhagic conjunctivitis (Texas Reports, pp. 501-10); varicella zoster (Texas Reports, pp. 511-15); cytomegalovirus infection (Texas Reports, pp. 523-27); and hepatitis B (Texas Reports, pp. 516-22). See also The Interferon System, pp. 307-19. However, large scale use of IFN as an antiviral agent requires larger amounts of IFN than heretofore have been available.

HuIFN has other effects in addition to its antiviral action. For example, it antagonizes the effect of colony stimulating factor, inhibits the growth of hemopoietic colony-forming cells and interferes with the normal differentiation of granulocyte and macrophage precursors (Texas Reports, pp. 343-49). It also inhibits erythroid differentiation in DMSO-treated Friend leukemia cells (Texas Reports, pp. 420-28). HuIFN may also play a role in regulation of the immune response. Depending upon the dose and time of application in relation to antigen, HuIFN- α can be both immunopotentiating and immunosuppressive in vivo and in vitro (Texas Reports, pp. 357-69). In addition, specifically sensitized lymphocytes have been observed to produce HuIFN- α after contact with antigen. Such antigen-induced HuIFN- α could therefore be a regulator of the immune response, affecting both circulating antigen levels and the expression of cellular immunity (Texas Reports, pp. 370-74). HuIFN is also known to enhance the activity of killer lymphocytes and

antibody-dependent cell-mediated cytotoxicity (R. R. Herberman et al., "Augmentation By Interferon Of Human Natural And Antibody Dependent Cell-Mediated Cytotoxicity", Nature, 277, pp. 221-23 (1979); P. Beverley and D. Knight, "Killing Comes Naturally", Nature, 278, pp. 119-20 (1979); Texas Reports, pp. 375-80). Both of these species are probably involved in the immunological attack on tumor cells.

Therefore, in addition to its use as a human antiviral agent, HuIFN has potential application in anti-tumor and anticancer therapy (The Interferon System, pp. 319-21). It is now known that IFNs affect the growth of many classes of tumors in many animals (The Interferon System, pp. 292-304). They, like other antitumor agents, seem most effective when directed against small tumors. The antitumor effects of animal IFN are dependent on dosage and time but have been demonstrated at concentrations below toxic levels. Accordingly, numerous investigations and clinical trials have been and continue to be conducted into the antitumor and anticancer properties of IFNs. These include treatment of several malignant diseases such as osteosarcoma, acute myeloid leukemia, multiple myeloma and Hodgkin's disease (Texas Reports, pp. 429-35). In addition, HuIFN has recently been shown to cause local tumor regression when injected into subcutaneous tumoral nodules in melanoma- and breast carcinoma-affected patients (T. Nemoto et al., "Human Interferons And Intralesional Therapy Of Melanoma And Breast Carcinoma", Amer. Assoc. For Cancer Research, Abs. nr. 994, p. 246 (1979)). Although the results of these clinical tests are encouraging, the antitumor and anticancer applications of IFN have been severely hampered by lack of an adequate supply of purified IFN.

Today, HuIFN- α is produced either through human cells grown in tissue culture or through human leukocytes collected from blood donors. 2.6×10^9 IU of crude HuIFN- α have been reported from 800 l of cultured Namalva cells (P. J. Bridgen et al., supra). At very large blood

centers, e.g., the Finnish Red Cross Center in Helsinki, Finland, the production capacity is about 10^{11} IU of crude HuIFN- α annually. Since dosage is typically 3×10^6 IU per patient per day, these sources are not adequate to provide the needed commercial quantities of HuIFN- α . Therefore, production of HuIFN- α by other procedures is desirable. Because the specific activity of IFN- α is high, in the order of 4.0×10^8 to 10^9 IU/mg, the amount of HuIFN- α required for commercial applications is low. For example, 100 grams of pure HuIFN- α would provide between 3 and 30 million doses.

Recent advances in molecular biology have made it possible to introduce the DNA coding for specific nonbacterial eukaryotic proteins into bacterial cells. In general, with DNA other than that prepared via chemical synthesis, the construction of such recombinant DNA molecules comprises the steps of producing a single-stranded DNA copy (cDNA) of a purified messenger RNA (mRNA) template for the desired protein; converting the cDNA to double-stranded DNA; linking the DNA to an appropriate site in an appropriate cloning vehicle to form a recombinant DNA molecule and transforming an appropriate host with that recombinant DNA molecule. Such transformation may permit the host to produce the desired protein.

Several non-bacterial proteins and genes have been obtained in E. coli using recombinant DNA technology. These include a protein displaying rat proinsulin antigenic determinants (L. Villa-Komaroff et al., "A Bacterial Clone Synthesizing Proinsulin", Proc. Natl. Acad. Sci. USA, 75, pp. 3727-31 (1978)), rat growth hormone (P. H. Seeburg et al., "Synthesis Of Growth Hormone By Bacteria", Nature, 276, pp. 795-98 (1978)), mouse dihydrofolate reductase (A.C.Y. Chang et al., "Phenotypic Expression In E. coli Of A DNA Sequence Coding For Mouse Dihydrofolate Reductase", Nature, 275, pp. 617-24 (1978)), human somatostatin (K. Itakura et al., "Expression In Escherichia coli Of A Chemically Synthesized Gene For The Hormone Somatostatin",

Science, 198, pp. 1056-63 (1977)); European patent applications 0,001,929, 0,001,930, and 0,001,931 and cognate applications in other countries), the A and B polypeptide chains of human insulin (D. V. Goeddel et al., "Expression In Escherichia coli Of Chemically Synthesized Genes For Human Insulin", Proc. Natl. Acad. Sci. USA, 76, pp. 106-10 (1979) and the European and related patent specifications, supra), antigens of human hepatitis B virus (C. J. Burrell et al., "Expression In Escherichia coli: Of Hepatitis B Virus DNA Sequences Cloned In Plasmid pBR322", Nature, 279, pp. 43-7 (1979) and M. Pasek et al., "Hepatitis B Virus Genes And Their Expression In E. coli", Nature, 282, pp. 575-79 (1979)), human growth hormone (D. V. Goeddel et al., "Direct Expression In Escherichia coli Of A DNA Sequence Coding For Human Growth Hormone", Nature, 281, pp. 544-51 (1979)), SV40 t antigen (T. M. Roberts et al., "Synthesis Of Simian Virus 40 t Antigen In Escherichia coli", Proc. Natl. Acad. Sci. USA, 76, pp. 5596-600 (1979)), and human fibroblast interferon (HuIFN- β) (T. Taniguchi et al., "Construction And Identification Of A Bacterial Plasmid Containing The Human Fibroblast Interferon Gene Sequence", Proc. Japan Acad., 55, Ser. B, pp. 464-69 (1979) together with personal communication 1980).

None of these recombinant DNA processes, however is directed, as is this invention, toward the synthesis of HuIFN- α . This is the problem to which the present invention is addressed. Its solution is not facilitated as were the above described recombinant DNA schemes by the availability of the sequence information required to prepare a synthetic gene (e.g., somatostatin) or of a cell type or virus rich in a particular DNA sequence (e.g., hepatitis viral antigen) or mRNA species (e.g., rat insulin) which allows preparation and identification of bacterial clones containing the desired hybrid DNA, or of a system allowing the selection of E. coli hosts that express the desired protein (e.g., mouse dihydrofolate reductase). Neither is it aided by the report of a

plasmid which is said to contain a DNA sequence that hybridizes to a mRNA from a poly(A) RNA, that mRNA producing HuIFN- β activity in oocytes (e.g., fibroblast interferon). Nor is the solution of the present invention addressed as is the apparent suggestion of Research Disclosure No. 18309, pp. 361-62 (1979) to preparing pure or substantially pure HuIFN- α mRNA before cloning of the HuIFN- α gene.

Finally, it should be recognized that the selection of a DNA sequence or the construction of a recombinant DNA molecule which hybridizes to a mRNA from polyA RNA, that mRNA producing HuIFN activity in oocytes, is not sufficient to demonstrate that the DNA sequence or the hybrid insert of the recombinant DNA molecule corresponds to HuIFN. Instead, only the production of a polypeptide that displays an immunological or biological activity of HuIFN can actually demonstrate that the selected DNA sequence or constructed recombinant DNA molecule corresponds to HuIFN. More importantly, it is only after HuIFN activity is shown that the DNA sequence, recombinant DNA molecule or sequences related to them may be employed to select other sequences corresponding to HuIFN in accordance with this invention.

It will therefore be appreciated from the foregoing that the problem of producing HuIFN- α with the use of recombinant DNA technology is much different than any of the above described processes. Here, a particular DNA sequence of unknown structure -- that coding for the expression of HuIFN- α in an appropriate host -- must be found in and separated from a highly complex mixture of DNA sequences in order for it to be used in the production of HuIFN- α . Moreover, this location and separation problem is exacerbated by the predicted exceedingly low concentration of the desired DNA sequence in the complex mixture and the lack of an effective means for rapidly analyzing the many DNA sequences of the mixture to select and separate the desired sequence.

DISCLOSURE OF THE INVENTION

The present invention solves the problems referred to by locating and separating DNA sequences that code for the expression of HuIFN- α in an appropriate host and thereby providing DNA sequences, recombinant DNA molecules and methods by means of which a host is transformed to produce a polypeptide displaying an immunological or biological activity of human leukocyte interferon.

By virtue of this invention, it is possible to obtain polypeptide(s) displaying an immunological or biological activity of HuIFN- α for use in antiviral, antitumor or anticancer agents and methods. This invention allows the production of these polypeptides in amounts and by methods hitherto not available.

As will be appreciated from the disclosure to follow, the DNA sequences and recombinant DNA molecules of the invention are capable of directing the production, in an appropriate host, of a polypeptide displaying an immunological or biological activity of HIFN- α . Replication of these DNA sequences and recombinant DNA molecules in an appropriate host also permits the production in large quantities of genes coding for these polypeptides. The molecular structure and properties of these polypeptides and genes may be readily determined. The polypeptides and genes are useful, either as produced in the host or after appropriate derivatization or modification, in compositions and methods for detecting and improving the production of these products themselves and for use in antiviral and antitumor or anticancer agents and methods.

This process is therefore distinguishable from the prior processes, above mentioned, in that this process, contrary to the noted prior processes, involves the preparation and selection of DNA sequences and recombinant DNA molecules which contain appropriate DNA sequences which code for at least one polypeptide displaying an immunological or biological activity of HuIFN- α .

It will be appreciated from the foregoing that a basic aspect of this invention is the provision of a DNA sequence which is characterized in that it codes for a polypeptide displaying an immunological or biological activity of HuIFN and is selected from the group consisting of the DNA inserts of Z-pBR322 (Pst)/HcIF-4c, Z-pBR322(Pst)/HcIF-2h, Z-pBR322(Pst)/HcIF-SN35, Z-pBR322(Pst)/HcIF-SN42, Z-pKT287(Pst)/HcIF-2h-AH6, DNA sequences which hybridize to any of the foregoing DNA inserts, DNA sequences, from whatever source obtained, including natural, synthetic or semi-synthetic sources, related by mutation, including single or multiple, base substitutions, deletions, insertions and inversions to any of the foregoing DNA sequences or inserts, and DNA sequences comprising sequences of codons which on expression code for a polypeptide displaying similar immunological or biological activity to a polypeptide coded for on expression of the codons of any of the foregoing DNA sequences and inserts and that these sequences permit the production of interferon and interferon-like polypeptides in hosts.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a schematic outline of one embodiment of a process of this invention for preparing a mixture of recombinant DNA molecules, some of which are characterized by inserted DNA sequences that code for polypeptides of this invention.

Figure 2 is a schematic outline of the initial clone screening process of this invention.

Figure 3 is a schematic outline of one embodiment of a clone screening process using DNA sequences prepared in accordance with the invention.

Figure 4 is a restriction map of one of the clones of the invention, the absolute position of each restriction site in this clone is not depicted. Figures 8-10 display more absolute positions for these restriction sites.

Figure 5 is a schematic outline of the process of determining the orientation of a DNA insert in one recombinant DNA molecule of this invention.

Figure 6 displays the partial nucleotide sequence of some cloning vehicles useful in accordance with this invention.

Figure 7 displays the results of a Sephadex G-100 fractionation of supernatant prepared from a bacterial culture of this invention.

Figures 8-10 display the nucleotide sequence of a DNA insert to a recombinant DNA molecule of this invention. The sequence is numbered from the nucleotide following the polyG 5' tail to the nucleotide before the polyA residues and polyC 3' tails. Nucleotides 57-125 represent a signal sequence and nucleotides 126-626 represent the "mature" interferon and the stop codon. The amino acid sequence of the signal sequence is depicted above its nucleotide sequence in lower case letters and the amino acid sequence of the "mature" inteferon is depicted above its nucleotide sequence in upper case letters. Various restriction endonuclease recognition sites in this gene are also depicted in Figures 8-10, these sites being more absolutely located than those set forth in Figure 4.

Figure 11 is a schematic comparison of the restriction maps of two DNA inserts of recombinant DNA molecules of this invention.

Figures 12-16 display the nucleotide sequences of two DNA inserts of recombinant DNA molecules of this invention. The sequences are numbered from the nucleotide following the polyG 5' tail to the nucleotide before the polyA residues and polyC 3' tails. The amino acid sequence of the signal sequence for each of these inserts is depicted above its respective nucleotide sequence in lower case letters and the amino acid sequence of the "mature" interferon is depicted above its nucleotide sequence in upper case letters.

Figure 17 displays a partial restriction map of Z-pBR322(Pst)/HcIF-II-206 and the sequencing strategy employed to determine the nucleotide sequence of the Hif-II-206 fragment displayed in Figures 12-16.

Figure 18 displays the partial restriction maps of a series of hybrid phages which hybridize to the Hif-2h fragment.

Figure 19 displays a partial restriction map of the hybrid insert of Z-pBR322Pst/HchrIF-35HB α and the sequencing strategy employed to determine its nucleotide sequence.

Figures 20-23 display the nucleotide sequence of the HchrIF-35HB α fragment and the amino acid sequence derived from it.

Figure 24 displays partial linkage maps for HuIFN- α related genes. The arrows show regions which form R-loops with induced leukocyte poly(A) RNA. The hatched box (chr-16) indicates the sequence which was inferred from blotting experiments, but was not revealed by R-loop mapping.

Figure 25 is a schematic outline of the construction of plasmid C8-IFN- α 1.

Figure 26 is a schematic outline of the construction of plasmid LAC-AUG(α 2).

Figure 27 displays the reconstruction of the AUG initiation codon and the CYS initial codon in the construction of LAC-AUG(α 2).

Figure 28 is a schematic outline of the construction of plasmid C8-IFN- α 2 and the hybrid molecules I, II, III and IV.

Figures 29-32 display the nucleotide sequence and amino acid sequence encoded thereby for IFN- α 4b and its signal sequence.

BEST MODE OF CARRYING OUT THE INVENTION

In order that the invention herein described may be more fully understood, the following detailed description is set forth.

In the description the following terms are employed:

Nucleotide--A monomeric unit of DNA or RNA consisting of a sugar moiety (pentose), a phosphate, and a nitrogenous heterocyclic base. The base is linked to the sugar moiety via the glycosidic carbon (1' carbon of the pentose). That combination of a base and a sugar is called a nucleoside. Each nucleotide is characterized by its base. The four DNA bases are adenine ("A"), guanine ("G"), cytosine ("C") and thymine ("T"). The four RNA bases are A, G, C and uracil ("U").

DNA Sequence--A linear array of nucleotides connected one to the other by phosphodiester bonds between the 3' and 5' carbons of adjacent pentoses.

Codon--A DNA sequence of three nucleotides (a triplet) which encodes through mRNA an amino acid, a translation start signal or a translation termination signal. For example, the nucleotide triplets TTA, TTG, CTT, CTC, CTA and CTG encode for the amino acid leucine ("Leu"), TAG, TAA and TGA are translation stop signals and ATG is a translation start signal.

Reading Frame--The grouping of codons during translation of mRNA into amino acid sequences. During translation the proper reading frame must be maintained. For example, the sequence GCTGGTTGTAAG may be translated in three reading frames or phases, each of which affords a different amino acid sequence:

GCT GGT TGT AAG--Ala-Gly-Cys-Lys

G CTG GTT GTA AG--Leu-Val-Val

GC TGG TTG TAA G--Trp-Leu-(STOP)

Polypeptide--A linear array of amino acids connected one to the other by peptide bonds between the α -amino and carboxy groups of adjacent amino acids.

Genome--The entire DNA of a cell or a virus. It includes inter alia the structural genes coding for the polypeptides of the substance, as well as operator, promoter and ribosome binding and interaction sequences, including sequences such as the Shine-Dalgarno sequences.

Structural Gene--A DNA sequence which encodes through its template or messenger RNA ("mRNA") a sequence of amino acids characteristic of a specific polypeptide.

Transcription--The process of producing mRNA from a structural gene.

Translation--The process of producing a polypeptide from mRNA.

Expression--The process undergone by a structural gene to produce a polypeptide. It is a combination of transcription and translation.

Plasmid--A non-chromosomal double-stranded DNA sequence comprising an intact "replicon" such that the plasmid is replicated in a host cell. When the plasmid is placed within a unicellular organism, the characteristics of that organism may be changed or transformed as a result of the DNA of the plasmid. For example, a plasmid carrying the gene for tetracycline resistance (Tet^R) transforms a cell previously sensitive to tetracycline into one which is resistant to it. A cell transformed by a plasmid is called a "transformant".

Phage or Bacteriophage--Bacterial virus many of which consist of DNA sequences encapsidated in a protein envelope or coat ("capsid").

Cloning Vehicle--A plasmid, phage DNA or other DNA sequences which are able to replicate in a host cell, which are characterized by one or a small number of endonuclease recognition sites at which such DNA sequences may be cut in a determinable fashion without attendant loss of an essential biological function of the DNA, e.g., replication, production of coat proteins or loss of promoter or binding sites, and which contain a marker suitable for use in the identification of transformed cells, e.g., tetracycline resistance or ampicillin resistance. A cloning vehicle is often called a vector.

Cloning--The process of obtaining a population of organisms or DNA sequences derived from one such organism or sequence by asexual reproduction.

Recombinant DNA Molecule or Hybrid DNA--A molecule consisting of segments of DNA from different genomes which have been joined end-to-end outside of living cells and have the capacity to infect some host cell and be maintained therein.

Expression Control Sequence--A sequence of nucleotides that controls and regulates expression of structural genes when operatively linked to those genes. They include the lac system, the trp system, major operator and promoter regions of phage λ , the control region of fd coat protein and other sequences known to control the expression of genes of prokaryotic or eukaryotic cells and their viruses.

Referring now to Figure 1, we have shown therein a schematic outline of one embodiment of a process for preparing a mixture of recombinant DNA molecules, some of which are characterized by inserted DNA sequences that code for polypeptides having an immunological or biological activity of human leukocyte interferon.

PREPARATION OF POLY(A) RNA CONTAINING HUMAN INTERFERON mRNA (IFN- α mRNA)

Human leukocytes were induced for 5 hours at 37°C with Sendai virus and extracted to yield a poly(A) RNA mixture containing human leukocyte interferon mRNA ("HuIFN- α mRNA"). Induction was by the Cantell procedure (The Interferon System, pp. 130-31 and the references cited therein). The poly(A) RNA mixture is illustrated without regard to its actual proportions in Figure 1. Induced leukocytes were harvested and 10^{11} cells were resuspended in 1 l of a solution containing 8 g NaCl, 0.2 g KCl, 1.15 g $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$ and 0.2 g KH_2PO_4 dissolved in 1 l of water ("PBS") and added slowly with vigorous stirring to 17 l 20 mM Tris-HCl (pH 7.5), 1 mM EDTA ("TE buffer"), 2% sodium dodecyl sulfate ("SDS") in a 50 l separatory funnel. Self-digested Pronase (Calbiochem) was added to 200 $\mu\text{g}/\text{ml}$ and the solution stirred for 1 h

at room temperature. 10^6 counts/minute ("cpm") of ^{125}I -globin mRNA were added as a marker for recovery of the poly(A) RNA and to control for mRNA degradation during subsequent steps. 2M Tris-HCl (pH 9) in an amount equal to 1/20 of the total volume ("1/20 vol") was added and the mixture extracted with vigorous stirring with 15 l of redistilled phenol for 10 min. Three l chloroform were added and the mixture stirred for 5 min. After allowing 30 min for phase separation, the aqueous phase was removed and extracted again with phenol and chloroform. The resultant aqueous phase, totalling 19.1 l, was combined with 60 g SDS. Nucleic acids were precipitated from the aqueous phase with 1/10 vol 3M sodium acetate (pH 5.5) and 2 vol ethanol.

After storage overnight at -20°C , the fibrous nucleic acid precipitate was removed by filtration through a plastic tea sieve. This material was then stirred with 200 ml TNE (50 mM Tris-HCl (pH 7.5), 100 mM NaCl, 5 mM EDTA) containing 0.5% SDS. It subsequently dissolved on addition of a further 350 ml of that solution. The non-fibrous precipitate was collected by centrifugation in 1 l Sorvall bottles in a Sorvall RC-3 centrifuge for 15 min at 5,000 rpm and dissolved in 350 ml TNE containing 0.5% SDS. The two TNE solutions were combined, extracted 3 times with 1 vol phenol, 3 times with 1/2 vol ether and 3 times with 1 vol ether. RNA recovery from the aqueous phase totalled 775 mg, as measured by absorbance at 260 nm.

Isolation of the poly(A) RNA mixture was achieved by repeated batch adsorption to oligo(dT) cellulose (type 7, P-L Biochemicals, Inc.). 2.7 grams oligo(dT) cellulose were added to 500 ml, i.e., about half of the RNA-containing solution described above. After stirring for 1 h at room temperature to effect adsorption of the poly(A) RNA to the oligo(dT) cellulose, the cellulose and the mixture of mRNAs bound to it were collected by centrifugation and washed once with 50 ml TNE and a second time with 15 ml TNE. The bound poly(A) RNA was then eluted by

five successive washes with 2 ml H₂O. The yield was 860 µg poly(A) RNA as measured by optical density (Preparation A). The supernatant RNA solution from the first adsorption was subjected to two further adsorption cycles, exactly as described above. The second and third adsorptions yielded 600 µg and 170 µg RNA respectively and were combined (Preparation B).

RNA was assayed for HuIFN-αmRNA by injection into Xenopus laevis oocytes (The Interferon System, pp. 93-95): RNA was dissolved in 15 mM Tris-HCl (pH 7.5), 88 mM NaCl ("TNK buffer") to give a concentration of about 1 mg/ml. Fifty nl of this solution were injected into each of 50 oocytes. The oocytes were incubated overnight at room temperature in Barth medium (Gurdon, J. Embryol and Exper. Morph., 20, pp. 401-414 (1968) and Barth, J. Embryol and Exper. Morph., 7, pp. 210-222 (1959)). The incubated oocytes were then rinsed and homogenized with a Pasteur pipette in a 1.5 ml Eppendorf centrifuge tube in 0.5 ml 52 mM Tris glycine buffer (pH 8.9). The mixture was centrifuged for 2 min in an Eppendorf centrifuge and the supernatant was drawn off and frozen at -20°C for assay. IFN-α activity was determined by the plaque reduction assay described by H. Strander and K. Cantell, "Production Of Interferon By Human Leukocytes In Vitro", Ann. Med. exp. Fenn., 44, pp. 265-73 (1966). One unit IFN-α reduces virus plaques by 50%. The potency of an IFN-α preparation is expressed relative to the human reference HuIFN-α 69/19 (International Symposium on Standardization of Interferon and Interferon Inducers, 1969). Alternatively, the assay was based on the reduction of cytopathic effect, essentially as described by W. E. Stewart, II and S. E. Sulkin, "Interferon Production In Hamsters Experimentally Infected With Rabies Virus", Proc. Soc. Exp. Biol. Med., 123, pp. 650-3 (1966), except that human CCL-23 cells were used and that challenge was with Mengo virus. The oocyte extracts had 300 IU of IFN-α activity per µg of RNA injected. In later assays incubation of injected oocytes

was for 48 hrs and only the incubation medium was assayed because most of the interferon is excreted by the oocytes (A. Colman and J. Morser, "Export Of Proteins From Oocytes Of Xenopus laevis", Cell, 17, pp. 517-26 (1979)). For further purification of the poly(A) RNA sufficient 0.5 M ethylene diamine tetraacetic acid ("EDTA") was added to the poly(A) RNA Preparation A to bring the concentration to 5 mM EDTA. The resultant solution was extracted twice with an equal vol of TNE-saturated phenol and 5 times with an equal vol of ether. It was then passed through a 0.1-ml Chelex-100 Bio-Rad column, heated for 90 sec at 100°C and layered onto a 13-ml 5-23% sucrose gradient containing 50 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.2 M NaCl. 10,000 cpm of 5'-terminally ³²P-labeled DNA fragments produced by simultaneous digestion of pBR322 with restriction enzymes HindIII and PstI (New England Biolabs), were added as size markers. Centrifugation was in an SW40 rotor at 10°C and 35,000 rpm for 16 h. Fractions (0.6 ml) were collected with an ISCO gradient collector at 1 ml/min. The fractions were assayed for HuIFN- α mRNA as described above and their position relative to the ³²P-DNA markers was noted for future reference. In subsequent centrifugations, HuIFN- α mRNA-containing fractions were identified relative to the markers. The fractions with HuIFN- α mRNA activity contained 80 μ g of poly(A) RNA. They were mixed with 2 vol TNE containing 0.5% SDS and 0.02% polyvinyl sulfate (in later preparations polyvinyl sulfate was omitted) and applied to a 50- μ l oligo(dT) cellulose column. After washing the column as described above, 40 μ g of the RNA mixture were eluted with 4 washes of 0.6 ml sterile distilled water. After ethanol precipitation, the RNA was dissolved to 1 mg/ml in 0.5 mM EDTA.

An assay for HuIFN- α mRNA activity was carried out as described above on a portion of the poly(A) RNA precipitate. It had a specific activity of 3600 IU interferon/ μ g of RNA injected. Therefore, the sucrose gradient had enriched the poly(A) RNA about 10-fold in

regard to HuIFN- α mRNA. In a subsequent, similar preparation about a 40-fold enrichment was obtained. Preparation B was purified similarly and, since it had a similar specific activity as Preparation A, the two were pooled.

At this point it should be recognized that even the poly(A) RNA product obtained from the sucrose gradient contains a very large number of different mRNA's. Except for the mRNA specific for IFN- α , the other mRNAs are undesirable contaminants (Figure 1). Unfortunately, these contaminant RNAs behave similarly to HuIFN- α mRNA throughout the remainder of the cloning process of this invention. Therefore, their presence in the poly(A) RNA will result in the ultimate preparation of a large number of unwanted bacterial clones which contain genes that code for polypeptides other than IFN- α . This contamination presents complex screening problems in the isolation of the desired IFN- α hybrid clones. In the case of IFN- α , the screening problem is further exacerbated by the lack of a sufficiently purified sample of HuIFN- α mRNA or DNA or portion thereof to act as a screening probe for the identification of the desired clones. Therefore, the screening process for the IFN- α clones is very time-consuming and difficult. Further, because only a very small percentage of IFN- α clones themselves are expected to express IFN- α in a biologically active or immunologically active form, the isolation of an active clone is a "needle in a haystack" screening process.

Advantageously, we may use recombinant DNA technology to provide a purified sample of HuIFN- α mRNA or cDNA or a portion thereof. This purified mRNA or cDNA can be used to screen rapidly very large numbers of bacterial clones and thereby markedly increase the probability of isolating a clone which expresses IFN- α in an active form.

SYNTHESIS OF cDNA MIXTURE CONTAINING HuIFN- α cDNA

The poly(A) RNA enriched for IFN- α mRNA (Preparation A+B) was used as a template to prepare single-stranded complementary DNA (cDNA) (Figure 1) (Cf, A. Efstratiadis *et al.*, "Full Length And Discrete Partial Reverse Transcripts Of Globin And Chorion mRNAs", *Cell*, 4, pp. 367-78 (1975) and references cited therein). The 800- μ l reaction mixture contained 40 mM Tris-HCl (pH 7.5), 30 mM NaCl, 5mM MgCl₂, 0.5 mM DTT (Cal-Biochem), 20 μ g/ml oligo(dT) 12-18 (P&L Biochemicals), 5 mM dGTP (Schwarz), dCTP (Laevosan) and dTTP (Sigma), 5 mM ³²P-dATP (NEN, specific activity 100,000 cpm/nmole), 60 μ g/ml poly(A) RNA and 280 units avian myeloblastosis virus (AMV) reverse transcriptase (a gift from Life Sciences, Inc., St. Petersburg, Florida). After incubation for 1 h at 37°C, 0.5 M EDTA and 20% SDS (recrystallized) were added to 10 mM EDTA and 0.1% SDS. The mixture was extracted with 1 vol phenol (distilled). The phenol phase was washed with 200 μ l 200 mM Tris-HCl (pH 7.5), 1 mM EDTA and 0.1% SDS, and the aqueous phases combined. These were extracted with an equal vol ether (Fluka, pro anal.) and chromatographed on a 5-ml Sephadex G-100 column in TNE. Fractions of 0.1 ml were collected at 0.3 ml/min. Fractions displaying radioactivity (as measured by Cerenkov radiation) were combined and 3 M sodium acetate added to 0.3M. The nucleic acids were precipitated with 2.5 vol of ethanol. After storage overnight at -20°C, the samples were centrifuged and the supernatant discarded. The precipitate was dissolved in 180 μ l distilled water and transferred to a siliconized Eppendorf tube. 20 μ l 5M NaOH were added and the mixture kept at room temperature for 40 min. 20 μ l of 5M sodium acetate, 100 μ l distilled water and 500 μ l ethanol were added. After cooling overnight at -20°C, the resulting precipitate was collected by centrifugation at a force equivalent to 10,000 times the force of gravity (10000 xg) for 20 min at 0°C. The yield of single-stranded cDNA was 10 μ g.

Again, it is to be understood that the single-stranded cDNA product prepared above is in reality a complex mixture of a large number of different cDNAs transcribed from the corresponding mRNAs present in the poly(A) RNA mixture (Figure 1). Only a very few of these cDNAs are IFN- α related, i.e., HiIFN- α cDNAs. Another factor also acts to complicate the cDNA mixture--each mRNA species of the poly(A) RNA mixture is usually not transcribed completely. Instead, for each mRNA species the transcription process may stop before the end of the mRNA is reached. Therefore, a large variety of cDNA species may be produced from each mRNA species (not shown in Figure 1). Each species will behave more or less similarly in the subsequent cloning process so that bacterial clones will be produced which contain recombinant DNA molecules having only a fragment of the gene for a particular protein. The presence of these fragment-containing clones even further complicates the final clone screening process.

The sizes of the various single-stranded cDNAs were determined by electrophoresis of a small aliquot on a alkaline 2% agarose gel using 30 mM NaOH, 2 mM EDTA as electrolyte (M. W. McDonnell et al., "Analysis Of Restriction Fragments Of T7 DNA And Determination Of Molecular Weights By Electrophoresis In Neutral And Alkaline Gels", J. Mol. Biol., 110, pp. 119-46 (1977)). The ^{32}P -cDNA had a length of 600-1000 nucleotides, relative to single-stranded globin cDNA and ^{32}P -labeled DNA fragments used as size markers.

PREPARATION OF DOUBLE-STRANDED cDNA

The single-stranded cDNA may be rendered double-stranded by treatment with DNA polymerase I (T. Maniatis et al., "Amplification And Characterization Of A β -Globin Gene Synthesized In Vitro", Cell, 8, pp. 163-82 (1976)). The precipitated single-stranded cDNA from above was dissolved in 200 μl H_2O , heated at 100°C for

2 min and incubated in 500 μ l 0.1 M heat denatured potassium phosphate buffer (pH 6.9), 10 mM MgCl_2 , 10 mM DTT (Calbiochem), 1 mM each of dATP (Merck), dGTP (Schwarz) and dCTP (Laevosan), 1 mM ^3H -dTTP (NEN, specific activity 100,000 cpm/nmole) and 150 units/ml of *E. coli* DNA polymerase I (Boehringer-Mannheim). After 6.5 h at 15°C, 0.5 M EDTA and 20% SDS were added to 10 mM EDTA and 0.1% SDS. The mixture was then extracted with 500 μ l phenol and the phenol phase was reextracted with 250 μ l 20 mM Tris-HCl (pH 7.5), 5 mM EDTA ("TE buffer"). The two aqueous phases were combined and chromatographed on a 5-ml Sephadex G-100 column under the same conditions described previously. Sodium acetate (3M) was added to 0.3 M and 2.5 vol ethanol were mixed into precipitate the DNA. A total of 13 μ g DNA was recovered.

The DNA was treated with nuclease S_1 , prepared by the method of R. C. Wiegand *et al.*, "Specificity Of The S_1 Nuclease From *Aspergillus oryzae*", *J. Biol. Chem.*, 250, pp. 8848-55 (1975). The precipitated DNA was dissolved in 250 μ l S_1 buffer (0.2 M NaCl, 50 mM sodium acetate (pH 4.5), 10 mM zinc sulfate) and warmed at 37°C for 30 min. 1.5 μ l S_1 enzyme (11 units/ μ l) were added and the mixture incubated at 37°C for 30 min. SDS and EDTA were added to 0.1% SDS and 5 mM EDTA, and the mixture was extracted with 250 μ l phenol. The phenol phase was washed with 100 μ l TE buffer. The aqueous phases were combined and chromatographed on a Sephadex G-100 (Pharmacia) column in TNE; 0.1-ml fractions were collected at 0.3 ml/min and the Cerenkov radiation of each fraction was determined. 8 μ g of double-stranded cDNA were recovered after precipitation with ethanol and sodium acetate as above.

Again, it must be recognized that the double-stranded cDNA produced above is a mixture of a large number of cDNAs and fragments thereof, only a very few of which are HuIFN- α cDNA or its fragments (Figure 1).

CLONING OF DOUBLE-STRANDED DNA

A wide variety of host/cloning vehicle combinations may be employed in cloning the double-stranded cDNA prepared as above described. For example, useful cloning vehicles may consist of segments of chromosomal, non-chromosomal and synthetic DNA sequences, such as various known bacterial plasmids, e.g., plasmids from E. coli including col E1, pCR1, pBR322 and their derivatives, wider host range plasmids, e.g., RP4, phage DNA, e.g., the numerous derivatives of phage λ , e.g., NM 989, and vectors derived from combinations of plasmids and phage DNAs such as plasmids which have been modified to employ phage DNA or other expression control sequences or yeast plasmids such as the 2 μ plasmid or derivatives thereof. Useful hosts may include bacterial hosts such as strains of E. coli, e.g., E. coli HB 101, E. coli X1776, E. coli X2282, E. coli MRC1 and strains of Pseudomonas, Bacillus subtilis, Bacillus stearothermophilus and other bacilli, yeasts and other fungi, animal or plant hosts such as animal (including human) or plant cells in culture or other hosts. Of course, not all host/vector combinations may be equally efficient. The particular selection of host/cloning vehicle combination may be made by those of skill in the art after due consideration of the principles set forth without departing from the scope of this invention.

Furthermore, within each specific cloning vehicle, various sites may be selected for insertion of the double-stranded DNA. These sites are usually designated by the restriction endonuclease which cuts them. For example, in pBR322 the PstI site is located in the gene for β -lactamase, between the nucleotide triplets that code for amino acids 181 and 182 of that protein. This site was employed by Villa-Komaroff et al., supra, in their synthesis of protein displaying rat proinsulin antigenic determinants. One of the two HindIII endonuclease recognition sites is between the triplets coding for

amino acids 101 and 102 and one of the several Tag sites at the triplet coding for amino acid 45 of β -lactamase in pBR322. In similar fashion, the EcoRI site and the PvuII site in this plasmid lie outside of any coding region, the EcoRI site being located between the genes coding for resistance to tetracycline and ampicillin, respectively. This site was employed by Itakura et al. and Goeddel et al. in their recombinant synthetic schemes, supra. These sites are well recognized by those of skill in the art. It is, of course, to be understood that a cloning vehicle useful in this invention need not have a restriction endonuclease site for insertion of the chosen DNA fragment. Instead, the vehicle could be joined to the fragment by alternative means.

The vector or cloning vehicle and in particular the site chosen therein for attachment of a selected DNA fragment to form a recombinant DNA molecule is determined by a variety of factors, e.g., number of sites susceptible to a particular restriction enzyme, size of the protein to be expressed, susceptibility of the desired protein to proteolytic degradation by host cell enzymes, contamination of the protein to be expressed by host cell proteins difficult to remove during purification, expression characteristics, such as the location of start and stop codons relative to the vector sequences, and other factors recognized by those of skill in the art. The choice of a vector and an insertion site for a particular gene is determined by a balance of these factors, not all selections being equally effective for a given case.

Although several methods are known in the art for inserting foreign DNA into a cloning vehicle or vector to form a recombinant DNA molecule, the method preferred for a first construction in accordance with this invention is described in Villa-Komaroff et al., supra, and displayed in Figure 1. This method is characterized by digesting the plasmid (in particular pBR322) with that restriction enzyme specific to the site chosen for the insertion (in particular PstI) and adding dGMP

tails to the termini by terminal transferase. dCMP tails are added to the 5' termini of the cut plasmid to regenerate the PstI site and permit linkage to a cDNA fragment carrying the complementary tails. In similar fashion, the double-stranded cDNA is elongated by the addition of dCMP tails to the 3' termini to allow joining to the tailed plasmid. The tailed plasmid and cDNA are then annealed to insert the cDNA in the appropriate site of the plasmid and to circularize the hybrid DNA, the complementary character of the tails permitting their cohesion (Figure 1). The resulting recombinant DNA molecule now carries a gene at the chosen restriction site (Figure 1).

Of course, other known methods of inserting DNA sequences into cloning vehicles to form recombinant DNA molecules are equally useful in this invention. These include, for example, direct ligation, synthetic linkers, exonuclease and polymerase-linked repair reactions followed by ligation, or extension of the DNA strand with DNA polymerase and an appropriate single stranded template followed by ligation.

It should, of course, be understood that the nucleotide sequences or cDNA fragment inserted at the selected site of the cloning vehicle may include nucleotides which are not part of the actual structural gene for the desired polypeptide or may include only a fragment of the complete structural gene for the desired protein. It is only required that whatever DNA sequence is inserted, a transformed host will produce a polypeptide having a biological or immunological activity of HuIFN- α or that the DNA sequence itself is of use as a hybridization probe to select clones which contain DNA sequences useful in the production of polypeptides having an immunological or biological activity of HuIFN- α .

The cloning vehicle or vector containing the foreign gene is employed to transform a host so as to permit that host to express the protein or portion thereof for which the hybrid DNA codes. The selection of an appropriate host is also controlled by a number of factors

recognized by the art. These include, for example, compatibility with the chosen vector, toxicity of proteins encoded by the hybrid plasmid, ease of recovery of the desired protein, expression characteristics, biosafety and costs. A balance of these factors must be struck with the understanding that not all hosts may be equally effective for expression of a particular recombinant DNA molecule.

In the present synthesis, the preferred initial cloning vehicle is the bacterial plasmid pBR322 and the preferred initial restriction endonuclease site therein is the PstI site (Figure 1). The plasmid is a small (molecular weight approx. 2.6 megadaltons) plasmid carrying resistance genes to the antibiotics ampicillin (Amp) and tetracycline (Tet). The plasmid has been fully characterized (F. Bolivar et al., "Construction And Characterization Of New Cloning Vehicles II. A Multi-Purpose Cloning System", Gene, pp. 95-113 (1977); J. G. Sutcliffe, "pBR322 Restriction Map Derived From The DNA Sequence: Accurate DNA Size Markers Up To 4361 Nucleotide Pairs Long", Nucleic Acids Research, 5, pp. 2721-28 (1978)). Insertion of the DNA product in this site provides a large number of bacterial clones each of which contains one of the DNA genes or fragments thereof present in the DNA product previously prepared. Again, only a very few of these clones will contain the gene for IFN- α or fragments thereof (Figure 1). The preferred host for initial cloning in accordance with this invention is E. coli HB 101. Other experiments were conducted with E. coli X1776, a host described in British patent 1,516,458 and placed on deposit with the American Type Culture Collection, Rockville, Maryland, USA, where it has been assigned ATCC No. 31244.

1. Preparation of PstI-Cleaved, dGMP-elongated pBR322

Plasmid pBR322 (20 μ g) was digested with 21 units PstI endonuclease (MRE Porton Downs or New England

Biolabs) in 150 μ l 10 mM Tris-HCl (pH 7.5), 6 mM $MgCl_2$, 50 mM NaCl, 6 mM 2-mercaptoethanol, 200 mg/ μ l bovine serum albumin ("BSA") (Calbiochem). After 2 h at 37°C, the mixture was extracted with 1 vol phenol-chloroform (1:1) and 1 vol ether and precipitated with ethanol.

Addition of homopolymeric dGMP tails (Figure 1) by terminal deoxynucleotidyl transferase (TdT) (purified according to F. J. Bollum, "Deoxynucleotide Polymerizing Enzymes From Calf Thymus Gland", in Methods in Enzymology, (L. Grossman and K. Moldave, eds.), Academic Press, New York, 128, pp. 591-611 (1968)) was done in a 328- μ l reaction volume containing 100 mM sodium cacodylate (pH 7.2), 10 mM NaH_2PO_4 , 5 mM $MgCl_2$, 1 mM dGTP, 50 μ g/ μ l BSA, and 3-6 units of TdT (purified as above) per μ g of DNA. Incubation was at 37°C for 20 min. EDTA was added to 10 mM and the mixture extracted as above and dialyzed for 2 days against TNE buffer.

2. Preparation of dCMP-elongated DNA

Double-stranded DNA was elongated with dCMP residues by standard procedures (E.g., Villa-Komaroff et al., supra). 150 ng of the double-stranded cDNA described above were incubated in 8 μ l 100 mM sodium cacodylate (pH 7.2), 2.5 mM $CoCl_2$, 50 μ g/ μ l BSA, 0.1 mM dCTP containing 3-6 units of purified TdT per μ g of DNA for 8 min at 27°C and then frozen at -20°C. As before, the dCMP-elongated DNA is a mixture of different species, only a very few of which are IFN-related (Figure 1).

3. Preparation of Ca^{++} -Treated E. coli X1776

A single colony of E. coli X1776 was inoculated into 100 ml tryptone medium (C. Weissmann and W. Boll, "Reduction Of Possible Hazards In The Preparation Of Recombinant Plasmid DNA", Nature, 261, pp. 428-29 (1976), supplemented with 100 μ g/ml diaminopimelic acid (Koch-Light Laboratories), 10 μ g/ml nalidixic acid (Calbiochem)

and 10 ug/ml tetracycline (Achromycin[®], American Cyanamid). The culture was grown at 37°C to an apparent optical density of 0.6 at 650 nm (OD₆₅₀) (as measured in a Beckman DB spectrophotometer) and chilled in ice for 30 min. The culture was then sedimented at 4000 rpm in a Sorvall H4 swinging bucket rotor, the cells washed with 50 ml 10 mM NaCl, repelleted by centrifugation, and resuspended in 20 ml 100 mM CaCl₂. The suspension was cooled in ice for 30 min, pelleted by centrifugation and resuspended in 4 ml of 100 mM CaCl₂ and kept on ice overnight for use. E coli HB101 was prepared for transformation by the method of M. Mandel and A. Higa, "Calcium-Dependent Bacteriophage DNA Infection", J. Mol. Biol., 53, pp. 159-62 (1970). Aliquots (0.5 ml) were kept frozen at -70°C and retained their activity for at least 3 months.

4. Annealing of dGMP-elongated pBR322 and dCMP-elongated DNA

The annealing of the tailed, PstI-cleaved pBR322 and tailed cDNA was as described in J. Van den Berg et al., "Comparison Of Cloned Rabbit And Mouse β -globin Genes Showing Strong Evolutionary Divergence Of Two Homologous Pairs Of Introns", Nature, 276, pp. 37-44 (1978). 8 ng of dCMP-elongated DNA product were mixed with 22 ng of dGMP-elongated PstI-cleaved pBR322 in 50 μ l TNE buffer. Incubation was for 4 successive 1 h stages at 65°C, 46°C, 37°C and 20°C. 20 μ l 100 mM Tris-HCl (pH 7.5), 100 mM CaCl₂, 100 mM MgCl₂ and 50 μ l TNE buffer were added and the mixture cooled in ice for 20 min.

The product is, of course, a large mixture of different recombinant DNA molecules and some cloning vehicles without inserted DNA sequences. However, each recombinant DNA molecule contains a cDNA segment at the PstI site. Each such cDNA segment may comprise a gene or a fragment thereof. Only a very few of the cDNA segments code for IFN or a portion thereof (Figure 1). The vast

majority code for one of the other proteins or portions thereof whose mRNA's were part of the poly(A) RNA used in the process of this invention (Figure 1).

5. Transfection Of E. coli Xl776
With The Annealed Hybrid Plasmids

The transfection of E. coli Xl776 with the mixture of recombinant DNA molecules was as described in J. Van den Berg et al., supra. P3 containment facilities were used for the transfection process and all subsequent steps in which the resulting transformed bacteria were handled. The annealed pBR322 recombinant DNA molecules were added to 100 μ l of Ca^{++} -treated E. coli Xl776, prepared previously, and the mixture cooled in ice for 20 min, heated at 20°C for 10 min, and 0.6 ml tryptone medium added. The mixture was plated onto 2 tryptone medium agar plates supplemented as above. Transfection efficiency was 3.3×10^4 colonies per μ g of annealed pBR322 transfecting DNA; native pBR322 gave 3×10^6 colonies per μ g.

Since plasmid pBR322 includes the gene for tetracycline resistance, E. coli hosts which have been transformed with a plasmid having that gene intact will grow in cultures containing that antibiotic to the exclusion of those bacteria not so transformed. Therefore, growth in tetracycline-containing culture permits selection of hosts transformed with a recombinant DNA molecule or recycled vector.

After 48 h at 37°C, individual colonies were picked and suspended in 100 μ l tryptone medium (supplemented as above) in the wells of microtiter plates (Dynatech). After incubation at 37°C overnight, 100 μ l 40% glycerol were mixed into each well. The plates were stored at -20°C and a library of 100,000 individual clones of transformed E. coli Xl776 was prepared.

These 100,000 clones contain a variety of recombinant DNA molecules representing complete or partial

copies of the mixture of mRNAs in the poly(A) RNA preparation from IFN-producing leukocytes (Figure 2). The majority of these will contain only a single recombinant DNA molecule. Only a very few of these recombinant DNA molecules are related to IFN. Accordingly, the clones must be screened to separate the IFN-related clones from the others.

SCREENING FOR A CLONE CONTAINING HuIFN- α CDNA

There are several approaches to screen for bacterial clones containing human leukocyte interferon cDNA ("HuIFN- α CDNA"). These include, for example, RNA selection hybridization (Alwine *et al.*, *infra*), differential hybridization (T. P. St. John and R. W. Davis, "Isolation Of Galactose-Inducible DNA Sequences From *Saccharomyces Cerevisiae* By Differential Plaque Filter Hybridization", *Cell*, 16, pp. 443-452 (1979); Hoeijmakers *et al.*, *infra*), hybridization with a synthetic probe (B. Noyes *et al.*, "Detection And Partial Sequence Analysis Of Gastrin mRNA By Using An Oligodeoxynucleotide Probe", *Proc. Natl. Acad. Sci. USA*, 76, pp. 1770-1774 (1979)) or screening for clones that produce the desired protein by immunological (L. Villa-Komaroff *et al.*, *supra*) or biological (A.C.Y. Chang *et al.*, *supra*) assays. We have chosen RNA selection hybridization as being the most convenient and promising method for primary screening of clones containing IFN- α CDNA.

A. RNA Selection Hybridization Assay

1. Overview Of The Initial Assay

Referring now to Figure 2, recombinant DNA was isolated from a culture of a mixture of 512 clones from the above library of clones (two mixtures of 2 clones shown in Figure 2) (Step A). The reason for selecting this batch size will be explained below. The recombinant

DNA molecules were cleaved, denatured and hybridized to leukocyte poly(A) RNA containing IFN- α mRNA prepared as before (Step B). All recombinant DNA molecule-poly(A) RNA hybrids were separated from the non-hybridized poly(A) RNA (Step C). The poly(A) RNA was recovered from the hybrids and purified (Step D). The recovered RNA was assayed for IFN- α mRNA activity as above (Step E). If, and only if, the mixture of recombinant DNA molecules contains a recombinant DNA molecule having an inserted nucleotide sequence capable of hybridizing to the IFNmRNA in the poly(A) RNA under stringent hybridization conditions, will the mRNA released from that hybrid cause the formation of IFN- α in oocytes, because mRNA released from any other recombinant DNA molecule-poly(A) RNA hybrid will not be IFN- α -related. If a group of 512 clones gave a positive response, the clones were regrouped in 8 lots of 64, and each lot assayed as before. This process was continued until a single clone responding to this assay was identified.

There is no assurance that the recombinant DNA molecules and bacterial clone transformed therewith, which are thus identified, contain the complete IFN- α cDNA sequence of IFN- α or even that the DNA sequence actually codes for IFN- α . However, the recombinant DNA molecules will certainly contain extensive nucleotide sequences complementary to the IFN- α mRNA coding sequence. Therefore, the recombinant DNA molecule may at least be used as a source of a probe to screen rapidly other recombinant DNA molecules and clones transformed with them to identify further sets of clones which may contain an authentic and complete IFN- α nucleotide coding sequence.

2. Theoretical Considerations

The conditions for the hybridization (Step B) are critical. The absolute concentrations and the ratio of recombinant DNA molecule and poly(A) RNA must be chosen so as to take into consideration reaction rate and

stoichiometry. The proper choice is difficult to make, because the proportion of IFN- α mRNA in the poly(A) RNA is not known. In order to assure controlled and adequate kinetics, the hybridization was carried out under conditions where the concentration of DNA sequences from the recombinant DNA molecules was in excess as compared to the estimated IFN- α mRNA concentration. In a mixture of 512 possible different recombinant DNA molecules, an IFN- α -related DNA sequence ("IFN- α R DNA") will either not occur (giving a negative assay), or it will constitute at least about 1/512 of the recombinant DNA molecules. The concentration of the recombinant DNA molecule mixture and therefore the concentration of the IFN- α R DNA, if any, can thus be adjusted in the hybridization step to ensure adequate hybridization rates. In addition, the amount of the IFN- α R DNA in the reaction mixture must be sufficient to bind enough IFN- α mRNA from the poly(A) RNA to allow detection of IFN- α after injection into oocytes of the mRNA recovered from the recombinant DNA molecule-poly(A) RNA hybrid.

In order to detect IFN- α by the assays available, its concentration should be 100 IU/ml or higher. Because 0.5 ml aliquots are required for replicate determinations, 50 IU should be generated in the oocytes. The poly(A) RNA from induced leukocytes, used previously, generates about 500 IU IFN- α upon injection of 1 μ g into oocytes. Therefore, at least 0.1 μ g poly(A) RNA has to be injected to generate the needed 50 IU. Model experiments with rabbit globin mRNA and rabbit β -globin cDNA clones showed that the overall recovery of 125 I-globin mRNA in the oocyte relative to 125 I-globin mRNA added to the hybridization mix was about 10%, and the recovery of mRNA activity about 5%. Therefore, at least $0.1/0.05 = 2$ μ g of leukocyte poly(A) RNA should be used for the hybridization assay. To ensure an adequate safety margin, 12 μ g of poly(A) RNA were used per assay.

To calculate how much DNA from the recombinant DNA molecules is required to bind the IFN- α mRNA in 12 μ g

of poly(A) RNA, the IFN- α mRNA content of poly(A) RNA was estimated. One μ g of poly(A) RNA generates 500 IU of IF. The specific activity of IFN- α lies between 2×10^8 and 10^9 IU/mg protein. 500 IU of IFN- α therefore correspond to between $500/2 \times 10^8 = 2.5 \times 10^{-6}$ mg (2.5 ng) and $500/10^9 = 5 \times 10^{-7}$ mg (0.5 ng) of interferon.

The relationship between the amount of IFN- α mRNA injected into an oocyte and the amount of IFN- α produced is unknown. In the case of β -globin mRNA, about 30 molecules of protein per molecule mRNA are produced per hour; this value is about 6 for β -globin (J.B. Gurdon et al., "Message Stability In Injected Frog Oocytes: Long Life Of Mammalian And β -Globin Messages", J. Mol. Biol., 80, pp. 539-51 (1973)). Assuming an average value of 20 for IFN- α , a molecular weight of 18000 for IFN- α and a molecular weight of 330'000 for IFN- α mRNA, then 26 mg ($18000/330000 \times 20 \times 24$) of IFN- α should be produced in 24 h per mg of IFN- α mRNA injected. If the specific activity of IFN- α is 2×10^8 /mg (2×10^2 IU/ng), then 1 ng IFN- α mRNA will yield $26 \times 2 \times 10^2 = 5.2 \times 10^3$ IU of IFN- α . If the specific activity is 10^9 /mg (10^3 IU/ng), the amount of IFN- α produced would be 2.6×10^4 IU. Because 1 μ g of leukocyte poly(A) RNA yields 500 IU of IFN- α , under the above assumed conditions, the concentration of IFN- α mRNA in 1 μ g poly(A) RNA would fall between 0.1 ng to 0.02 ng and the proportion of IFN- α mRNA in leukocyte poly(A) RNA would lie between 1:10'000 and 1:50'000. Therefore, 12 μ g of poly(A) RNA contains about 1.2 ng to 0.2 ng IFN- α mRNA.

Should the translation ratio of the IFN- α mRNA in the oocytes be lower by an order of magnitude than the average for globin mRNA, the IFN- α mRNA content of the poly(A) RNA would be 10 times higher than calculated above, or between about 1:1000 to 1:5000. And, 12 μ g of poly(A) RNA would then contain about 12 ng to 2 ng of IFN- α mRNA. On the other hand, should the translation ratio of the IFN- α mRNA in the oocytes be higher by an order of magnitude than the average for globin mRNA, the

IFN- α mRNA content of the poly(A) RNA would be 10 times lower than calculated above, or between about 1:100'000 and 1:500'000. And, 12 μ g of poly(A) RNA would then contain 0.1 ng to 0.02 ng IFN- α mRNA.

Plasmid pBR322 has 4361 b.p. The complete cDNA of IFN- α mRNA would add about 800-1000 b.p. to pBR322 on formation of pBR322-IFN- α cDNA to a total of about 5200-5400 b.p. Its molecular weight would thus be about 12 times ($2 \times 5200/800$) that of the IFN- α mRNA alone. Therefore, in order to bind the IFN- α mRNA calculated above to be present in 12 μ g poly(A) RNA required for the assay, an amount of recombinant DNA molecules equal to 12 times the amount of the IFN- α mRNA will be required (stoichiometric amount).

Because the IFN- α mRNA content of the poly(A) RNA used to prepare the recombinant DNA molecules had been increased 10 to 40-fold over that of the crude poly(A) RNA, the group of 512 clones should have 10 to 40 times more clones containing the desired IFN- α mRNA than calculated from the above.

If IFN- α mRNA is 1 part in 1000 of the crude poly(A) RNA, then 12 μ g of poly(A) RNA contain 12 ng IFN- α mRNA and the stoichiometric amount of IFN- α cDNA plasmid is 144 ng. Since a group of 512 clones will contain at least 5 with IFN- α cDNA inserts, the amount of total hybrid plasmid DNA required is 14.8 μ g ($144 \times 512/5 \times 10^{-3}$). If IFN- α mRNA is 1 part in 10'000, then 12 μ g of poly(A) RNA contain 1.2 ng IFN- α mRNA and the amount of IFN- α cDNA plasmid required is 14.4 ng. A group of 512 clones will contain either 0 or 1 IFN- α cDNA insert, so that the amount of total hybrid plasmid DNA required is 7.4 μ g ($14.4 \times 512 \times 10^{-3}$). If IFN- α mRNA is 1 part in 100'000, then the amount of total hybrid plasmid DNA required is 0.74 μ g ($1.44 \times 512 \times 10^{-3}$). In order to ensure that the hybridization reaction will proceed under DNA excess conditions (i.e., excess recombinant DNA as compared to poly(A) RNA), 20 μ g of the mixture (about 1.4 to 30-fold excess) was chosen for the assay.

Hybridization must be conducted under conditions which ensure (a) that the hybridized portion of the poly(A) RNA is recovered intact and in a biologically active form, (b) that non-specific DNA-mRNA association is prevented, and (c) that the hybridization reaction goes to at least 75% completion. These conditions are most likely to be met by hybridization in 80% formamide, 0.4M NaCl (J. Casey and N. Davidson, "Rates Of Formation And Thermal Stability Of RNA:DNA And DNA:DNA Duplexes At High Concentrations Of Formamide", Nucleic Acids Res., 4, pp. 1539-52 (1977)). In this solution, hybridization can be conducted at about 40°C (rather than the 60°-70°C required when formamide is omitted). Lower temperatures are preferred to minimize damage to the poly(A) RNA. We chose a hybridization temperature of 56°C. This is about 3° below the $T_{1/2i}$ (J. Casey and N. Davidson, supra) and about 10-13° below $T_{1/2d}$ (Hamaguchi & Geidushek, J. Amer. Chem. Soc., 84, p. 1329). Therefore, this temperature should not allow hybridization of sequences with less than about 87% homology, since a 1% mismatch lowers the $T_{1/2d}$ by 1° (T. F. Bonner et al., "Reduction In The Rate Of DNA Reassociation By Sequence Divergence", J. Mol. Biol., 81, pp. 123-35 (1973)).

In the present hybridization, self-hybridization of DNA is not a major problem because the mixture of DNA's being used consists of the same vector (pBR322) and a variety of cDNA inserts. Therefore, most of the DNA sequences will be heteroduplexes in which the inserts are available for hybridization to poly(A) RNA. It is very unlikely that complementary cDNA inserts which form part of different duplexes will interact because of topological constraints. In any event, DNA:DNA reassociation is minimized under the reaction conditions used (J. Casey and N. Davidson, supra).

To determine the hybridization time required to ensure at least 75% reaction, a second order rate equation was employed:

$$t = \frac{\ln \frac{Co - Ro + Ro (1 - \frac{R}{Ro})}{(1 - \frac{R}{Ro}) Co}}{k_R (Co - Ro)} = 3.9 \text{ h}$$

where:

R = molar nucleotide concentration of hybridized RNA

Co = molar nucleotide concentration of initial DNA to be hybridized

Ro = molar nucleotide concentration of initial RNA to be hybridized

k_R = rate constant for RNA-DNA hybridization

t = time (sec)

and:

$$\frac{R}{Ro} = 0.75 \text{ (75\% reaction completion)}$$

$$k_R = 472 \text{ (} k_R = 1/12 k_d \text{ (J. Casey and N. Davidson, supra))}$$

where: k_d = second order rate constant for DNA under the chosen conditions of hybridization

and: $k_d = 1.7 \times 10^5 \times L^{1/2} \times N^{-1}$
(J. R. Hutton and J. G. Wetmur, "Renaturation Of Bacteriophage ϕ X174 DNA-RNA Hybrid: RNA Length Effect And Nucleation Rate Constant", J. Mol. Biol., 77, pp. 495-500 (1973))

L = 900 (chain length in b.p.; about 900 are present in the full IFN- α cDNA insert)

N = 900 (complexity in b.p.
of the hybrid chain;
here the complexity
is 900 because the 900
nucleotides of the
IFN- α mRNA join with the
complementary 900
nucleotides of the
IFN- α cDNA insert)

$$C_0 = 2.5 \times 10^{-7}$$

(Based on a 40 μ l solution
containing the previously
determined 20 μ g of recombi-
nant DNA molecules to be
used in the assay, again
assuming that the IFN- α cDNA
insert will be 1/12 of a
recombinant DNA molecule
and will occur in at least
1 of the 512 clones, and
assigning 662 as the average
molecular weight of one DNA
base pair)

$$R_0 = 8.7 \times 10^{-8}$$

(Based on a 40 μ l solution
containing the previously
determined 12 μ g of poly(A)
RNA to be used in the
assay, again assuming that
the poly(A) RNA contains
1:10'000 parts IFN- α mRNA
(given the large excess of
DNA a different proportion
will have little effect on
the rate of hybridization)
and assigning 343 as the
average molecular weight of
one ribonucleotide of RNA)

3. Execution Of The Initial Assay

Step A-Preparation and Cleavage of the Recombinant DNA Molecule Mixture

The desired number of bacterial clones was inoculated onto tryptone medium agar plates supplemented as above, by transferring to it an aliquot from each microtiter well with use of a mechanical device. After incubation at 37°C, each clone had given rise to a colony of several mm diameter. All colonies were washed off the plate(s) and pooled to give an inoculum used to inoculate 1 l of tryptone medium supplemented as above in a 2 l Erlenmyer flask. The culture was shaken at 37°C to an apparent OD₆₅₀ of about 0.8 (estimated visually). One volume of supplemented tryptone medium and chloramphenicol to 170 µg/ml were added to the culture which was further shaken at 37°C for 16 h. 20 ml chloroform were added and the culture shaken again for 10 min at 37°C to kill the bacteria (C. Weissmann and W. Boll, *supra*). The culture was decanted from the chloroform and the cells were harvested by centrifugation (Sorvall GS3 rotor) for 15 min at 6000 rpm and 4°C. About 1-2 g of cells were obtained for each 1-liter preparation. The cells were suspended in 30 ml 20 mM Tris-HCl (pH 7.5), centrifuged for 20 min at 5000 rpm and 4°C (Sorvall SW rotor) and resuspended in 30 ml 50 mM Tris-HCl (pH 7.5). 0.25 vol of lysozyme solution (10 mg/ml in 50 mM Tris-HCl (pH 7.5)) were added and after cooling for 10 min at 0°C 0.33 vol (based on the vol of the original 50 mM Tris-HCl-culture suspension) 0.5 M EDTA (pH 8.0) were gently mixed in without shaking. After another 10 min at 0°C, 1/16 vol (again based on the original volume) of 2% Triton X-100 were added. After 60 min, the sample was centrifuged for 60 min at 10'000 rpm and 0°C in a Sorvall SW rotor. The supernatant was transferred to a beaker containing a magnetic stirrer, and 3M NaOH was added with stirring until a pH of 12.5 was reached, as measured at 20°C, using a glass electrode and an Orion Research model

601 pH meter, standardized with Beckman pH 10 Carbonate Buffer Standard (No. 3505). After stirring 10 min at 20°C, the pH was adjusted to 8.5. After 3 min further stirring 1/9 vol 5 M NaCl and 1 vol phenol (distilled and equilibrated with 9.5 M NaCl) were added and vigorous stirring continued for 5 min. The phases were separated by centrifugation (GSA Sorvall rotor) at 10'000 rpm and 0°C for 10 min. The supernatant containing Form I DNA (circular double-stranded DNA) was carefully removed from the interphase (which contains single-stranded DNA) and extracted 3 times with chloroform. (Phenol must be largely removed at this step). The Form I DNA fraction will contain those recombinant DNA molecules (pBR322-cDNA insert) originally used in transforming those host cells which form part of the 512 clones chosen for assay.

Pancreatic RNAase A (5 mg/ml, preheated 10 min at 85°C) was added to the Form I DNA to a concentration of 20 µg/ml and the mixture incubated 60 min at 37°C. 1/5 vol 5 M NaCl were added and the mixture adjusted with 30% polyethylene glycol 6000 (Union Carbide, autoclaved 20 min at 120°C) up to a final concentration of 7.5% PEG. After 2-16 h at -10°C, the precipitate was collected in a Sorvall SW Rotor for 20 min at 8'000 rpm and 0°C, dissolved in 0.075 M NaCl, 0.0075 M Na-citrate to an absorbance of 20 at 260 nm, and adjusted to 0.5% SDS. The solution was incubated for 30 min at 37°C with 0.5 mg/ml Pronase (self-digested at 20 mg/ml, 2h at 37°C) and extracted 3 times with 1 vol distilled phenol and 2 times with 1 vol chloroform. The sample (up to 2 ml of a 1 mg/ml DNA solution) was centrifuged through a 5 to 23% sucrose gradient in 50 mM Tris-HCl (pH 7.5), 1 mM EDTA for 15 h at 21'000 rpm and 15°C using an SW 27 Beckman Rotor. Fractions were collected and the OD₂₆₀ monitored. DNA-containing fractions were pooled and the DNA precipitated with sodium acetate and ethanol. 20 to 100 µg of the Form I DNA mixture were recovered by centrifugation.

Twenty µg of purified Form I DNA were digested in 150 µl 10 mM Tris-HCl (pH 7.5), 6 mM MgCl₂, 50 mM

NaCl, 6 mM 2-mercaptoethanol, 200 µg/ml BSA or gelatin and 20 units HindIII (New England Biolabs). The HindIII restriction enzyme cleaves the Form I DNA at a site within the pBR322 moiety (It is unlikely that the cDNA moiety is also cleaved, but if it is, the assay should not be substantially affected). After 2 h at 37°C, an aliquot (1%) was analyzed by electrophoresis through a 1% agarose gel in 50 mM Tris-acetate (pH 7.8), 2 mM EDTA for 1 h at 50 mA to ascertain whether digestion was complete. If digestion was not complete, more HindIII was added and incubation continued for 2 h. When the Form I DNA was converted totally to linear molecules, Pronase (Calbiochem), EDTA and SDS were added to 0.5 mg/ml, 10 mM and 0.5% respectively. After 30 min at 37°C, the solution was extracted with 30 µl phenol-chloroform (1:1). The organic phase was washed with 50 µl 20 mM Tris-HCl (pH 7.5), 1 mM EDTA, and the combined aqueous phases extracted 3 times with ether, filtered through a 0.1-ml Chelex column, collected in an EDTA-boiled Pyrex[®] tube and precipitated with 1/10 vol 3M sodium acetate and 2.5 vol ethanol. After standing overnight at -20°C, the DNA was collected by centrifugation.

Step B - Hybridization of the DNA with Poly(A) RNA

Two hybridization mixtures were prepared. Mixture I contained 4 µl of 10-fold concentrated hybridization buffer (4M NaCl, 0.1 PIPES (pH 6.4, 1,4 piperazine-diethane sulfonic acid, Sigma), 50 mM EDTA, 0.5 µl (about 5 ng ¹²⁵I-globin mRNA (5000 cpm) and 6 µl induced leukocyte poly(A) RNA (2 µg/µl), an amount sufficient to generate 6000 IU of IFN when injected into oocytes. Mixture II contained 10 µg of the HindIII digested Form I DNA from above and 0.1 µg of PstI-digested Z-pBR322(H3)/RcβG-4.13 (a pBR322 derivative that contains the β-globin sequence in the HindIII site) (Mantei et al., "Rabbit β-globin mRNA Production In Mouse L Cells Transformed With Cloned Rabbit β-globin Chromosomal DNA", Nature,

281, pp. 40-46 (1979)). The ^{125}I -globin mRNA in mixture I and the β -globin DNA in mixture II serve as internal positive controls for the hybridization assay. Both mixtures were dried in a stream of nitrogen gas. 40 μl of 80% formamide were added to the residue of mixture II and the solution was denatured for 10 min at 100°C and chilled quickly in ice. The denatured solution was used to dissolve the residue of mixture I and the resulting solution incubated at 56°C for 4 h.

Step C - Separation Of Hybridized Poly(A)
RNA-DNA From Non-Hybridized Poly(A)
RNA

After dilution to 1 ml with cold 0.9 M NaCl, 0.09 M Na-citrate and formamide (100%) to 4% (by volume) the solution was filtered at 0.5 ml/min through a Millipore filter (0.45 μm pore size), the filter having been first tested for its capacity to retain RNA-DNA hybrids, because not all filters obtained from the manufacturer were equally efficient.

Step D - Purification Of Hybridized Poly(A)
RNA

The above filter, with poly(A) RNA hybrids attached, was immersed in 1 ml 0.15 M NaCl, 0.015 M Na-citrate, 0.5% SDS for 10 min at 37°C, rinsed with 50 mM Tris-HCl (pH 7.5), 10 mM MgCl_2 , 2 mM CaCl_2 and placed in 0.6 ml of fresh buffer. After the addition of 5 μl iodoacetate-treated DNAase (5mg/ml) (S. B. Zimmermann and G. Sandeen, Anal. Biochem., 14, p. 269 (1966); P. A. Price et al., "Alkylation Of A Histidine Residue At The Active Site Of Bovine Pancreatic Deoxyribonuclease", J. Biol. Chem., 244, pp. 924-32 (1969)), the filter was incubated at 37°C for 10 min.

The filter was removed and the solution extracted with 1 vol phenol and 1 vol ether and passed through a 0.1-ml Chelex column. 5 μg of carrier RNA (purified

yeast RNA) were added to the solution and the RNA precipitated with sodium acetate and ethanol. The precipitate was collected by centrifugation at 10'000 xg, dissolved in 100 μ l 1 mM EDTA, heated for 90 sec at 100°C, and TNE and SDS added to 2 x TNE and 0.5% SDS. The RNA was adsorbed to a 100- μ l oligo(dT) cellulose column, eluted with four washes of 0.3 ml distilled water and precipitated with sodium acetate and ethanol. After 16 h at -20°C the precipitated RNA was separated by centrifugation and dissolved in 2 μ l TNK buffer.

Step E - Determination Of IFN- α RNA Activity

The poly(A) RNA solution from above was injected into 40 oocytes (about 50 nl per oocyte). The oocytes were incubated at 23°C for 24-48 hours, homogenized and centrifuged (or the incubation medium recovered) and assayed as described previously for IFN- α .

4. Subsequent Assay - Hybridization To Filter-Bound DNA

Most subsequent assays of a recombinant DNA molecule from a single clone were carried out with DBM or DPT paper-bound DNA, because the assay conditions were no longer critical and the assay is more convenient. DPT paper gave lower backgrounds and was used preferentially. DBM paper was prepared as described (J. C. Alwine *et al.*, "Method For Detection Of Specific RNAs In Agarose Gels By Transfer To Diazobenzyl Oxymethyl-Paper And Hybridization With DNA Probes", Proc. Natl. Acad. Sci. USA, 14, pp. 5350-54 (1977)). APT paper was prepared by a procedure of B Seed (pers. commun.): Sheets of Whatman 540 paper (20 g) were agitated for 16 h at 20°C with a mixture of 70 ml 0.5 M NaOH, 2 mg/ml NaBH₄ and 30 ml 1,4-butanediol diglycidyl ether. The paper was then transferred to a solution of 10 ml 2-aminothiophenol in 40 ml acetone and agitated for 10 h. The paper was exhaustively washed

with acetone, 0.1 N HCl, H₂O, 0.1 N HCl, H₂O and dried. APT paper was diazotized to DPT paper as described for the conversion of ABM to DBM paper (Alwine et al., supra).

DNA (up to 15 µg) was bound to 50 mm² diazotized ABM (DBM) or diazotized APT (DPT) paper as described by J.H.J. Hoeijmakers et al. "The Isolation Of Plasmids Containing DNA Complementary To Messenger RNA For Variant Surface Glycoproteins Of Trypanosoma Brucei", Gene, in press, 1980) and set forth below.

Hybrid plasmid DNA was digested with endonuclease PstI, treated with 500 µg Pronase per ml, 0.5% SDS, and 10 mM EDTA for 30 min at 37°C, extracted with phenol and ether, passed through a 0.1-ml Chelex column, and precipitated with ethanol. The heat-denatured DNA (up to 5 µg, with a small amount of ³²P-DNA added as tracer) was incubated overnight at 0°C with 1 cm² DBM or DPT paper in 200 µl 25 mM potassium phosphate buffer (pH 6.5). Filters were washed three times for 5 min at room temperature with 50 mM potassium phosphate buffer (pH 6.5), 1% glycine and three times with 99% recrystallized formamide. A further incubation with 99% formamide for 2 min at 68°C was followed by three washes in 50 mM potassium phosphate buffer (pH 6.5) at 20°C and two washes in 0.4 M NaOH at 37°C for 10 min. About 40-60% of the radioactivity was retained on the filters. The filters were incubated for 3 h at 38°C in pre-hybridization medium A, supplemented with 1% glycine, using 330 µl per filter. Medium A contains 50% formamide, 5 x SSC, 0.04% polyvinyl pyrrolidone, 0.04% Ficoll (Pharmacia), 0.1% SDS, 25 µg poly(A) (P & L) and 100 µg yeast RNA (BDH, extracted six times with phenol and precipitated with ethanol). The filters were washed twice in medium A and then hybridized for 16 h at 38°C with poly(A) RNA as indicated (usually 5-8 µg) in medium A under paraffin oil. The RNA was added as follows: one wet DNA filter was blotted and put in a sterile Petri dish, 20-40 µl of the RNA solution were pipetted on this filter and a second DNA filter (either a duplicate or a control) was put on top and the

sandwich was covered with a sterile paraffin oil. After the hybridization the filters were successively washed in medium A (2 times), in a solution containing 1 x SSC, 0.2% SDS, 1 mM EDTA (3 times, 10 min at 20°C each), medium A (2 h at 38°C) and in 50% formamide, 5 x SSC, 0.1% SDS (3 times, 10 min at 20°C). Hybridized RNA was eluted by heating for 1 min at 100°C in 200 µl 10 mM Tris-HCl (pH 7.4), 1 mM EDTA and 0.1% SDS. The elution step was repeated twice, the eluates were combined and the RNA was precipitated with ethanol after addition of 2 µg yeast RNA (purified as above). The washed pellet was vacuum dried, dissolved in 3 µl H₂O and injected into oocytes. IFN-α activity was assayed as above.

5. Results Of The RNA Selection Hybridization Assay

The assays from 8 groups of 512 clones (i.e., groups T, Y, j, K, . , O, ε and π were negative. The assays from 4 groups of 512 clones (i.e., groups I, δ, N and λ) were positive, albeit not consistently. The positive assays are reported in the following format: IU/ml of IFN-α produced by the RNA released from poly(A) RNA-DNA hybrid (assay from control hybridization using Z-pBR322(H3)/Rcβ G-4.13, supra); the assays in which the experimental results were higher than the background control are underscored.

<u>Group</u>	<u>IU/ml</u>
I	<60 (<60); <u>110</u> (<20); <110 (<110); <110 (<110); <35 (<35)
δ	<u>20</u> (<20)
N	<u>35</u> (<20); <110 (<110); <u>200</u> (<110)
λ	<60 (<60); <u>60</u> (<20); <110 (<110); <110 (<110)

Group λ was subdivided into 8 subgroups of 64 clones and hybridized and assayed as before. The subgroups gave the following results, presented in the same format as above:

<u>Subgroup</u>	<u>IU/ml</u>
λ - I	<35 (<35); <35 (<35)
λ - II	<u>130</u> (<30); <45 (<45)
λ - III	<u>225</u> (<35); <u>35</u> (<30); <u>35</u> (<30); <u>600</u> (<30); <20 (<20)
λ - IV	<u>85</u> (<35); <25 (<25)
λ - V	<35 (<35)
λ - VI	<35 (<35)
λ - VII	<35 (<35)
λ - VIII	<35 (<35)

Subgroup λ -III was subdivided into 8 sets of 8 clones, and hybridized and assayed:

<u>Set</u>	<u>IU/ml</u>
λ -III-1	<20 (<20); <20 (60); <u>35</u> (<30)
λ -III-2	<35 (<35); <30 (<30); <u>150</u> (<20); <u>600</u> (<35); <u>110</u> (60)
λ -III-3	<25 (<25); <30 (<30)
λ -III-4	<u>30</u> (<30); <20 (<20); <20 (60)
λ -III-5	30(?) (<35); <20 (<20); <35 (60)
λ -III-6	<30 (<30); <20 (<20)
λ -III-7	<30 (<20)
λ -III-8	<30 (<20)

Because the first positive result was achieved with the set λ -III-4, the individual colonies of this set (designated A to H) were hybridized and assayed:

λ -III-4-B	<35* (<35); <20 (60)
λ -III-4-C	35 (60); <u>60</u> * (<35); <u>111</u> * (<11); 11* (<11); <u>20</u> (<20)

Therefore, clone λ -III-4-C contains a recombinant DNA molecule capable of hybridizing IFN- α mRNA.

* The DBM paper method was used in this assay.

The recombinant DNA molecule in this clone is designated: Z-pBR322(Pst)/HcIF-4C ("Hif-4C"), and the bacterial strain containing it: E. coli X1776 (Z-pBR322 (Pst)/HcIF-4C) ("E. coli Hif-4C"). This nomenclature indicates that the recombinant DNA molecule originated in Zurich (Z) and is plasmid pBR322 containing at the PstI site a HIFN- α cDNA ("HcIF"); the particular recombinant DNA molecule being derived from clone λ -III-4-C ("4C").

RECLONING AND CHARACTERIZATION OF Z-pBR322(Pst)/HcIF-4C

Since primary clones of transformed cells occasionally contain more than one species of recombinant DNA molecule (Efstratiadis et al., "The Primary Structure Of Rabbit β -globin mRNA As Determined From Cloned DNA", Cell, 10, pp. 571-85 (1977)), Hif-4C was isolated from E. coli X1776 (Hif-4C) clones and purified as described above. Samples of Hif-4C and pBR322 were digested with PstI and analyzed by electrophoresis on a 1% agarose gel. Hif-4C gave two bands, one with the mobility of Pst-cleaved pBR322, the other with a mobility corresponding to about 320 b.p.

E. coli HB101 was transformed with the isolated Hif-4C as described above. Six clones of tetracycline-resistant, transformed bacteria were picked, small cultures prepared and Form I DNA purified and analyzed by PstI cleavage and agarose gel electrophoresis as before. All samples showed cleavage patterns identical to Hif-4C. One of these recloned recombinant DNA molecules was designated Z-pBR322(Pst)/HcIF-4c ("Hif-4c") and used for further experimentation. The lower case "c" designates a recloned DNA molecule.

To determine the capacity of Hif-4c and its cDNA insert to hybridize to IFN- α mRNA, Hif-4c (115 μ g) was digested to completion with 125 units of PstI, extracted with phenol and chloroform, and precipitated with ethanol as described above. An aliquot (10 μ g) was 5' terminally labeled (to serve as a tracer in subsequent steps) by

dissolving it in 100 μ l 50 mM Tris-HCl (pH 7.5), passing it through a 0.1-ml Chelex 100 column and treating it with 0.6 units bacterial alkaline phosphatase for 1 h at 65°C. Tenfold concentrated TNE (40 μ l) was added and the solution extracted 3 times with 1 vol phenol and 3 times with 1 vol chloroform. The DNA was precipitated with 2 vol ethanol at -20°C overnight and collected by centrifugation. For further purification, the sample in 0.5 ml TNA was adsorbed to 0.25-ml DEAE cellulose (Whatman DE52, prewashed with 2 ml 150 mM NaCl, 50 mM Tris-HCl (pH 7.5), 2 mM EDTA) ("NET-buffer"), washed with 2 ml of NET buffer, eluted with 0.4 ml 1.5 M NaCl, 20 mM Tris-HCl (pH 7.5), 2 mM EDTA and precipitated with ethanol as above. The DNA was incubated with γ - 32 P-ATP (specific activity about 5000 Ci/mmol) and polynucleotide kinase, (A.M. Maxam and W. Gilbert, "A New Method For Sequencing DNA", Proc. Natl. Acad. Sci. USA, 74, pp. 560-564 (1977)) and purified by chromatography on a 3-ml Sephadex-G50 column in TNE. The eluted fractions were pooled and the 32 P-DNA precipitated with ethanol as above; yield, about 10^7 dpm.

The unlabeled PstI-cleaved Hif-4c DNA (90 μ g) was mixed with 6×10^5 dpm of 32 P-labeled PstI cleaved Hif-4c DNA from above and electrophoresed through a 10 x 20 x 0.7 cm, 2% horizontal agarose gel in 50 mM Tris-acetate buffer (pH 7.8) using a 2.5 cm slot. An x-ray film was exposed to the gel and the position of the 320-bp fragment determined. The gel strip containing the radioactive band (1.3×10^5 dpm) was cut out, crushed by pressing through a plastic 2-ml syringe and extracted overnight at 4°C by agitation with ten times the gel vol of NET buffer. The DNA was adsorbed to a 0.1-ml hydroxyapatite column (prewashed with 1 ml NET buffer). The column was washed with 1 ml 0.1 M K-phosphate buffer (pH 7.5) and the DNA eluted with 0.2 ml 1 M K-phosphate buffer (pH 7.5). The eluate was diluted 10-fold with sterile distilled H₂O and the DNA adsorbed to and eluted from DEAE and precipitated with ethanol as described above. This DNA is called "Hif-4c fragment".

The Hif-4c fragment (120 ng) was bound to DPT paper (0.5 x 0.5 cm) as described above. As a control, 120 ng β -globin cDNA fragment excised with HindIII from the hybrid plasmid Z-pBR322(H3)Rc β G-4.13 (F. Meyer et al., "Transposition Of AT-linked, Cloned DNA From One Vector To Another", Experimentia, 35, p. 972 (1979); N. Mantei et al., supra) and processed similarly. Hybridization of duplicate filters to poly(A) RNA (in 20 μ l), washing of the filters and recovery of the RNA from the filters were as described above. After injection into oocytes the following IFN- α activities were detected:

<u>DNA fragment</u>	<u>amount of leukocyte poly(A) RNA* (μg)</u>	<u>time of hybridization</u>	<u>IFN-α activity (IU/ml)** (duplicate assay)</u>
Hif-4c	2.5	16 h	250;100
β -globin cDNA	2.5	16 h	4;1
Hif-4c	7.5	16 h	3000;1000
β -globin cDNA	7.5	16 h	4;30
Hif-4c	7.5	5 h	1000;1000
β -globin cDNA	7.5	5 h	10;1

Thus, Hif-4c contains an insert capable of hybridizing to IFN- α mRNA.

IDENTIFICATION OF CLONES OF E. COLI CONTAINING
RECOMBINANT DNA MOLECULES CROSS-HYBRIDIZING TO
THE INSERT IN Hif-4c

Since the cDNA insert in recombinant DNA molecule Hif-4c was only about 320 b.p., or a third of the estimated size of IFN- α mRNA, the purified Hif-4c fragment

* 1 μ g of this RNA gave 4600 IU/ml.

** Oocyte supernatant after 48 h incubation, assayed by cytopathic effect reduction (W.E. Stewart, II and S.E. Sulkin, supra).

described above was used as a probe to screen for bacterial clones containing recombinant DNA molecules having related hybrid DNA inserts (Figure 3).

The 64 bacterial clones constituting subgroup λ -III described above were stamped onto a Millipore membrane (8 cm diameter), placed on an agar plate (supplemented with diaminopimelic acid, nalidixic acid and tetracycline, as above) and incubated for 24 h at 37°C. The filter was placed onto a 0.75 ml drop of 0.5 M NaOH and after 2-3 min transferred onto a paper towel to remove excess liquid; the step was repeated. The filter was neutralized, using 1 M Tris-HCl (pH 7.5), and washed with 1.5 M NaCl - 0.5 M Tris-HCl (pH 7.4) in a similar fashion as above and air dried. The filter was dipped in 0.3 M NaCl, air dried and heated at 80°C for 2 h in a vacuum.

Hif-4c Pst fragment (30 ng) was ^{32}P -labeled by nick translation (A.J. Jeffreys and R.A. Flavell, "The Rabbit β -Globin Gene Contains A Large Insert In The Coding Sequence", Cell, 12, pp. 1097-1108 (1977)) using β - ^{32}P dATP and α - ^{32}P dCTP (specific activity, 40 Ci/mmol each). The filter bearing the λ -III colonies was prehybridized in 4 x SET (SET is 0.15 M NaCl, 30 mM Tris-HCl (pH 8.0), 1 mM EDTA), 0.1% (w/v) Ficoll, 0.1% polyvinylpyrrolidone, 0.1% (w/v) BSA, 0.5% SDS, and 200 $\mu\text{g/ml}$ denatured, fragmented salmon sperm DNA for 7 h at 68°C and hybridized with 2×10^5 cpm of ^{32}P -labeled Hif-4c fragment in 4 x SET, 0.02% (w/v) Ficoll, 0.02% polyvinylpyrrolidone, 0.02% w/v BSA, 0.5% SDS and 200 $\mu\text{g/ml}$ denatured salmon sperm DNA at 68°C for 16 h. The filter was rinsed with SET-0.5% SDS at room temperature, washed with 2 x SET - 0.5% SDS for 5 h at 68°C, replacing the solution once, and with 3 mM Trizma base at room temperature for 4 h, replacing the solution once. After drying the filter, an x-ray film was exposed to the filter for 80 h using a screen. Three colonies gave a strong positive response, namely λ -III-7D, λ -III-2H and λ -III-4C, and 2 colonies a weak one, namely λ -III-1E, λ -III-3D.

Small cultures were prepared from the Hif-4c related clones, Form I DNA was purified, cleaved with PstI and analyzed by agarose gel electrophoresis as described above. All Form I DNAs gave rise to a large fragment (plasmid pBR322 moiety) and a small one (hybrid insert). The recombinant DNA molecule from λ -III-2H released the largest insert, namely about 900 b.p. This recombinant DNA molecule was designated Z-pBR322(Pst)/HcIF-2H ("Hif-2H") and its insert "Hif-2H fragment".

Hif-2H was tested for its capacity to bind IFN- α mRNA by binding it to DPT paper (4 μ g/100mm²) and hybridizing it to poly(A) RNA (0.3 μ g/ μ l), all as described above, for 16 h and determining IFN- α mRNA activity:

<u>DNA sample</u>	<u>IFN-α activity (IU/ml)*</u>
Hif-2H	250 \pm 50 (average of 4 determinations)
Z-pBR322(H3)/Rc β G-4.13	30 (average of 2 determinations)
pBR322	20

Hif-2H was recloned as described for Hif-4C and designated Hif-2h.

In a further experiment an additional set of E. coli clones containing recombinant DNA molecules was prepared and colonies hybridizing to the labeled Hif-4c fragment were identified. In order to ensure a high yield of plasmids with long cDNA inserts, part of the double-stranded ³²P-labeled leukocyte cDNA prepared enzymatically from leukocyte poly(A) RNA (the same cDNA preparation as described above) was fractionated by size by centrifuging through a sucrose density gradient, using the same procedure described for the centrifugation of the poly(A) RNA. The fractions containing the cDNA with a sedimentation velocity corresponding to a 600 b.p. DNA

* Assayed by cytopathic effect reduction.

fragment or greater were pooled and the cDNA recovered after ethanol precipitation. The cDNA was elongated with dCMP residues, hybridized to dGMP-elongated Pst I-cleaved pBR322 and the hybrid DNA used to transform E. coli as before, except that E. coli HB101 was used. The bacteria were distributed onto 8-cm diameter Millipore filters, placed on Tryptone medium agar plates (containing 10 µg/ml tetracycline) and grown until small colonies appeared. A replica filter was prepared by pressing a fresh, moist Millipore filter onto the colony-bearing filter, peeling it off, placing it face upward on an agar plate containing 4.4% glycerol and incubating it until small colonies appeared. This colony-bearing filter was covered with a further Millipore filter, frozen at -55°C and stored (D. Hanahan and M. Meselson, "A Protocol For High Density Plasmid Screening", Sept. 1978, personal communication). Eighteen filters, bearing a total of about 5000 colonies were prepared. One replica of each filter was used for hybridization to the ³²P-labeled, Pst I-excised Hif-4c DNA fragment, exactly as described above. About 185 positive colonies were identified on an autoradiogram, recloned on Millipore filters and identified once more by hybridization. 95 clones giving the strongest hybridization response were designated Z-pBR322(Pst)/HcIF-SN1 to SN95 and used for further investigation.

It is, of course, evident, given the ability of Hif-2h to produce a polypeptide displaying an immunological or biological activity of HuIFN (infra), that Hif-2h and other DNA sequences related to it, e.g. Hif-4c, may be employed in this method of clone screening equally well on other clones containing DNA sequences arising from recombinant DNA technology, synthesis, natural sources or a combination thereof or clones containing DNA sequences related to any of the above DNA sequences by mutation, including single or multiple, base substitutions, insertions, inversions, or deletions to select other DNA sequences and clones which also code for HuIFN. Therefore, such DNA sequences and their identification also fall within

this invention (e.g., infra). It is also to be understood that DNA sequences, which are not screened by the above DNA sequences, yet which as a result of their arrangement of nucleotides code on expression for the polypeptides coded for by the expression of the above DNA sequences also fall within this invention.

FURTHER CHARACTERIZATION OF Hif-2h DNA INSERT

As described above recombinant DNA molecule Hif-2h contains an insert of about 900 b.p., and hybridizes to human leukocyte interferon mRNA. The following additional characteristics were determined.

1. Hybrid Arrested Translation

If mRNA is hybridized to a cloned, complementary cDNA, the translation of the mRNA is inhibited, however heat denaturation of the hybrid releases translatable mRNA (B.M. Paterson et al., "Structural Gene Identification And Mapping By DNA-mRNA Hybrid-Arrested Cell-Free Translation", Proc. Natl. Acad. Sci. USA, 74, pp. 4370-74 (1977)). 2.2 µg Pst I-cleaved Hif-2h, and as a control 2 µg HindIII-cleaved Z-pBR322(H3)/RcβG-4.13 ("RcβG") were denatured in 10 µl 80% (vol/vol) deionized formamide - 20 mM PIPES buffer (pH 6.4) for 10 min at 80°C. The solution was added to an Eppendorf tube into which leukocyte poly(A) RNA (5 µg), NaCl (4 µ moles) and EDTA (10 nmoles) had been dried down. The mixture was heated for 7 h at 48°C under a layer of paraffin oil, cooled and diluted with 200 µl H₂O. The two samples were divided into equal parts, and one of each was heated at 100°C for 30 sec. The nucleic acids were precipitated with ethanol, dissolve in 3 µl H₂O and assayed for IFN-αmRNA activity in oocytes as above:

	Le poly(A)			
<u>DNA</u>	<u>RNA input</u>		<u>Treatment</u>	<u>IFN-α (IU/ml)*</u>
Hif-2h (1.1 μ g)	2.5 μ g		hybridized	400
Hif-2h (1.1 μ g)	2.5 μ g		hybridized and	
			denatured	2000
Rc β G (1 μ g)	2.5 μ g		hybridized	3000
Rc β G (1 μ g)	2.5 μ g		hybridized and	
			denatured	3000
Hif-2h (0.5 μ g)	1 μ g		none	2000
-	1 μ g		none	3000
-	1 μ g		none	2000

Therefore, Hif-2h, when hybridized with poly(A) RNA, inhibited the translation of the IFN- α mRNA in the poly(A) RNA; after denaturing the hybrid, the IFN- α mRNA was again translatable. This experiment confirms that Hif-2h contains sequences complementary to IFN- α mRNA.

2. Analysis By Restriction Enzyme Cleavage And Determination Of Nucleotide/Amino Acid Sequences And Restriction Map

Digestions of Hif-2h with various restriction enzymes (New England Biolab) were carried out, and the resulting products analyzed by agarose gel electrophoresis. The underlined fragments are not common to pBR322 and Hif-2h:

<u>Restriction enzyme</u>	<u>Fragment sizes</u>	
	<u>Hif-2h**</u>	<u>pBR322***</u>
<u>Pst</u> I	<u>885</u> + 20,4361	4361
<u>Eco</u> RI	<u>1426</u> ,3820	<u>4361</u>
<u>Bgl</u> II	<u>5246</u>	not cleaved

* The oocyte medium was assayed after 48 h by the cytopathic effect inhibition method.

** Although internally consistent herein the absolute size of these fragments is best indicated by reference to Figs 8-10.

*** From Sutcliffe (supra).

<u>EcoRI</u> + <u>BglII</u>	336,4960	4361
<u>EcoRI</u> + <u>PstI</u>	209,676,748	748,3611
	3611	
<u>BspI</u>	921,587,540,504,	587,540,
		504,457,
	457,434,2 x 231,	434,267,
		234
	+14 fragments	+14 fragments
	200 bp	200 bp
<u>MboII</u>	1616,884	not done
	and others	

In addition, 5' terminally ^{32}P -labeled PstI cleaved Hif-2h was cleaved with several restriction enzymes and the sizes of the radioactive fragments derived from the cDNA insert in that recombinant DNA molecule were determined:

<u>Restriction enzyme</u>	<u>^{32}P-fragments*</u>
<u>EcoRI</u>	676,209
<u>HindIII</u>	no cleavage
<u>BspI</u>	799, 86
<u>HpaII</u>	no cleavage
<u>HhaI</u>	no cleavage
<u>BamHI</u>	no cleavage
<u>Hinf</u>	210, 62
<u>BglII</u>	545,340

From these data a restriction map of Hif-2h was deduced (Figure 4). The map is not definitive in regard to the absolute location of the sites; and maybe incomplete in regard to MboII sites within the insert. Only the sites closest to the insert are given within the pBR322 moiety. The arrow indicates the orientation of the IFN- α cDNA coding strand.

Although the actual structure of the Hif-2h fragment or other inserts in clones of this invention or the amino acid sequence or structure of the polypeptides coded therefrom is not required for one of skill in the art to make and use the invention described and claimed herein, the above data and restriction map were included

* Although internally consistent herein the absolute size of these fragments is best indicated by reference to Figures 8-10.

in the original application hereto as the best available information on the structure of fragment at the time of filing the original application. Since that time, as expected (supra, p. 9, lines 27-29), these data and restriction map for the Hif-2h fragment have been refined using well-known techniques of nucleotide sequencing and restriction analysis. E.g., A. M. Maxam and W. Gilbert, "A New Method For Sequencing DNA", Proc. Natl. Acad. Sci. USA, 74, pp. 560-64 (1977). Plasmid DNA was prepared by Method B (N. M. Wilkie, et al., "Hybrid Plasmids Containing An Active Thymidine Kinase Gene Of Herpes Simplex Virus 1", Nucleic Acids Research, 7, pp. 859-77 (1979)) and restricted by various restriction enzymes essentially as recommended by the supplier, except that 200 µg/ml gelatin replaced the bovine serum albumin in the enzyme buffers. (EcoRI was a gift from W. Boll, BspI a gift from A. Kiss and other enzymes were obtained from New England Biolabs.)

Restricted DNA (20 µg) was extracted with phenol, precipitated with ethanol, dissolved in 0.05 M Tris-HCl (pH 8), and passed over a small column of Chelex-100. Fragments with flush or 5'-overhanging ends were dephosphorylated by treatment with 0.2 units calf intestinal alkaline phosphatase (Boehringer) per pmol DNA 5' ends in 200 µl 0.05 M Tris-HCl (pH 8) for 60 min at 37°C. The enzyme was inactivated by heating 60 min at 65°C. For DNA fragments with 3' overhanging ends, bacterial alkaline phosphatase (Worthington) was used as described (A. M. Maxam and W. Gilbert, supra) except that incubation was at 65°C for 30 min. The dephosphorylated DNA was purified by adsorption to and elution from DEAE-cellulose (W. Muller et al., "Site-Directed Mutagenesis In DNA: Generation Of Point Mutations In Cloned B-Globin Complementary DNA At The Positions Corresponding To Amino Acids 121-123", J. Mol. Biol., 124, pp. 343-58 (1978)) or subjected to polyacrylamide gel electrophoresis where required (see below). Fragments recovered from a polyacrylamide (or agarose) gel in 0.15 M NaCl, 0.05 M

Tris-HCl (pH 8) were adsorbed to a 0.1-ml hydroxyapatite (Biorad HTP) column, washed 4 times with 1 ml 0.1 M potassium phosphate buffer (pH 7) and eluted with 0.3 ml 1 M potassium phosphate buffer (pH 7). The solution was diluted tenfold and the DNA adsorbed to DEAE cellulose and recovered as described (W. Muller *et al.*, *supra*).

After ethanol precipitation, the DNA was 5'-terminally labeled with [γ - 32 P] ATP (12-34 μ Ci per pmol DNA 5' ends) and polynucleotide kinase (New England Biolabs or P-L Biochemicals Inc.) essentially as described by A. M. Maxam and W. Gilbert, *supra*, except that the DNA was not denatured before the kinase reaction. Specific activities of 1-1.5 μ Ci [32 P] phosphate per pmol DNA 5' ends were obtained.

For sequencing, labeled fragments were cleaved with a second restriction enzyme and the products separated by electrophoresis through a 5% polyacrylamide gel in tris-borate-EDTA buffer. The desired fragments were extracted from the gel and purified (Muller *et al.*, *supra*). The various fragments for sequencing were prepared as follows (the number indicates the nominal fragment chain length in base pairs, the labeled site is indicated by an asterisk):

(1) cleavage of Hif-2h with BspI, isolation of BspI-BspI-232 and BspI-BspI-949 by 5% polyacrylamide gel electrophoresis in Loening's buffer (U. E. Loening "The Fraction Of High Molecular Weight Ribonucleic Acid By Polyacrylamide Gel Electrophoreses", *J. Biochem.* p. 102 (1967));

(2) cleavage of Hif-2h with BspI, labeling, cleavage with PstI, isolation of BspI*-PstI-83 and BspI*-PstI-827;

(3) cleavage of Hif-2h with BglII, labeling, cleavage with PstI, isolation of BglII*-PstI-336 and BglII*-PstI-570;

(4) cleavage of Hif-2h with MboII, labeling, digestion with PstI and HindII (to cleave an interfering

350 bp pBR322 fragment), isolation of MboII*-PstI-519 and MboII*-PstI-351;

(5) cleavage of Hif-2h with EcoRI, labeling, cleavage with PstI, isolation of EcoRI*-PstI-708 and EcoRI*-PstI-198;

(6) cleavage of Hif-2h with PstI, labeling, cleavage with BglII, isolation of PstI*-BglII-570 and PstI*-BglII-336;

(7) cleavage of Hif-2h with AvaII, labeling, cleavage with PstI and BglII, isolation of AvaII*-PstI-186 and AvaII*-BglII-147;

(8) cleavage of Hif-2h with PvuII, labeling, cleavage with PstI and BglII, isolation of PvuII*-PstI-486.

The fragments were degraded according to the method of A. M. Maxam and W. Gilbert, *supra*, with the modifications described in protocols provided by the same authors in September 1978. The products were fractionated on 0.1 x 25 x 36 cm 12% polyacrylamide gels (acrylamide/bisacrylamide = 18/1) in 50 mM tris-borate, 1 mM EDTA (pH 8.3), with runs of 2, 8, 18 and 26 h at 900 V following a 6 h prerun at 700 V. Best results were obtained when the gels were kept at room temperature 2-3 days before use.

Each stretch of the cDNA insert was sequenced from both strands and each restriction site which served as labeled terminus was sequenced using a fragment spanning it. The nucleotide sequence thus obtained is depicted in Figures 8-10. As is to be expected the positions of the various restriction sites in this insert are more absolutely located than those determined by restriction enzyme cleavage alone and depicted in Fig. 4.

Referring now to Figs. 8-10, the heteropolymeric part of the insert is flanked by 23G residues at the 5' end and by 7A residues (probably reflecting the poly(A) terminus of the mRNA) followed by 15C residues at the 3' terminus. For reference, the insert is numbered from the first nucleotide following the dG tail to the last nucleotide before the polyA residues. An ATG initiation

triplet in position 57-59 and a TAA termination triplet at position 624-626 define a reading frame uninterrupted by nonsense codons. Both other reading frames in this region of the insert contain 18 and 12 nonsense codons respectively. Moreover, the only other sequences, *i.e.*, in different reading frames, flanked by an ATG or GTG and a termination signal, which code for a polypeptide of 25 amino acids or more, are located between nucleotides 226 and 304, 640 and 778 and 683 and 743, respectively. Therefore, the region between nucleotides 57 and 626 most likely includes the nucleotide sequence of the Hif-2h fragment that codes for a polypeptide displaying a biological or immunological activity of IFN- α in accordance with this invention.

It should of course be understood that cloned cDNA from polyA RNA by the usual procedures (A. Efstratiadis *et al.*, *supra*) lacks 5' terminal nucleotides and may even contain artifactual sequences (R. I. Richards *et al.*, "Molecular Cloning And Sequence Analysis Of Adult Chicken B-Globin cDNA", *Nucleic Acids Research*, 7, pp. 1137-46 (1979)). Therefore, it is not certain that the ATG located at nucleotides 57-59 is in fact the first ATG of authentic mRNA. However, for the purposes of the following description, it is assumed that the ATG at nucleotides 57-59 is the first ATG of authentic mRNA.

By comparing the polypeptide coded by this region of the insert with that sequence of 35 amino terminal amino acids of authentic human lymphoblastoid interferon --SerAspLeuProGlnThrHisSerLeuGlyAsnArgArgAla-LeuIleLeuLeuAlaGlnMetGlyArgIleSerLeuPheSerCysLeuLys-AspArgHisAsp-- determined by K. C. Zoon *et al.*, *supra* and M. Hunkapiller and L. Hood, *supra*, it appears that the chosen reading frame is correct and that nucleotides 57-124 may code for a signal sequence which precedes the nucleotide sequence coding for the "mature" polypeptide because alignment of the published sequence with the determined sequence (from the 24th amino acid onward) displays extensive coincidence (*i.e.*, 26 of 35 amino acids).

In eukaryotic mRNAs the first AUG triplet from the 5' terminus is usually the initiation site for protein synthesis (M. Kozak, "How Do Eukaryotic Ribosomes Select Initiation Regions In Messenger RNA", *Cell*, 15, pp. 1109-25 (1978)). The codon in the Hif-2h fragment corresponding to the first amino acid of lymphoblastoid interferon is 22 codons from the first AUG (and 14 codons from the second one) indicating that the sequence coding for interferon may be preceded by a sequence determining a signal peptide of 23 (or less likely 15) amino acids. The longer of the presumptive signal sequences contains an uninterrupted series of 11 hydrophobic amino acids (and the shorter one, one of 6 hydrophobic amino acids). This accumulation of hydrophobic residues is characteristic of signal sequences (cf., B. D. Davis and P. C. Tai, "The Mechanism Of Protein Secretion Across Membranes", *Nature*, 283, pp. 433-38 (1980))

The nucleotide sequence apparently corresponding to "mature" IFN- α polypeptide comprises 498 nucleotides, which code for 166 amino acids. Assuming that there is no carboxyterminal processing, the molecular weight of the interferon polypeptide is 19'388. The base composition of the coding sequence is 50% GC. The codon usage within the interferon coding sequence is in reasonable agreement with that compiled for mammalian mRNAs in general (R. Grantham, et al., "Codon Catalog Usage And The Genome Hypothesis", *Nucleic Acids Research*, 8, pp. 49-62 (1980)). Any deviations observed may be ascribed to the small numbers involved.'

The structure of the polypeptide depicted in Figure 8-10 for the Hif-2h fragment, of course, does not take into account any modifications to the polypeptide caused by its interaction with in vivo enzymes, e.g., glycosylation. Therefore, it must be understood that this structure may not be identical with IFN- α produced in vivo, but it still has very similar, if not identical, biological and immunological properties. Neither, does

this structure exclude the likelihood that other modifications such as mutations, including single or multiple, base substitutions, deletions, insertions, or inversions or chemical derivatizations of this structure will not produce compounds that also display IFN- α activity.

3. Determination Of The Plus Strand Of The Inserted IFN- α CDNA

The DNA strand that has the same sequence as the mRNA is designated as plus strand, and its complement as minus strand. The plus strand of the IFN- α CDNA insert was identified as outlined in Figure 5. Hif-2h DNA was cleaved with the restriction enzyme BglII, the termini labeled with ^{32}P -phosphate (as described above for PstI-cleaved termini) and the DNA digested with PstI, to give longer (545 b.p. (570 b.p. as determined in the more refined analysis reported above)) and shorter 340 bp (336 bp as determined in the more refined analysis reported above)) radioactive fragments. These fragments were denatured and hybridized to poly(A) RNA from induced leukocytes in 80% formamide, 0.4 M NaCl, i.e., under conditions where DNA-DNA reassociation does not occur (supra). The nucleic acids were digested with nuclease S1, which degrades all single-stranded nucleic acids, in particular the non-hybridized ^{32}P -DNA, and the products were separated on a polyacrylamide gel (R. F. Weaver and C. Weissmann, "Mapping Of RNA By A Modification Of The Berk-Sharp Procedure", Nucleic Acid Research, 7, pp. 1175-93 (1979)). An autoradiogram showed that only the shorter -nucleotide fragment had been hybridized and protected by the poly(A) RNA, identifying the 5'-labeled shorter-nucleotide strand as the minus strand. The orientation of the plus strand is therefore as given in Figure 4 and Figure 5 (right hand side).

4. Demonstration That Poly(A) RNA From
Non-Induced Human Leukocytes Does
Not Hybridize To Hif-2h DNA

An experiment identical to that described in the preceding section was carried out, however the poly(A) RNA was from non-induced human leukocytes, prepared by the same procedure as in the case of Sendai virus-induced leukocytes. No detectable amount of labeled DNA was protected. By comparison to the results of the preceding section the poly(A) RNA from non-induced cells contains less than about 1/20 the amount of mRNA hybridizable to Hif-2h than does poly(A) RNA from induced cells.

SYNTHESIS OF A POLYPEPTIDE WITH INTERFERON ACTIVITY
BY E. COLI CONTAINING RECOMBINANT DNA MOLECULES
RELATED TO Z-pBR322(Pst)/HcIF-4c

The PstI site of pBR322 lies within the β -lactamase (penicillinase) gene. Therefore, when a coding DNA segment (e.g., a cDNA comprising all or part of a gene) is ligated into the position in the proper orientation and proper reading frame, a fused protein may result. The protein would consist of the amino-terminal portion of β -lactamase followed by the amino acid sequence for which the inserted DNA sequence codes (L. Villa-Komaroff et al., supra). If the inserted DNA segment comprises a DNA sequence containing its own initiation signal, and has a sequence preceding it with a termination signal in phase with the β -lactamase sequence, termination and re-initiation may occur at the second initiation signal and a non-fused, active protein may result (A.C.Y. Chang et al., supra). To ensure that the DNA insert related to Hif-4c was inserted in the proper reading frame for expression within the β -lactamase gene, a set of derivatives of pBR322, namely pKT279, pKT280 and pKT287 (constructed by K. Talmadge, personal communication, 1979) was employed. Each of these derivatives has a PstI site located such that a DNA insert ligated into that site will be in a different reading frame from an insert at

the PstI site of the other derivatives of the set (Figure 6). Therefore, the set permits the insertion of DNA into the β -lactamase gene in all three reading frames. The PstI-excised insert from Hif-2h was prepared as described for the fragment Hif-4c. The Hif-2h Pst fragment (10 ng) was mixed with PstI-cleaved pBR322, pKT279, pKT280 or pKT287 (10 ng in each case) in 20 μ l of 10 mM Tris-HCl (pH 7.5), 6 mM MgCl₂, 100 mM NaCl, 6 mM β -mercaptoethanol, 200 μ g/ml gelatin and 0.1 mM ATP and incubated with 0.1 units T₄ DNA ligase (New England Biolabs) for 16 h at 10°C. The resulting recombinant DNA molecules are designated Z-pBR322(Pst)/HcIF-2h, Z-pKT279(Pst)/HcIF-2h, Z-pKT280(Pst)/HcIF-2h and Z-pKT287(Pst)/HcIF-2h. E. coli HB101 was transformed with each of these recombinant DNA molecules and transformed colonies were selected on tetracycline-containing agar plates as described previously. Since tetracycline-resistant clones of transformed bacteria may also contain the recyclized vector, bacterial colonies of each set were grown on Millipore filters and colonies hybridizing to ³²P-labeled Hif-4c fragment were identified and selected as described above. These strains were designated as follows,

E. coli HB101 (Z-pBR322(Pst)/HcIF-2h-AH1) to (-AH3);

E. coli HB101 (Z-pKT279(Pst)/HcIF-2h-AH1) to (-AH8);

E. coli HB101 (Z-pKT280(Pst)/HcIF-2h-AH1) to (-AH8);

E. coli HB101 (Z-pKT287(Pst)/HcIF-2h-AH1) to (-AH8).

Extracts of some of the above strains as well as of some of the strains Z-pBR322(Pst)/HcIF-SN1 to 95 were tested for IFN- α activity. Bacteria were grown in Tryptone medium to stationary phase, harvested, washed with 1/20 vol (based on the vol of the culture) 50 mM Tris-HCl (pH 8), 30 mM NaCl and frozen. After thawing, the cells were resuspended in the volume indicated below of the previous buffer and lysozyme was added to 1 mg/ml. After 60 min at 0°C the suspensions were frozen (in an ethanol-dry ice bath) and thawed (at 37°C) 5 times, and centrifuged 10 min at 12'000 rpm in a GSA Sorvall rotor. In some cases part of the supernatant (S30) was further

centrifuged at 100'000xg in a Type 65 Spinco rotor and the supernatants (S100) recovered. Such supernatants were screened for IFN- α activity by the cytopathic effect reduction assay (Expt.1). The colonies showing a positive response in Expt. 1 were reassayed as well as 49 clones from the set Z-pBR322/HcIF-SN-1 to SN-95 described above at a lower dilution (Expt. 2).

Source of extract:

E. coli HB 101 transformed by:

Expt. 1*

	<u>Preparation</u>	<u>IFN-α activity (IU/ml)</u>
Z-pBR322(Pst)/HcIF-2h	S30	?
Z-pBR322(Pst)/HcIF-2h-AH-1 to 3	S30	?
Z-pKT279(Pst)/HcIF-2h-AH-1 to 7	S30	?
Z-pKT279(Pst)/HcIF-2h-AH-8	S30	positive
Z-pKT280(Pst)/HcIF-2h-AH-2, 6, 7	S30	?
Z-pKT280(Pst)/HcIF-2h-AH-1, 3, 4, 5	S30	positive
Z-pKT287(Pst)/HcIF-2h-AH-1, 2, 3, 4, 5, 8	S30	?
Z-pKT287(Pst)/HcIF-2h-AH-6, 7	S30	positive

Source of extract:

E. coli HB101 transformed by:

Expt. 2**

	<u>Preparation</u>	<u>IFN-α activity (IU/ml)</u>
Z-pKT279/HcIF-2h-AH-8	S30 and S100	300
Z-pKT280/HcIF-2h-AH-1, 3, 4, 5	S30 and S100	300
Z-pKT287/HcIF-2h-AH-6 and 7	S30 and S100	300
Z-pBR322(Pst)/HcIF-SN-4, 5, 7, 9, 10, 11, 13, to 16	S30	neg (<10)
Z-pBR322(Pst)/HcIF-SN-18 to 22, 24, 25, 27, 30 to 34	S30	neg (<10)
Z-pBR322(Pst)/HcIF-SN-38 to 41, 43 to 48	S30	neg (<10)

* Expt. 1: Extracts assayed at 1:150 final dilution

** Expt. 2: Extracts assayed at 1:6 final dilution

Z-pBR322(Pst)/HcIF-SN-1 to 3,6, 8,12,17, 23,26	S30	10
Z-pBR322(Pst)/HcIF-SN-28,29,36, 37,49	S30	10
Z-pBR322(Pst)/HcIF-SN-35,42	S30	200

Some of the more active producers from above were examined in more detail. Cultures were grown to late log phase (apparent OD₆₅₀ about 0.9)* and the cells lysed as above, in 1/50 of the culture volume. The following activities were found, using Z-pBR322(Pst)/HcIF-SN32 as a negative control:

<u>Source of extract:</u>		<u>IFN-α activity**</u>
<u>E. coli HB101 transformed by:</u>	<u>Preparation</u>	<u>(IU/ml)</u> <u>(dup. assays)</u>
Z-pKT279(Pst)/HcIF-2h-AH8	S30, S100	100; 300
Z-pKT280(Pst)/HcIF-2h-AH3	S30, S100	1000; 1000
Z-pKT287(Pst)/HcIF-2h-AH6	S30, S100	200; 200
Z-pBR322(Pst)/HcIF-SN35	S30, S100	1000; 1000
Z-pBR322(Pst)/HcIF-SN42	S30, S100	300; 100
Z-pBR322(Pst)/HcIF-SN32	S30, S100	0; 0

It is to be understood that the actual protein produced by these strains has not been analyzed structurally to determine whether or not it is produced fused to amino acids unrelated to IFN or with all or part of IFN's signal sequence. However, whatever protein is produced, it displays an immunological or biological activity of IFN. Therefore, the protein as expressed is useful. Most importantly, the activity of the protein demonstrates

* Three thousand liters or larger cultures of IFN- α can be grown without loss of IFN activity.

** It is to be understood that the above reported expression may reflect interferon production by genes under the control of the penicillinase expression control sequence.

that the DNA sequence which codes for it is a DNA sequence related to HuIFN- α . It therefore is within the skill of the art to employ that DNA sequence as is demonstrated herein to select other like HuIFN- α related DNA sequences and to provide the basis for other constructions that will express mature interferon or other variants thereof or will improve the yield of the particular protein expressed.

The level of production of a protein is governed by two major factors: the number of copies of its gene within the cell and the efficiency with which those gene copies are transcribed and translated. Efficiency of transcription and translation (which together comprise expression) is in turn dependent upon nucleotide sequences, normally situated ahead of the desired coding sequence. These nucleotide sequences or expression control sequences define, inter alia, the location at which RNA polymerase interacts to initiate transcription (the promoter sequence) and at which ribosomes bind and interact with the mRNA (the product of transcription) to initiate translation. Not all such expression control sequences function with equal efficiency. It is thus of advantage to separate the specific coding sequences for the desired protein from their adjacent nucleotide sequences and fuse them instead to other known expression control sequences so as to favor higher levels of expression. This having been achieved, the newly engineered DNA fragment may be inserted into a multicopy plasmid or a bacteriophage derivative in order to increase the number of gene copies within the cell and thereby further to improve the yield of expressed protein.

Several expression control sequences may be employed as described above. These include the operator, promoter and ribosome binding and interaction sequences (including sequences such as the Shine-Dalgarno sequences) of the lactose operon of E. coli ("the lac system"), the corresponding sequences of the tryptophan synthetase system of E. coli ("the trp system"), the major operator

and promoter regions of phage λ (O_L^P and O_R^{PI}), the control region of the phage fd coat protein, or other sequences which control the expression of genes of prokaryotic or eukaryotic cells and their viruses. Therefore, to improve the production of a particular polypeptide in an appropriate host, the gene coding for that polypeptide may be prepared as before and removed from a recombinant DNA molecule containing it and reinserted into a recombinant DNA molecule closer to its former expression control sequence or under the control of one of the above expression control sequences. Such methods are known in the art.

Further increases in the cellular yield of the desired products depend upon an increase in the number of genes that can be utilized in the cell. This is achieved, for illustration purposes, by insertion of recombinant DNA molecules engineered in the way described previously into the temperate bacteriophage λ (NM989), most simply by digestion of the plasmid with a restriction enzyme, to give a linear molecule which is then mixed with a restricted phage λ cloning vehicle (e.g., of the type described by N. E. Murray et al., "Lambdoid Phages That Simplify The Recovery Of In Vitro Recombinants", Molec. gen. Genet. 150, pp. 53-61 (1977) and N. E. Murray et al., "Molecular Cloning Of The DNA Ligase Gene From Bacteriophage T4", J. Mol. Biol., 132, pp. 493-505 (1979)) and the recombinant DNA molecule produced by incubation with DNA ligase. The desired recombinant phage is then selected as before and used to lysogenise a host strain of E. coli.

Particularly useful λ cloning vehicles contain a temperature-sensitive mutation in the repression gene cI and suppressible mutations in gene S, the product of which is necessary for lysis of the host cell, and gene E, the product which is the major capsid protein of the virus. With this system the lysogenic cells are grown at 32°C and then heated to 45°C to induce excision of the prophage. Prolonged growth at 37°C leads to high levels

of production of the protein, which is retained within the cells, since these are not lysed by phage gene products in the normal way, and since the phage gene insert is not encapsidated it remains available for further transcription. Artificial lysis of the cells then releases the desired product in high yield.

In an initial attempt to increase the yield of polypeptide, displaying a biological or immunological activity of human leukocyte interferon, produced from hosts transformed with Z-pBR322(Pst)/HcIF-SN35 by the processes above described, a restriction map of the DNA insert in the hybrid was determined. This mapping revealed that as compared to Hif-2h, Hif-SN35 was lacking a PstI site flanking the 3' end of the sequence, part of the signal sequence was missing (up to and including codon 7) and the AvaII site in the signal sequence had been replaced by a BspI site. Therefore, Hif-SN35 is likely a polymorphic or allelic variant of Hif-2h.

The plasmid Hif-SN35 was opened with PstI and the resulting DNA strand chewed back at both ends using standard procedures and the LAC-Alu fragment (*infra*) inserted therein and the plasmid reclosed. The actual structure of the modified plasmid, identified as Z-pBR322(Pst)/HcIF-SN35-AHL6, and the amino acid sequence at the amino terminal end of the protein produced in E. coli have been determined. The nucleotide sequence of this construction reveals that the LAC-Alu fragment was attached one amino acid away from the first amino acid of IFN- α 1(SN35). The amino acid sequence of the amino terminal portion of the protein expressed in E. coli revealed that a fused protein was produced having six amino acids fused to the IFN- α 1(SN35) sequence. However, hosts transformed with the modified plasmid produce about 100 times more polypeptide displaying a biological or immunological activity of human leukocyte interferon as compared to hosts transformed with unmodified Z-pBR322(Pst)/HcIF-SN35.

Referring now to Figure 25, another attempt to improve the yield of polypeptide, displaying an immunologi-

cal or biological activity of human leukocyte interferon, produced by hosts transformed with Z-pBR322(Pst)/HcIF-SN35 (SN35 in Figure 25) is depicted. The hybrid plasmid was cleaved using standard procedures with BspI (a gift of Dr. Kiss). After heat inactivation (65°C, 30 min) of the restriction enzyme, the mixture was adjusted to 50 mM Tris-HCl (pH 8) and heated (37°C, 30 min). Following extraction with phenol and ether, the largest, α lcDNA fragment was isolated on low temperature gelling agarose (0.8%) and HindIII linkers attached. The modified fragment was then joined to HindIII-cleaved plasmid HS-pBR322(Eco)/lacUV5-150 ("LAC-150")* (a gift of H. Schaller) by melting the fragment-containing gel pieces (about 20 μ l each) at 65°C, cooling to 37°C and adding 20 U per μ l T4 DNA ligase. After 16 h at 15°C, ligation occurred in the solidified gel (H. Lehrach, personal communication 1980). One tenth vol 100 mM Tris-HCl (pH 7.5), 100 mM CaCl₂, 100 mM MgCl₂ were added to the sample and it was heated 5 min at 65°C and cooled to 37°C. The samples were then used to transform Ca²⁺ treated E. coli HB101, incubated at 0°C for 20 min, heated at 42°C for 1 min and for 10 min at 20°C. After addition of 1 mol tryptone medium, the samples were incubated 60 min at 37°C and plated on to agar plates containing ampicillin. Plasmid DNA was separated from these cultures, as before, and the hybrid plasmid containing the IFN- α 1 insert with its 5' end adjoining the LAC fragment identified by restriction analysis. The plasmid was then cleaved with EcoRI using conventional procedures and digested with exonuclease BAL-31 (0.06 U/ml, 2-4 min at 30°C) to remove the overhanging EcoRI tail of the LAC fragment and to shorten the β -galactosidase coding segment.

* This plasmid contains the lac promoter HaeIII-202 bp fragment (W. Gilbert *et al.*, "Lactose Operator Sequences And The Action Of Lac Repressor" in Protein Ligand Interactions, H. Sund and G. Blauer, eds. (Berlin, Walter de Gruyter), pp. 193-206 and K. Backman *et al.*, "Maximizing Gene Expression On A Plasmid Using Recombination In Vitro", Cell, 13, pp. 65-71 (1978)) flanked by an EcoRI linker at its 3' end.

To ensure that the treated plasmid contained the complete IFN- α 1 coding sequence, the plasmid was then cleaved with BglII, using conventional procedures, worked up as before and the largest fragment separated on agarose gel (0.8%). This fragment was then combined with a BspI-BglII fragment from Z-pBR322(Pst)/HcIF-SN35 and the resulting hybrid plasmid used to transform E. coli HB101 as before. The transformed colonies were screened for IFN activity and one clone having a high level of IFN activity was selected. This clone was designated E. coli HB101 (C8-IFN- α 1) and its hybrid plasmid C8-IFN- α 1.

DNA sequence analysis of C8-IFN- α 1 revealed that the coding sequence following the initiator triplet determined the first seven amino acids of β -galactosidase, a Pro residue generated by the fusion, amino acids 16 to 23 of the IFN- α 1 signal sequence and the IFN- α 1 (SN35) sequence. E. coli minicell strains (DS410) transformed with hybrid plasmid C8-IFN- α 1 produce about 50 million units IFN per liter or about 2500 times more polypeptide displaying an immunological or biological activity of HuIFN as compared to minicells transformed with unmodified Z-pBR322(Pst)/Hif-SN35. Amino acid sequencing of the polypeptide produced by plasmid C8-IFN- α 1 confirms that the product is a fused protein having seven amino acids from β -galactosidase, one amino acid generated from the fusion and amino acids 16-23 of the IFN- α 1 signal sequence fused to IFN- α 1.

Further, examples of various constructions to improve the protein yields in accordance with this invention are discussed in connection with other forms of IFN- α (infra).

PROPERTIES OF INTERFERON ACTIVITY PRODUCED BY
E. COLI TRANSFORMED WITH HYBRID PLASMIDS

1. Sensitivity Of IFN- α Activity To Trypsin

50 μ l samples of authentic HuIFN- α (specific activity, 1.2×10^6 U/mg; 50U), and the S100 extracts described above of E. coli HB101 (Z-pKT287(Pst)/HcIF-2h-AH6) ("Hif-287-6 extracts") (200 U/ml, 10 U) and of E. coli HB101 (Z-pBR322(Pst)/HcIF-SN35) ("Hif-35 extracts") (1000 U/ml; 50 U) were incubated with various amounts of trypsin, as indicated below, for 30 min at 37°C. Since the S100 extracts have a high protein content, while the HuIFN- α does not, a mixture of HuIFN- α and the control S100 extract Hif-32 was tested in parallel:

<u>IFN-α preparation</u>	<u>Trypsin (μg)</u>	<u>IFN-α activity (units)</u>
HuIFN- α (50 units in 50 μ l Hif-32 S100 extract)	0	50
	0.1	50
	1	50
	10	5
	50	0
	0	15
Hif-287-6 S100 extract (10 units)	0.1	15
	1	5
	10	1
	50	0
	0	15
	0.1	15

Therefore the IFN- α of the extracts is sensitive to trypsin and hence a protein.

Hif-35 S100 extract (50 units)	0	30
	0.1	20
	1	20
	10	2
	50	0
	0	30

2. Behavior On Chromatography On Sephadex G-100

Extract Hif-35 (1 ml) and the S100 extract of E. coli HB101 (Z-pBR322(Pst)/HcIF-SN32) ("Hif-32 extracts") were chromatographed on a 32-ml Sephadex G-100 column at 4°C in 50mM K-phosphate buffer (pH 7.4). Cytochrome c

(0.2 mg) was added as an internal marker. The flow rate was 2 ml/hr and 1.0 ml fractions were collected. The absorbance at 280 nm, and 405 nm (cytochrome c), and the IFN- α activity were determined. As shown in Figure 7, the IFN- α activity of Hif-35 extracts was eluted before cytochrome c, with a k_D value of about 0.45. Therefore, the apparent molecular weight of the substance was between about 20'000 and 30'000*; no activity was detected in the fractions of control extract Hif-32.

3. Inhibition Of The Interferon Activity
Of Hif-35 And Hif-287-6 By Antibody
Against Human Leukocyte Interferon.

HuIFN- α (specific activity 1.2×10^6 IU/mg), and the Hif-35/Hif-287-6 S100 extracts were incubated with various dilutions of sheep antiserum against HuIFN- α (prep. K. Cantell, Feb. 24, 1976, specific activity 450'000 units/ml) in 100 μ l Modified Eagles Medium (MEM) with 10% calf serum for 30 min at 37°C and 45 μ l were assayed for IFN- α activity by the cytopathic effect reduction assay. (The antibody itself did not cause a cytopathic effect):

<u>IFN-α Preparation</u> <u>(units)</u>	<u>Anti-leukocyte-</u> <u>interferon antibody</u> <u>(units)</u>	<u>Residual</u> <u>IFN-α activity</u> <u>(IU)</u>
IFN- α (10)	0	5
	0.18	~0.5
	9	<0.1
	450	<0.1
Hif-35 extract (25)	0	15
	0.18	15
	9	<0.1
	450	<0.1
Hif-287-6 extract (25)	0	15
	0.18	15
	9	<0.1
	450	<0.1
none	0	<0.1
	450	<0.1

* The molecular weight determined by nucleotide sequencing and assuming no carboxyterminal processing is 19,388.

To show that the action of the antibody was not due to an unspecific effect, such as proteolytic degradation, a similar experiment was performed with the mouse interferon system:

<u>IFN Preparation</u> <u>(Units)</u>	<u>Anti-leukocyte-</u> <u>interferon antibody</u> <u>(units/ml)</u>	<u>IFN activity</u> <u>(units/ml)</u> <u>mouse system</u>
mouse preparation	4500	100
(100 units)	90	100
	18	100

Thus, antibodies directed against HuIFN- α specifically inhibit the IFN- α activity of polypeptides produced in E. coli transformed with certain recombinant DNA molecules containing the HcIF-2h DNA sequence. The apparently lower affinity of the antibody for the IFN- α produced in E. coli may reflect structural differences between the latter and natural HuIFN- α , for example, absence of carbohydrate moiety, presence of signal sequence, or fusion to part of the β -lactamase sequence.

4. Reduced Activity Of Hif-35 And Hif-287-6 Extracts On Mouse Cells

Human CCL23 cells or Mouse L929 cells were treated with E. coli extracts, HuIFN- α (prep., K. Cantell, specific activity 1.2×10^6 units/mg) or mouse IF (N.I.H. standard), were challenged with virus (Mengo virus in the case of human cells and VSV in the case of mouse cells) and the IFN- α activity determined by the cytopathic effect reduction assay:

<u>Addition</u>	<u>IFN activity (units/ml)</u>	
	<u>human system</u>	<u>mouse system</u>
mouse - interferon (120 units/ml)	-	120
Hif-35 extracts	100	13
	1000	120
Hif-287-6 extracts	30	4
	300	40
HuIFN- α (100 units/ml)	100	4
HuIFN- α (1000 units/ml)	1000	40

These results show that Hif-35 and Hif-287-6 extracts have a protective action on human cells and only a slight effect (~10%) on mouse cells, as is typical for human interferon.

5. Effect On Some Cell Function

Extracts from E. coli HB101 (Z-pBR322(Pst)/Hif-SN35-AHL6) were compared with authentic IFN for its effect on some cell functions. The E. coli made IFN- α displayed the following properties of natural IFN- α : (1) it enhances natural killing activity of human lymphocytes; (2) it enhances antibody-dependent cell-mediated cytotoxicity; (3) it suppresses antigen- and mitogen-induced leukocyte migration inhibition; and (4) it inhibits growth of IFN-sensitive Burkitt lymphoma cells. These properties are indicia of E. coli synthesized IFN- α 's activity against human tumors and cancers.

6. Activity Of IFN- α Without Amino-Terminal Sequences

IFN- α without amino-terminal sequences has also been made in E. coli and shown to display activity consistent with IFN activity.

To construct the appropriate recombinant DNA molecule, plasmid Hif-2h (supra) was partially digested with EcoRI and BamHI and the fragment containing the IFN- α coding sequence separated on agarose gel and combined with the non-IFN- α coding sequence obtained from an EcoRI/BamHI restriction of plasmid Hif-SN35 which is missing a PstI site adjacent the 3' end of the hybrid insert as compared to Hif-2h (supra). The resulting plasmid was then restricted with PstI/BglI to remove a portion of the amino terminal part of the IFN- α coding sequence. Inserted in its place were a series of IFN- α fragments prepared by digestion of plasmid Hif-2h with PvuII, treatment with Bal exonuclease, attachment of PstI

linkers and restriction with BglIII. The resulting plasmids thus contained a series of IFN- α 1 coding sequences which lacked various portions of their amino terminal sequences. One of these (plasmid 2H-M8) was digested with PstI and its nucleotide sequence determined. Sequencing revealed that the plasmid 2H-M8 contained several nucleotides between the Pst site and the first codon (CYS) of IFN- α 1. Therefore, the PstI digested plasmid 2H-M8 was treated with T4 polynuclease/dATP, S1 exonuclease and digested with SalI. This procedure generated a series of fragments whose IFN- α 1 coding sequences were missing portions of their amino terminal end. These fragments were then placed under LAC control by operatively linking them to a GUA-LAC fragment prepared from plasmid Lac3V8 by digestion with EcoRI, treatment with exonuclease S1 and digestion with SalI. The resulting series of plasmids thus had IFN- α 1 coding sequences lacking various portions of their amino terminal ends attached in a counterclockwise direction via an AUG to a fragment containing the LAC promoter.

Some of these plasmids were sequenced. One began at the fifth amino acid of IFN- α 1 and one at the tenth amino acid of IFN- α 1. In E. coli minicells (DS410) both of these plasmids produced polypeptides that displayed IFN activity. Therefore, not all of the IFN- α 1 protein is required for IFN activity.

IDENTIFICATION OF CLONES OF E. COLI CONTAINING
RECOMBINANT DNA MOLECULES WHOSE DNA INSERTS
WEAKLY CROSS-HYBRIDIZE TO THE INSERT IN Hif-4c
AND HAVE A DIFFERENT RESTRICTION MAP THAN THE
Hif-2h FRAGMENT

The comparison of the first 35 amino acids of authentic lymphoblastoid interferon (Zoon et al., supra, and M. Hunkapiller and L. Hood, supra) and the sequence deduced from Hif-2h fragment shows 9 differences. In all cases, the codons for the differing amino acids could be related by one-base changes. The amino acid compositions determined directly for authentic lymphoblastoid interferon

on the one hand and deduced from the sequence of the Hif-2h fragment on the other, also show striking differences in regard to their content of Gly, Pro, Cys and Met. These differences are too large to be explained by polymorphism. Instead, they most likely reflect the existence of at least two non-allelic genes, because the degree of divergence of the two proteins (26% mismatch) is similar to that between, for example, human and sheep B-globin (23% mismatch). Accordingly, the clones that displayed weak hybridization to fragment Hif-4c, identified previously (supra), were examined and a clone E.coli HB101 (Z-pBR322(Pst)/HcIF-II-206) was identified.

The hybrid plasmid Z-pBR322(Pst)/HcIF-II-206 ("HcIF-II-206") of this clone and its DNA insert "Hif-II-206 fragment" are weakly hybridizing to Hif-4c and Hif-2h fragment. E. coli transformed with plasmid Hif-II-206 produces polypeptides displaying a biological or immunological activity of HuIFN- α . Hif-II-206 fragment has a restriction map that is distinct from that determined for the Hif-2h fragment. A comparison of the two restriction maps is set forth in Fig. 11. Again, the absolute location of the restriction sites in the Hif-II-206 fragment is not determined by restriction mapping alone. However, sequencing of the nucleotide sequence of this insert, using the standard procedures described above, permits more absolute determination of these locations. However, because of the differences in the restriction map of the Hif-II-206 fragment as compared to the Hif-2h fragment, it is clear that the interferon genes of the two inserts have different nucleotide sequences.

Referring now to Figures 12-16, the nucleotide sequences of the inserted DNA sequence -- Hif-II-206 fragment -- of culture HcIF-G and the inserted DNA sequence -- Hif-2h fragment -- of culture HcIF-E (infra), determined previously, and the corresponding amino acid sequences of the proteins coded for by those nucleotide sequences are displayed. The nucleotide sequence of the Hif-II-206 fragment -- the Pst I fragment (790bp) of the

plasmid DNA isolated using the procedure of Method B as described by N. M. Wilkie et al., Nucl. Acids Res., 7, pp. 859-77 (1979) from culture HcIF-G -- was determined using the standard procedure of A. M. Maxam and W. Gilbert, supra. The sequencing strategy employed is shown in Figure 17.

Restricted DNA (usually about 10 µg) was 5' terminally labelled as described by N. Mantei et al., Gene, 10, pp. 1-10 (1980). Labeled fragments were cleaved with a second restriction enzyme, the products separated by electrophoresis through a 5% polyacrylamide gel in Tris-borate-EDTA buffer (A. C. Peacock and C. W. Dingman, Bioch., 6, pp. 1818-27 (1967)), extracted from the gel, and purified as described by W. Muller et al., J. Mol. Biol., 124, pp. 343-58 (1978).

Referring now to Figure 17, the various fragments for sequencing were prepared as follows:

25 and 26 -- cleavage of Hif-II-206 with PstI, labelling, cleavage with BglII, isolation of a PstI*-BglII fragment (257bp) ("25") and a PstI*-BglII fragment (279bp) ("26");

21, 22 and 23 -- cleavage of Hif-II-206 with PvuII, labelling, cleavage with BglII, isolation of a PvuII*-BglII fragment (88bp) ("21"), a PvuII*-BglII fragment (176bp) ("22") and a PvuI*-BglII fragment (214bp) ("23");

11, 12, 13, and 14 -- cleavage of Hif-II-206 with BglII, labelling, cleavage with PstI, isolation of a BglII*-PstI fragment (279bp) ("14") and a comigrating mixture of a BglII*-PstI fragment and a BglII*-BglII* fragment. Cleavage of the mixture with PvuII and isolation of a BglII*-PstI fragment (257bp) ("13"), a BglII*-PvuII fragment (176bp) ("12") and a BglII*-PvuII fragment (88bp) ("11").

27L, 27U, 41, 43, 44 and 45 -- cleavage of Hif-II-206 with HinfI, labelling, isolation of precursor fragments: HinfI*-HinfI* (113bp) ("27P"), HinfI*-HinfI* (146bp) ("28P"), HinfI*-HinfI* (159bp) ("30P"), HinfI*-

HinfI* (397bp) ("31P") and HinfI*-HinfI* (1522bp) ("32P"). Cleavage of 28P with MboII and isolation of a fragment HinfI*-MboII (112bp) ("41"). Cleavage of 30P with MboII and isolation of a fragment HinfI*-MboII (126bp) ("43"). Cleavage of 31P with PstI and isolation of a fragment HinfI*-PstI (151bp) ("44"). Cleavage of 32P with PstI and isolation of a fragment HinfI*-PstI (139bp) ("45"). Strand separation of 27P to yield two strands ("27U" and "27L").

All segments, except 27U and 27L, were sequenced on both strands and across the restriction sites that served as origins for sequencing, except for the BglII site at position 185.

From a comparison of the amino acid sequence coded for by the two inserts it is apparent that the Hif-II-206 fragment codes for an interferon-like protein having one less amino acid than does the Hif-2h fragment (amino acid 44 (Asp) present in Hif-2h is missing in Hif-II-206). Moreover, 10% of the nucleotide positions and 17% of the derived amino acid residues of the two fragments are different.

In addition, when compared to the 35 amino acids determined for the amino terminus of lymphoblastoid interferon (K. C. Zoon et al., Science, 207, pp. 527-28 (1980)), the insert Hif-II-206 codes for a protein that differs in 5 residues from the 35 amino acid residues determined by Zoon et al., supra. Therefore, at least three different IFN genes of the leukocyte type (α type) must exist -- Hif-2h fragment, Hif-II-206 fragment and the gene coding for Zoon's IFN. In accordance with the newly proposed nomenclature for interferon, hereinafter the proteins coded for by these genes will be identified as follows:

<u>IFN Gene Source</u>	<u>Protein</u>
Hif-2h	IFN- α 1
Hif-II-206	IFN- α 2
IFN from lymphoblastoid cells (Zoon et al. (<u>supra</u>))	IFN- α 3

The differences between IFN- α 1 and IFN- α 2 are also reflected in their varying activities on human CCL23 and bovine embryonic kidney (BEK) cells:

<u>Extract</u>	<u>Relative IFN Activity*</u>		
	<u>CCL23</u>	<u>BEK</u>	<u>Ratio</u>
Hif-II-206 (IFN- α 2)	1.7	1.0	1.7:1
Hif-SN35** (IFN- α 1)	0.05	1.0	1:20

Therefore, IFN- α 1 is about 30 times less active on human cells than IFN- α 2. Yet, they are both about equally active on bovine cells. Therefore, IFNs may, in addition to their use as antiviral and antitumor or anticancer agents in humans, also be useful in treating these conditions in cattle. For example, preparations of HuIFN- α could be employed in a standard manner (supra) in treating FMDV and other well known viral infections of cattle. This is even more particularly true for IFN- α 1 since its activity on bovine cells is about 20 times greater than its activity on human cells.

* E. coli HB101 containing the hybrid plasmid were grown in tryptone medium with shaking to an OD₆₅₀ of about 1-2. The cells were harvested, weighed, resuspended in 1/100 or 1/20 of the original culture volume and lysed by the lysozyme-freeze-thaw method (S. Nagata et al., Nature, 284, pp. 316-20 (1980)). The S-30 supernatants were tested by CPE reduction assay in microtiter plates. Extracts from control cells were negative (<1 I.U./ml). Human CCL23 cells and bovine embryonic kidney (BEK) cells (FLOW) were grown in MEM-10% fetal calf serum. Exposure to IFN-containing extracts was for 24 h. The cells were challenged with an appropriate dilution of Mengo virus and stained 24 h later. Values were estimated visually relative to partially purified leukocyte IFN (preparation PIF, a gift of K. Cantell) of known titer. This preparation was about 3x more active on human than on bovine cells.

** The comparison of the restriction maps of Hif-SN35 and Hif-2h suggest that they are polymorphic variants of each other (supra).

Because of the improved yield attained in the case of IFN- α 1 with construction C8 (supra), a similar construction was made for IFN- α 2. Z-pBR322(Pst)/Hif-II-206 was cleaved completely with BspI and partially with PvuII (at P₁) (Figure 28) and the 867 bp fragment was isolated from a 6% polyacrylamide gel. This fragment was then ligated to a 2590 bp PvuII fragment of C8-IFN- α 1.* The resulting hybrid plasmid was used to transform E. coli HB101 and the clones screened for IFN activity. One clone displaying a high activity was selected and designated E. coli HB101 (C8-IFN- α 2).

DNA sequencing of hybrid plasmid C8-IFN- α 2 revealed that it like C8-IFN- α 1 had a coding sequence following the initiator triplet that determined the first seven amino acids of β -galactosidase, a Pro residue generated by the fusion and amino acids 16 to 23 of the IFN- α 2 signal sequence. Therefore, again a fused protein containing IFN- α 2 is likely to be expressed.

Minicells transformed with this plasmid gave 100-200 million units per l of IFN or 20000 to 40000 times higher yields of IFN- α 2 than minicells transformed with unmodified Z-pBR322(Pst)/Hif-II-206.

A comparison of the relative yields of C8-IFN- α 1 (~50 x 10⁶ units/liter) and C8-IFN- α 2 (~100-200 million units per liter) is at first surprising because IFN- α 2 has been shown to be about 30 times more active than IFN- α 1 on human cells (supra). However, quantitative analysis of the amount of the two proteins made by minicells transformed with the two C8 plasmids revealed that in C8-IFN- α 1 about 5 to 6 times more protein than in C8-IFN- α 2 was being made. Therefore, the yields measured on the basis of IFN activity had been skewed by the much greater amount of protein made in the case of C8-IFN- α 1.

Referring now to Figure 26, another construction in an attempt to improve the yield of IFN- α 2 is described.

* C8-IFN- α 1 has three PvuII restriction sites (Figure 28). The 2590 bp fragment is between P₁ and P₃.

First, an expression plasmid containing the LAC Alu fragment was prepared by restricting the known lac promoter with AluI and extending the fragment as shown (for one termini) in Figure 27 with EcoRI linkers (prepared by collaborative research). The extended fragment was then inserted into pBR322 at the EcoRI site and a small EcoRI-EcoRI fragment deleted from the construction. The resulting plasmid, designated 404 in Figure 26 was cleaved with HindIII and PvuII for insertion of the IFN- α 2 containing fragment. That IFN- α 2 fragment was prepared by partial Sau3A restriction of Z-pBR322(Pst)/HcIF-II-206 ("206" in Figure 26), extension of the Sau3A fragment with HindIII linkers (Figure 27) and cleavage with BspI. After insertion of this fragment into the HindIII-PvuII cleaved plasmid 404, the resulting plasmid was restricted with HindIII and EcoRI, treated with S1 nuclease to bring the LAC promoter closer to the IFN- α 2 gene and religated. This construction identified as plasmid LAC-AUG(α 2) has the IFN- α 2 DNA sequence under the control of the LAC promoter. Moreover, the IFN- α 2 sequence immediately follows the initiating AUG codon of that promoter (see Figure 27). Therefore, at least a portion of the IFN produced by these plasmids will be mature IFN, e.g., IFN without any amino acids from the signal sequence. In minicells the yield of IFN- α 2 obtained with plasmid LAC-AUG(α 2) was 5-10 million units per liter.

Another IFN- α 2 construction based on similar linking principles has also been made. Here, the penicillinase expression control sequence of pBR322 has been connected via an AUG initiator codon to the IFN- α 2 gene

from Hif-II-206.* This plasmid designated as β -lac-AUG(α 2) when used to transform host cells affords the production of IFN- α 2 without fusion to other protein sequences. In minicells, yields of 50-100 million units per liter have been observed. This plasmid is the most preferred plasmid for use in accordance with this invention. It is also preferred for use with the other IFN- α genes disclosed herein. The preferred host in accordance with this invention is E. coli DS410 (ara azide TonA lac y Tsx min a min b gal λ xyl step^R). This strain has been deposited together with an example of the preferred plasmid β -lac-AUG(α 2) as HcIF-K.

Other constructions using various promoter sequences, ribosome binding sites, Shine-Dalgarno sequences and DNA sequences between the promoter and the AUG initiator codon and using various of the IFN- α genes disclosed herein can also be constructed using similar methods and principles. These constructions are of course envisioned by this invention and are within the scope thereof.

HYBRID MOLECULES OF IFN- α 1 AND IFN- α 2

A number of hybrid molecules of IFN- α 1 and IFN- α 2 have been constructed. Surprisingly these hybrid constructions have quantitatively different properties and activities as compared either of their parents -- IFN- α 1 or IFN- α 2.

* This construction could most effectively be made by partial digestion of pBR322 with MboII, treating with S1, attaching EcoRI linkers as before and reinserting the fragment into the EcoRI site of pBR322 and deleting one EcoRI site. The resulting plasmid (" β -lac-AUG plasmid") could then be combined with an S1 treated (mild) HindIII linker-BspI fragment of 206 described previously. After cleavage with EcoRI and treating with S1 and phosphatase, the expression plasmid β -lac-AUG(α 2) is isolated. Constructions with other genes could be done in a similar fashion by merely utilizing the constructed β -lac-AUG plasmid for insertion of other genes or constructions or more preferably by using the Sau3A site in plasmid β -lac-AUG(α 2) itself.

Referring now to Figure 28, a schematic outline of the construction of four of these hybrid molecules is displayed. For convenience these hybrid molecules are designated as plasmids I, II, III and IV. In these constructions digestions with restriction enzymes (obtained from Biolabs, with the exception of BspI, a gift of Dr. Kiss) were carried out essentially as recommended by the suppliers. Partial DNA cleavages were carried out with decreased amounts of enzyme. After heat inactivation (65°C, 30 min) of the restriction enzyme, the samples were adjusted to 50 mM Tris-HCl (pH 8) and if required calf intestine alkaline phosphatase (Boehringer) (1 vol per µg DNA) were added. After 30 min at 37°C, the samples were extracted with phenol and ether. In most cases the DNA fragments were separated on low temperature gelling agarose (0.8%). For ligation the fragment containing gel pieces (about 20 µl each) were melted at 65°C, cooled to 37°C and 20 U per µl T4 DNA ligase was added. The mixture was kept at 15°C for 16 hrs and ligation occurred in the solidified gel (H. Lehrach, personal communication (1980)). One tenth vol of 100 mM Tris-HCl (pH 7.5), 100 mM CaCl₂, 100 mM MgCl₂ was added, the sample heated 5 min at 65°C and cooled to 37°C. The samples were then added to Ca²⁺ - treated mini cells, incubated at 0°C for 20 min, heated 1 min at 42°C and 10 min at 20°C and 1 ml tryptone medium added. After incubation for 60 min at 37°C, the cultures were plated on to agar plates containing the appropriate antibiotics. All of the plasmids were characterized by nucleotide sequence analysis across the joint in the IFN sequence.

Hybrid molecule I, an α-1(PvuII)α-2 hybrid, was constructed by partially cleaving C8-IFN-α1 (supra) (C8-α1 in Figure 28) with PvuII, dephosphorylated, cleaved with PstI and the PstI-PvuII(P₂) 1346 bp fragment isolated. This fragment was ligated to a 2135 bp PstI(a)-PvuII(P₂) fragment prepared by totally cleaving C8-IFN-α2 (supra) (C8-α2 in Figure 28) with PvuII and partially cleaving it with PstI.

Hybrid molecule II, an α -1(BglII) α -2 hybrid, was constructed by cleaving hybrid molecule I with BglII. After dephosphorylation, the large BglII fragment was isolated and ligated to the small BglII fragment of C8-IFN- α 2. After cloning, the hybrid plasmid having the small BglII fragment in the correct orientation was identified by restriction analysis.

Hybrid molecule III, an α -2(PvuII) α -1 hybrid, was constructed by partially cleaving C8-IFN- α 1 with PvuII, dephosphorylating, cleaving with AvaI and isolating the 1686 bp PvuII(P₂)-AvaI and 3233 bp PvuII(P₁)-AvaI fragments. These fragments were then ligated to the 300 bp PvuII(P₁)-PvuII(P₂) fragment of the HcIF-II-206 (*supra*) (SN206 in Figure 28) and the plasmid containing the small PvuII fragment identified by assaying transformed *E. coli* strains for IFN- α activity.

Hybrid molecule IV, an α -2(BglII) α -1 hybrid, was constructed by cleaving C8-IFN- α 1 with BglII and AvaI and the 1776 bp fragment isolated. This fragment was then ligated to the 3543 bp BglII-AvaI fragment of hybrid molecule III.

The biological activities of the different interferon species relative to each other were also determined. Cultures of minicells (DS410) transformed with the various plasmids were grown and the bacteria harvested by centrifugation, washed with PBS, suspended in PBS (about 1/20 of the original vol), incubated for 60 min at 0°C with 1 mg/ml lysozyme, 10 mM EDTA, freeze thawed four times, sheared by passing five times through a syringe and cleaved by centrifugation.

The activities on human, mouse, guinea pig and bovine cells were as follows:

<u>Protein Source</u>	<u>Relative IFN Activity</u>			
	<u>Human (FS4)</u>	<u>Bovine (BEK)</u>	<u>Mouse (L)</u>	<u>Guinea Pig</u>
C8-IFN- α 2	1	1	0.01	0.03
C8-IFN- α 1	0.03	1	0.3	0.03
Hybrid I	0.001	1	0.003-0.008	0.003-0.01
Hybrid II	0.001	1	0.001	0.01

Hybrid III	1	1	1	0.3
Hybrid IV	0.1	1	2	0.1-0.005
Negative Control	--	--	--	--

Surprisingly, all of the interferons have about the same activity on bovine cells, yet IFN- α 1 and the two hybrid IFN's (I and II) which have the amino terminal moiety of IFN- α 1 have about a 10- to 1000-fold lower activity on human cells than IFN- α 2 and the two hybrid IFN's (III and IV) having the amino terminal moiety of IFN- α 2. It is even more striking that the two hybrid IFN's (I and II) with the amino terminal part of IFN- α 1 have more than a 10-fold lower activity on human cells than IFN- α 1 itself. Yet, one of the hybrids (III) with the amino terminal part of IFN- α 2 has about the same activity on human cells as IFN- α 2.

IDENTIFICATION OF CHROMOSOMAL GENES FOR IFN- α

A collection of hybrid phage derived from fragments of fetal human chromosomal DNA which had been generated by partial cleavage with HaeIII and AluI, and joined with EcoRI linkers to λ Charon 4A arms has been prepared by R. M. Lawn et al., Cell, 15, pp. 1157-74 (1978). This gene bank was screened by an "in situ" procedure (W. D. Benton and R. W. Davis, Science, 196, pp. 180-82 (1977); T. Maniatis et al., Cell, 15, pp. 687-701 (1978)) using as a probe the ^{32}P -labelled IFN- α 1cDNA insert excised from pBR322(Pst)/Hif-2h. Sixteen hybridization-positive phage clones were isolated from 240,000 plaques by repeated plaque purification (T. Maniatis et al., supra). Ten of the hybrid phage DNA preparations were cleaved with HindIII, TacI, HhaI, BamHI, EcoRI and BglII, respectively, and the fragments separated by electrophoresis on an agarose gel, transferred to a Millipore membrane (E. M. Southern, J. Mol. Biol., 98, pp. 503-17 (1975)) and hybridized with the ^{32}P -labelled

Hif-2h cDNA insert. Figure 18 summarizes the results in the form of partial restriction maps and various tables. As displayed there, for each hybrid phage DNA preparation at least a few characteristic restriction sites were established and the region(s) hybridizing to the IFN- α 1 gene probe delineated (black arrow).*

Referring now to Figures 18 and 24, it can be seen that clones chr-3 and chr-26 may represent DNA segments which overlap over much of their length because they have several EcoRI and HindIII fragments in common. In addition, the hybridizing portion of chr-1 and one of the hybridizing portions of chr-10 may be the same because the HindIII-HindIII and EcoRI-EcoRI fragments, which hybridize with the Hif-2h probe have the same length (3.2kb and 0.95kb, respectively). It also appears that the "right-hand" hybridizing portion of chr-16 (labelled "r" in Figure 18) may be identical with the hybridizing portion of chr-35, although orientated in the opposite direction, inasmuch as each of the two clones yields a 1.4kb BglII-BglII fragment and a 2kb EcoRI-EcoRI fragment which hybridize with the Hif-2h cDNA probe. Therefore, most likely the chr-16 and chr-35 inserts overlap.

Accordingly, it appears that the 13 hybridizing portions of the 11 hybrid chromosomal DNAs fall into not less than 10 distinct classes -- chr-1, chr-3, chr-12, chr-13, chr-16 (left hand, labelled "l" in Figure 18) chr-26, chr-30, chr-35, chr-19 and chr-27.

Referring now to Figure 24, the overlapping of chr-1, chr-3, chr-10 and chr-26 and that of chr-16 and chr-35 are displayed.

The above data suggests that the genome of an individual human contains not less than 10 different DNA sequences that cross-hybridize to Hif-2h. This conclusion is reinforced by the fact that the proportion of fragment

* The shaded area on chr-16 in Figures 18 and 24 represents a sequence which hybridizes weakly to Hif-2h cDNA but did not display R-looping.

Hif-2h-related sequences detected in the clone bank is about 1 in 16,000. Assuming a value of 3×10^9 bp for the haploid human genome, the expected value for a single gene copy with an average DNA fragment size of 16kb (the average value of the clones examined) is about 1:190,000. Therefore, the frequency of Hif-2h related fragments is 12 times higher than expected for a single gene.

In comparison to these data, when Lawn et al (supra) screened 300,000 plaques from the same gene bank with a β -globin cDNA probe, only 2 positive clones were identified -- the expected value being 1.6:300,000. Therefore, there may be 10-15 distinct chromosomal DNA segments in the human genome that cross-hybridize to the Hif-2h fragment or IFN- α 1cDNA.

FURTHER CHARACTERIZATION OF Hif-chr35

As an illustration only, the hybridizing sequence of chr-35 ("Hif-chr 35") was further characterized. It is to be understood that the hybridizing portions of the previously described chromosomal hybrid phages could similarly be characterized and handled without departing from the scope of this invention.

The hybridizing portion of chr-35 ("Hif-chr35") (and the right-hand segment of Hif-chr16 to which it is most likely identical, supra) is the only hybridizing chromosomal DNA portion with a BglII site. Since IFN- α 1 and IFN- α 2 cDNA's have 1 and 2 BglII sites, respectively, within their coding sequences, it seems likely that Hif-chr35 is a counterpart to one of the two previously cloned interferon genes. Hif-chr35's strong hybridization to the 3' terminal Hif-2h cDNA fragment (containing only the 3' non-coding region) compared to the weaker hybridization of the other of the chromosomal DNAs to this probe supports a likely correspondence of Hif-chr35 and Hif-2h (F. Kafatos et al., Proc. Natl. Acad. Sci USA, 74, pp. 5618-22 (1977)).

To analyze the Hif-chr35 fragment further, a HindIII-BamHI fragment was excised from chr-35. This fragment (3.4kb) contains the hybridizing portion ("Hif-chr35") of chr-35. This fragment was subcloned into the PstI site of pBR322 using the well-known dC-dG tailing procedures (L. Villa-Komaroff et al., supra) and E.coli HB101 transformed with the resultant recombinant DNA molecule using well-known procedures (e.g., S. Nagata et al., Nature, 284, pp. 316-20 (1980)).

Clones of these transformants were screened by in-situ colony hybridization (D. Hanahan and M. Meselson, Gene, 10, pp. 63-67 (1980)) with the ^{32}P -labelled Hif-2h fragment (supra) and plasmid DNA -- Z-pBR322(Pst)/HchrIF-35HB "HchrIF-35HB" -- was separated from the positive clones (N. M. Wilkie et al., Nucleic Acids Res., 7, pp. 859-77 (1979); Method B). The orientation of the hybrid insert "HchrIF-35HB fragment" in the plasmid with respect to the β -lactamase gene of pBR322 was determined by EcoRI cleavage and sizing of the resulting fragments. The insert orientated to coincide with that of β -lactamase was designated α and the opposite orientation β .

Cultures of these positive clones were grown to an apparent $\text{OD}_{650} = 0.8$ and the bacteria harvested and lysed by the lysozyme-freeze-thaw method described in S. Nagata et al., supra. Seven of the 10 clones examined showed IFN- α activities of 75 to 500 units/g of cells (cytopathic effect reduction assay).

The DNA insert of one of these 7 IFN-producing clones -- E. coli HB 101 (Z-pBR322(Pst)/HchrIF-35HB α) -- was further characterized by restriction analysis and nucleotide sequence determination. Plasmid DNA (HchrIF-35HB α) was prepared from the clone as described previously and the restriction sites determined by Smith-Birnsteil mapping (H. O. Smith and M. L. Birnsteil, Nucl. Acids Res., 3, pp. 2387-98 (1976)): HchrIF-35HB α was digested with EcoRI, labelled at the 5' termini and digested with BglII (and PstI to cleave an undesired fragment of about 1kb). The 1.04kb EcoRI-BglII (3' proximal) and the 0.96kb

EcoRI-BglII (5' proximal) fragments were isolated by agarose gel electrophoresis as described by A. C. Peacock and C. W. Dingman, Biochemistry, 6, pp. 1818-27 (1967)). Both fragments were partially cleaved with HinfI, BspI and MboII respectively and the products separated on a 1% agarose gel in Tris-acetate buffer (ph 7.8) containing 1 µg/ml ethidium bromide. After staining, the radioactive bands were visualized by autoradiography. The BstNI and HgiAI sites were similarly determined on the 1.04kb (3' proximal) fragment. The results of the analysis is shown in Figure 19.

For nucleotide sequence determination, HchrIF-35HBα was cleaved with various restriction enzymes, the products separated by electrophoresis through a 5% polyacrylamide gel in Tris-borate-EDTA buffer (A. C. Peacock and C. W. Dingman, supra) and extracted from the gel and purified as described by W. Maller et al., J. Mol. Biol., 124, pp. 343-58 (1978).

The sequencing strategy employed is depicted in Figure 19 and described as follows:

1 and 2 -- cleavage of HchrIF-35HBα with BglII, labelling, cleavage with EcoRI and PstI and isolation of a BglII*-EcoRI fragment (940bp) ("1") and a BglII*-EcoRI (360bp) ("2");

3 and 4 -- cleavage of HchrIF-35HBα with EcoRI, labelling, cleavage with BspI and isolation of an EcoRI*-BspI fragment (680bp) ("3") and an EcoRI*-BspI, fragment (880 bp) ("4");

5, 6, 7 and 8 -- cleavage of HchrIF-35HBα with PvuII, labelling, cleavage with BglII and EcoRI and isolation of PvuII*-EcoRI fragment (780bp) ("5"), a PvuII*-BglII (215bp) ("6"), a PvuII*-BglII fragment (90bp) ("7"), and a PvuII*-EcoRI fragment (290bp) ("8");

9 and 10 -- cleavage of HchrIF-35HBα with EcoRI, isolation by 1% agarose gel electrophoresis in Tris-borate EDTA buffer of a 1300 bp EcoRI-EcoRI fragment, further cleavage with HinfI and isolation of a HinfI-HinfI fragment (450bp), and a HinfI-HinfI fragment (180bp).

Labelling, of the larger HinfI-HinfI fragment and cleavage with MboII permitted isolation of a HinfI*-MboII (190bp) ("9"). Labelling of the shorter HinfI-HinfI fragment and cleavage with AvaII permitted isolation of a HinfI*-AvaII fragment (150bp) ("10");

11 -- cleavage of HchrIF-35HB α with MboII, labelling, cleavage with BglII, and isolation of a MboII*-BglII fragment (465bp) ("11")

12, 13 and 14 -- cleavage of HchrIF-35HB α with BspI and BglII, isolation by agarose electrophoresis as above of a 1200bp BspI-BglII fragment and (a) cleavage with HgiAI, labelling, cleavage with MboII and isolation of an HgiAI*-MboII fragment (300bp) ("12") and an HgiAI*-MboII fragment (360bp) ("13"), or (b) cleavage with BstNI, labelling, cleavage with EcoRI and isolation of a BstNI*-EcoRI fragment (380bp) ("14").

The various fragments were sequenced by the Maxam-Gilbert procedure (supra). All fragments were sequenced on both strands and across the restriction sites that served as origins for sequencing.

A comparison of the nucleotide sequence of the coding region of HchrIF-35HB α and that of Hif-2h (coding region) (Figures 8-10 compared to Figures 20-23) reveals that they are identical. In particular, it is surprising that there is no indication of the presence of introns within the coding sequence of the HchrIF-35HB α fragment, i.e., between the HinfI site in the 5' non-coding region and the EcoRI site in the 3' non-coding region. Thus, no intron could be detected in the chromosomal sequence corresponding to mature IFN- α mRNA..

FURTHER CHARACTERIZATION OF Hif-chr 26 AND Hif-chr 3

The gene-containing segments of chr-3 and chr-26, which appear identical by heteroduplex analysis but differ in at least one BglII restriction site were examined by nucleotide sequencing. Five nucleotide

differences in 725 base pairs were found. Only two of these appear in the coding sequences. Since not only the genes, but at least 3.5 Kbp preceding and 6.0 Kbp following them formed a perfect heteroduplex and because of the relatively low sequence divergence which entails only 2 amino acid changes, it appears that Hif-chr3 and Hif-chr26 are allelic forms of the same gene. These are designated IFN- α 4a (Hif-chr3) and IFN- α 4b (Hif-chr26). The nucleotide sequence and corresponding amino acid sequence of IFN- α 4b determined by conventional sequencing techniques described previously is displayed in Figures 29-32.

A comparison of Figures 29-32 with Figures 8-10, 12-16 and 20-23 reveals that the proteins encoded by each of the sequences differ from each other in about 15% of their residues. This divergence is typical for products of non-allelic genes which have diverged 20-90 million years ago.

Expression Of Hif-chr35 In Mouse Cells

Plasmid Z-pBR322(Pst)/HchrIF-35HB α (supra) was used as a source of a Hif-chr35 fragment for expression in mouse cells. The plasmid was restricted with PstI and treated with 5' exonuclease to remove the 5' dG tails. This fragment was then inserted into a 5' dG-tailed KpnI fragment of a plasmid prepared by joining the BamHI-BamHI fragments of pBR322 and polynoma DNA. The resulting vector was used to transform mouse 3T3 cells using the calcium phosphate technique (N. Mantei et al., Nature, 281 pp. 40-46 (1979)). These transformed cells are designated for convenience Mouse 3T3 (polynoma-Hif-chr35). After 20-40 hours, assays revealed an IFN- α activity of 300 units/ml of IFN- α on human cells and about 3000 units/ml of IFN- α on bovine cells.

It should of course be understood that the nucleotide sequences depicted in Figures 8-10, 12-16, 20-23 and 29-32 do not take into account any modifications to the nucleotide sequences such as mutation, including

single or multiple, base substitutions, insertions, inversions or deletions which may have already taken place or which may subsequently be employed. Moreover, the sequence also does not take into account the possible substitution of other codons coding for the same amino acid as a codon depicted in these figures. Therefore, it should be understood that such modified sequences as code for polypeptides displaying an immunological or biological activity of IFN- α are also within this invention.

In addition, it is to be understood that the amino acid sequences depicted in Figures 8-10, 12-16, 20-23 and 29-32 do not take into account any modifications to the polypeptides caused by their interaction with in vivo or in vitro agents e.g. in vivo glycosylation enzymes. Therefore, it must be understood that fragments and derivatives of these polypeptides that display an immunological or biological activity of IFN- α are also part of this invention.

PRODUCTION OF POLYPEPTIDES DISPLAYING AN
IMMUNOLOGICAL OR BIOLOGICAL ACTIVITY OF
INTERFERON IN BACTERIAL HOSTS

Since the cytopathic effort reduction assay (W. E. Stewart II and S. E. Sulkin, S. E. Proc. Soc. Exp. Biol. Med., 123, pp. 650-53 (1966)) can detect minute amounts of IFN -- less than one active molecule per bacterial cell -- lysates of E. coli HB101 infected with the ten hybrid λ phages, described previously, were assayed for the presence of IFN. Seven of the eleven phages (all except chr-10; chr-12, chr-19 and chr-27) gave lysates containing IFN activity ranging from 3 to 50 units/ml. In the case of chr-10 and chr-12, the hybridizing (to Hif-2h) HindIII-HindIII or EcoRI-EcoRI fragments, subcloned into the PstI site of pBR322, as described previously, expressed IFN- α activity in E. coli. Since E. coli is believed to be incapable of splicing mRNA (O. Mercereau-Puijalon and P. Kourilsky, Nature,

279, pp. 647-49 (1979)), these IFN- α chromosomal genes most likely do not contain introns in their coding region.

FINAL CONCLUSIONS

We have isolated a set of recombinant DNA molecules containing cDNA prepared from poly(A) RNA from Sendai virus-treated (induced) human leukocytes, representatives of which have the following properties:

(1) They hybridize to poly(A) RNA from induced but not from non-induced human leukocytes.

(2) They hybridize to IFN- α mRNA as shown by their capacity to select this RNA from a mixture of RNAs, and by their capacity to inhibit (reversibly) translation of interferon mRNA in the hybrid arrested translation assay.

(3) E. coli containing certain members of the set produce a compound with the following properties:

(a) It is sensitive to trypsin

(b) It exhibits IFN- α activity in a human cell system and only slight activity in a mouse cell system

(c) It has a molecular weight between 20'000 and 30'000 (19,388 based on the nucleotide sequencing of Figs. 8-10)

(d) The IFN- α activity is specifically inhibited by antibody to human leukocyte interferon.

(4) The DNA inserts of the hybrid plasmids of this invention are able, in addition to their ability to select IFN- α mRNA from a mixture of RNAs, to select IFN- α DNA from mixtures of various sources including cDNAs and from hybrid phage gene banks of chromosomal DNA.

(5) A number of different chromosomal genes for IFN- α exist. It is unexpected that these genes lack introns and permit direct expression of interferon and interferon-like polypeptides in appropriate hosts.

(6) At least three of the nucleotide sequences of the DNA inserts of these recombinant DNA molecules are different and suggest the existence of at least three non-allelic genes for IFN- α .

(7) The proteins coded for by these three nucleotide sequences are different from the 35 amino acids determined from authentic lymphoblastoid interferon.

(8) Hybrid proteins prepared for various combinations of IFN- α gene segments display quantitatively different properties than each other or their parents and proteins having additional amino acids fused to IFN- α or proteins comprising IFN- α without a portion of its amino terminal sequence display IFN activity.

These properties demonstrate that the recombinant DNA molecules described by this invention contain at least a part of the coding sequence for human leukocyte interferon and that some of these plasmids lead to expression in E. coli of a polypeptide with an immunological or biological activity of human leukocyte interferon. It should also be evident that the polypeptides disclosed herein may be fragmented, modified or derivatized, as is well known in the protein art, without departing from the scope or disclosure of this invention.

Micro-organisms and recombinant DNA molecules prepared by the processes described herein are exemplified by cultures deposited in the culture collection Deutsche Sammlung von Mikroorganismen, in Gottingen, West Germany on January 7, 1980, and identified as HcIF-A to E:

- A: E. coli HB101 (Z-pBR322(Pst)/HcIF-4c)
- B: E. coli HB101 (Z-pBR322(Pst)/HcIF-2h)
- C: E. coli HB101 (Z-pBR322(Pst)/HcIF-SN35)
- D: E. coli HB101 (Z-pBR322(Pst)/HcIF-SN42)
- E: E. coli HB101 (Z-pKT287(Pst)/HcIF-2h-AH6)

These cultures were assigned accession numbers DSM 1699-1703, respectively.

In addition, micro-organisms and recombinant DNA molecules prepared by the processes described herein are exemplified by cultures deposited in the culture collection of the American Type Culture Collection, Rockville, Maryland on March 27, 1980 and identified as HcIF-G to H, and assigned ATCC accession numbers 3/633 and 3/634 respectively:

G: E. coli HB101 (Z-pBR322(Pst)/HcIF-II-206)

H: E. coli HB101 (Z-pBR322(Pst)/HcIF-SN35-AHL6)

Other micro-organisms prepared by the processes described herein are exemplified by cultures deposited in the culture collection Deutsche Sammlung von Mikroorganismen, in Gottingen, West Germany on October 1, 1980 and identified as HchrIF-A through J, and assigned accession numbers DSM 1914-1923:

- A. subcloned HindIII fragment of chr 3 in E. coli HB101;
- B. subcloned EcoRI fragment of chr 12 in E. coli HB101;
- C. subcloned HindIII fragment of chr 12 in E. coli HB101;
- D. subcloned EcoRI fragment of chr 13 in E. coli HB101;
- E. subcloned EcoRI fragment of chr 23 in E. coli HB101;
- F. subcloned HindIII fragment of chr 23 in E. coli HB101;
- G. subcloned EcoRI fragment of chr 26 in E. coli HB101;
- H. subcloned HindIII fragment of chr 26 in E. coli HB101;
- I. subcloned HindIII/BamHI fragment of chr 35 in E. coli HB101;
- J. subcloned BamHI fragment of chr 35 in E. coli HB101.

Finally, micro-organisms prepared by the processes described herein are exemplified by cultures deposited in the American Type Culture Collection, Rockville, Maryland, on December 15, 1980 and identified as HchrIF-K through HchrIF-Q and HcIF-I through HcIF-K, and assigned ATTC accession numbers as follows:

31760 HchrIF-K
31761 HchrIF-L
31762 HchrIF-M
31763 HchrIF-N
31764 HchrIF-O
31765 HchrIF-P
31766 HchrIF-Q

31767 HcIF-I
31768 HcIF-J
31769 HcIF-K

K. subcloned Tac-Tac fragment of chr 23 in E. coli HB101.

L. subcloned BglII-BglII fragment of chr 10 1 in E. coli HB101.

M. subcloned HindIII-HindIII fragment of chr 10r in E. coli HB101.

N. subcloned BglII-BglII fragment of chr 26 in E. coli HB101.

O. subcloned HindIII-HindIII fragment of chr 30 in E. coli HB101.

P. subcloned BglII-Tac fragment of chr 13 in E. coli HB101.

Q. subcloned BglII-Tac fragment of chr 16 1 in E. coli HB101.

HcIF-I: E. coli DS410 (C8-IFN- α 2)

HcIF-J: E. coli DS410 (LAC-AUG(α 2))

HcIF-K: E. coli DS410 (β -Lac-AUG(α 2))

While we have hereinbefore presented a number of embodiments of this invention, it is apparent that our basic construction can be altered to provide other embodiments which utilize the processes and compositions of this invention. Therefore, it will be appreciated that the scope of this invention is to be defined by the claims appended hereto rather than the specific embodiments which have been presented hereinbefore by way of example.

CLAIMS

1. A DNA sequence selected from the group consisting of the DNA inserts of Z-pBR322(Pst)/HcIF-4c, Z-pBR322(Pst)/HcIF-2h, Z-pBR322(Pst)/HcIF-SN35, Z-pBR322(Pst)/HcIF-SN42, Z-pKT287(Pst)/HcIF-2h-AH6, DNA sequences which hybridize to any of the foregoing DNA inserts, DNA sequences, from whatever source obtained, including natural, synthetic or semi-synthetic sources, related by mutation, including single or multiple, base substitutions, deletions, insertions and inversions to any of the foregoing DNA sequences or inserts, and DNA sequences comprising sequences of codons which on expression code for a polypeptide displaying similar immunological or biological activity to a polypeptide coded for on expression of the codons of any of the foregoing DNA sequences and inserts.

2. A DNA sequence according to claim 1 wherein said DNA sequence which hybridizes to said DNA insert is selected from the group consisting of the DNA inserts of Z-pBR322(Pst)/HcIF-II-206 or Z-pBR322 (Pst)/HcIF-SN35-AHL6, DNA sequences which hybridize to any of the foregoing DNA inserts, DNA sequences, from whatever source obtained, including natural, synthetic or semi-synthetic sources, related by mutation, including single or multiple, base substitutions, deletions, insertions and inversions to any of the foregoing DNA sequences or inserts, and DNA sequences comprising sequences of codons which on expression code for a polypeptide displaying similar immunological or biological activity to a polypeptide coded for on expression of the codons of any of the foregoing DNA sequences and inserts.

3. A DNA sequence according to claims 1 or 2 wherein said DNA sequence which hybridizes to said DNA insert is selected from the group consisting of Hif-chr1, Hif-chr3, Hif-chr12, Hif-chr13, Hif-chr16-2, Hif-chr26, Hif-chr30, Hif-chr35, DNA sequences which hybridize to any of the foregoing DNA sequences, DNA sequences, from

whatever source obtained, including natural, synthetic, or semi-synthetic sources, related by mutation, including single or multiple, base substitutions, deletions, insertions and inversions, to any of the foregoing DNA sequences and DNA sequences comprising sequences of codons which on expression code for a polypeptide similar in immunological or biological activity to a polypeptide coded for on expression of any of the foregoing DNA sequences.

4. A DNA sequence according to claim 1 or 2 wherein said DNA sequence which hybridizes to said DNA insert is selected from the group consisting of Hif-chr19, Hif-chr27, DNA sequences which hybridize to any of the foregoing DNA sequences, DNA sequences, from whatever source obtained, including natural, synthetic, or semi-synthetic sources, related by mutation, including single or multiple, base substitutions, deletions, insertions and inversions, to any of the foregoing DNA sequences and DNA sequences comprising sequences of codons which on expression code for a polypeptide similar in immunological or biological activity to a polypeptide coded for on expression of any of the foregoing DNA sequences.

5. A DNA sequence according to any one of claims 1 to 4 selected from the group consisting of DNA sequences of the formula: ATGGCCTCGCCCTTGCTTTACTGATGGTC CTGGTGGTGCTCAGCTGCAAGTCAAGCTGCTCTCTGGGCTGTGATCTCCCTGAGACC CACAGCCTGGATAACAGGAGGACCTTGATGCTCCTGGCACAAATGAGCAGAATCTCT CCTTCCTCCTGTCTGATGGACAGACATGACTTTGGATTTCCTCCAGGAGGAGTTTGA TGGCAACCAGTTCCAGAAGGCTCCAGCCATCTCTGTCTCCATGAGCTGATCCAGCAG ATCTTCAACCTCTTTACCACAAAAGATTCATCTGCTGCTTGGGATGAGGACCTCCTA GACAAATTCTGCACCGAACTCTACCAGCAGCTGAATGACTTGGAAGCCTGTGTGATG CAGGAGGAGAGGGTGGGAGAACTCCCCTGATGAATGCGGACTCCATCTTGGCTGTG AAGAAATACTTCCGAAGAATCACTCTCTATCTGACAGAGAAGAAATACAGCCCTTGT GCCTGGGAGGTTGTCAGAGCAGAAATCATGAGATCCTCTCTTTATCAACAACTTGC AAGAAAGATTAAAGGAGGAAGGAATAA, TGTGATCTCCCTGAGACCCACAGCCTGGA TAACAGGAGGACCTTGATGCTCCTGGCACAAATGAGCAGAATCTCTCCTTCCTCCTG TCTGATGGACAGACATGACTTTGGATTTCCTCCAGGAGGAGTTTGATGGCAACCAGTT CCAGAAGGCTCCAGCCATCTCTGTCTCCATGAGCTGATCCAGCAGATCTTCAACCT

CTTTACCACAAAAGATTTCATCTGCTGCTTGGGATGAGGACCTCCTAGACAAATTCTG
 CACCGAACTCTACCAGCAGCTGAATGACTTGAAGCCTGTGTGATGCAGGAGGAGAG
 GGTGGGAGAACTCCCCTGATGAATGCCGACTCCATCTTGGCTGTGAAGAAATACTT
 CCGAAGAATCACTCTCTATCTGACAGAGAAGAAATACAGCCCTTGTGCCTGGGAGGT
 TGTGAGAGCAGAAATCATGAGATCCCTCTCTTTATCAACAACTTGCAAGAAAGATT
 AAGGAGGAAGGAATAA and fragments and derivatives thereof,
 said fragments and derivatives coding for polypeptides
 displaying an immunological or biological activity of
 IFN- α .

6. A DNA sequence according to any one of
 claims 1 to 4 selected from the group consisting of DNA
 sequences of the formula: TTAAGTGGTGGCCCTCCTGGTGCTCAGCTGC
 AAGTCAAGCTGCTCTGTGGGCTGTGATCTGCCTCAAACCCACAGCCTGGGTAGCAGG
 AGGACCTTGATGCTCCTGGCACAGATGAGGAGAATCTCTCTTTTCTCCTGCTTGAAG
 GACAGACATGACTTTGGATTTCCCCAGGAGGAGTTTGGCAACCAGTTCCAAAAGGCT
 GAAACCATCCCTGTCCTCCATGAGATGATCCAGCAGATCTTCAATCTCTTCAGCACA
 AAGGACTCATCTGCTGCTTGGGATGAGACCCTCCTAGACAAATTCTACACTGAACTC
 TACCAGCAGCTGAATGACCTGGAAGCCTGTGTGATACAGGGGGTGGGGGTGACAGAG
 ACTCCCCTGATGAAGGAGGACTCCATTCTGGCTGTGAGGAAATACTTCCAAAGAATC
 ACTCTCTATCTGAAAGAGAAGAAATACAGCCCTTGTGCCTGGGAGGTTGTCAGAGCA
 GAAATCATGAGATCTTTTCTTTGTCAACAACTTGCAAGAAAGTTTAAGAAGTAAG
 GAATGA, TGTGATCTGCCTCAAACCCACAGCCTGGGTAGCAGGAGGACCTTGATGC
 TCCTGGCACAGATGAGGAGAATCTCTCTTTTCTCCTGCTTGAAGGACAGACATGACT
 TTGGATTTCCCCAGGAGGAGTTTGGCAACCAGTTCCAAAAGGCTGAAACCATCCCTG
 TCCTCCATGAGATGATCCAGCAGATCTTCAATCTCTTCAGCACAAGGACTCATCTG
 CTGCTTGGGATGAGACCCTCCTAGACAAATTCTACACTGAACTCTACCAGCAGCTGA
 ATGACCTGGAAGCCTGTGTGATACAGGGGGTGGGGGTGACAGAGACTCCCCTGATGA
 AGGAGGACTCCATTCTGGCTGTGAGGAAATACTTCCAAAGAATCACTCTCTATCTGA
 AAGAGAAGAAATACAGCCCTTGTGCCTGGGAGGTTGTCAGAGCAGAAATCATGAGAT
 CTTTTTCTTTGTCAACAACTTGCAAGAAAGTTTAAGAAGTAAGGAATGA and
 fragments and derivatives thereof, said fragments and
 derivatives coding for polypeptides displaying an immunol-
 ogical or biological activity of IFN- α .

7. A DNA sequence according to any one of
 claims 1 to 4 selected from the group consisting of DNA
 sequences of the formula: ATGGCCCTGTCCTTTTCTTTACTGATGGCC

GTGCTGGTGCTCAGCTACAAATCCATCTGTTCTCTGGGCTGTGATCTGCCTCAGACC
CACAGCCTGGGTAATAGGAGGACCTTGATACTCCTGCAACAAATGGGAAGAATCTCT
CATTTCTCCTGCCTGAAGGACAGACATGATTTCCGATTCCCCGAGGAGGAGTTTGAT
GGCCACCAGTTCCAGAAGACTCAAGCCATCTCTGTCCTCCATGAGATGATCCAGCAG
ACCTTCAATCTCTTCAGCACAGAGGACTCATCTGCTGCTTGGGAACAGAGCCTCCTA
GAAAAATTTTCCACTGAACTTTACCAGCAACTGAATGACCTGGAAGCATGTGTGATA
CAGGAGGTTGGGGTGGGAAGAGACTCCCCTGATGAATGTGGACTCCATCCTGGCTGTG
AGGAAATACTTCCAAAGAATCACTCTTTATCTAACAGAGAGAAGAAATACAGCCCTT
GTGCCTGGGAGGTTGTAAAAAAGATTAAGGAGGAAGGATTGA, TGTGATCTGCCTC
AGACCCACAGCCTGGGTAATAGGAGGACCTTGATACTCCTGCAAGAAATGGGAAGAA
TCTCTCATTTCTCCTGCCTGAAGGACAGACATGATTTCCGATTCCCCGAGGAGGAGT
TTGATGGCCACCAGTTCCAGAAGACTCAAGCCATCTCTGTCCTCCATGAGATGATCC
AGCAGACCTTCAATCTCTTCAGCACAGAGGACTCATCTGCTGCTTGGGAACAGAGCC
TCCTAGAAAAATTTTCCACTGAACTTTACCAGCAACTGAATGACCTGGAAGCATGTG
TGATACAGGAGGTTGGGGTGGGAAGAGACTCCCCTGATGAATCTGGACTCCATCCTGG
CTGTGAGGAAATACTTCCAAAGAATCACTCTTTATCTAACAGAGAAGAAATACAGCC
CTTGTGCCTGGGAGGTTGTCAGAGCAGAAATCATGAGATCCCTCTCGTTTTCAACAA
ACTTGCAAAAAAGATTAAGGAGGAAGGATTGA and fragments and deriva-
tives thereof, said fragments and derivatives coding for
polypeptides displaying an immunological or biological
activity of IFN- α .

8. A recombinant DNA molecule comprising a
DNA sequence according to any one of the preceeding
claims.

9. The recombinant DNA molecule according to
claim 8, wherein said DNA sequence is operatively linked
to an expression control sequence.

10. A recombinant DNA molecule according to
claim 9, wherein the expression control sequence is
selected from the group consisting of a lac system, a
 β -lac system, a trp system, major operator and promotor
regions of phage λ , the control region of fd coat protein,
and other sequences which control the expression of genes
of prokaryotic or eukaryotic cells and their viruses.

11. A recombinant DNA molecule according to
claim 9 or 10 selected from the group consisting of
C8-IFN- α 1, C8-IFN- α 2, LAC-AUG(α 2) and β -lac-AUG(α 2).

12. A host transformed with at least one recombinant DNA molecule according to any one of claims 8 to 11.

13. The host of claim 12 selected from the group consisting of strains of E. coli, Pseudomonas, Bacillus subtilis, Bacillus stearothermophilus, other bacilli, yeasts, other fungi, mouse or other animal or plant hosts and human tissue cells.

14. The transformed host according to claim 12 or 13 selected from the group consisting of E. coli HB101 (Z-pBR322(Pst)/HcIF-4c), E. coli HB101 (Z-pBR322(Pst)/HcIF-2h), E. coli HB101 (Z-pBR322(Pst)/HcIF-SN35), E. coli HB101 (Z-pBR322(Pst)/HcIF-SN42), and E. coli HB101 (Z-pKT287(Pst)/HcIF-2h-AH6).

15. The transformed host according to claim 12 or 13 selected from the group consisting of E. coli HB101 (Z-pBR322(Pst)/HcIF-II-206) and E. coli HB101 (Z-pBR322(Pst)/HcIF-SN35-AHL6).

16. The transformed host according to claim 12 or 13 selected from the group consisting of HchrIF-A, HchrIF-B, HchrIF-C, HchrIF-D, HchrIF-E, HchrIF-F, HchrIF-G, HchrIF-H, HchrIF-I, and HchrIF-J.

17. The transformed host according to claim 12 or 13 selected from the group consisting of E. coli DS410 (C8-IFN- α 1), E. coli DS410 (C8-IFN- α 2), E. coli DS410 (LAC-AUG(α 2)), E. coli DS410 HB101 (β lac-AUG(α 2)) and Mouse 3T3 (polynoma-Hif-chr35).

18. The transformed host according to claim 12 or 13 selected from the group consisting of HchrIF-K, HchrIF-L, HchrIF-M, HchrIF-N, HchrIF-O, HchrIF-P, HchrIF-Q and hosts transformed with Hif-chr19 and Hif-chr27.

19. A polypeptide or fragments and derivatives thereof displaying an immunological or biological activity of human leukocyte interferon produced by the transformed host according to any one of claims 12 to 18.

20. A polypeptide that it is coded for by a DNA sequence according to any one of claims 1 to 7.

21. A polypeptide or fragments and derivatives thereof according to claim 19 or 20 selected from the group consisting of IFN- α 1, IFN- α 2, IFN- α 4a and IFN- α 4b.

22. A polypeptide or fragments and derivatives thereof according to any one of claims 19 to 21 selected from the group consisting of polypeptides of the formula:
 METALASERPROPHEALALEULEUMETVALLEUVALVALLEUSERCYSLYSSERSER
 CYSSERLEUGLYCYSASPLEUPROGLUTHRHISERLEUASPASNARGARGTHRLEU
 METLEULEUALAGLNMETSERARGILESERPROSERSERCYSLEUMETASPARGHIS
 ASPPHEGLYPHEPROGLNGLUGLUPHEASPGLYASNGLNPNHEGLNLYSALAPROALA
 ILESERVALLEUHIISGLULEUILEGLNGLNILEPHEASNLEUPHETHRTHRLYSASP
 SERSERALAALATRPASPGLUASPLEULEUASPLYSPHECYSTHRGLULEUTYRGLN
 GLNLEUASNASPLEUGLUALACYSVALMETGLNGLUGLUARGVALGLYGLUTHRPRO
 LEUMETASNALAASPSEIRILELEUALAVALLYSLYSTYRPHEARGARGILETHRLEU
 TYRLEUTHRGLULYSLYSTYRSERPROCYSALATRPGLUVALVALARGALAGLUILE
 METARGSERLEUSERLEUSERTHRASNLEUGLNGLUARGLEUARGARGLYSGLU,
 CYSASPLEUPROGLUTHRHISERLEUASPASNARGARGTHRLEUMETLEULEUALA
 GLNMETSERARGILESERPROSERSERCYSLEUMETASPARGHISASPPHEGLYPHE
 PROGLNGLUGLUPHEASPGLYASNGLNPNHEGLNLYSALAPROALAILESERVALLEU
 HISGLULEUILEGLNGLNILEPHEASNLEUPHETHRTHRLYSASPSEIRALAALA
 TRPASPGLUASPLEULEUASPLYSPHECYSTHRGLULEUTYRGLNGLNLEUASNASP
 LEUGLUALACYSVALMETGLNGLUGLUARGVALGLYGLUTHRPROLEUMETASNALA
 ASPSEIRILELEUALAVALLYSLYSTYRPHEARGARGILETHRLEUTYRLEUTHRGLU
 LYSTYSTYRSERPROCYSALATRPGLUVALVALARGALAGLUILEMETARGSERLEU
 SERLEUSERTHRASNLEUGLNGLUARGLEUARGARGLYSGLU, and polypeptides
 from whatever source obtained related to any of the
 foregoing polypeptides by mutation, including single or
 multiple, base substitutions, deletions, insertions and
 inversions of the DNA sequences which code for them.

23. A polypeptide or fragments and derivatives thereof according to any one of claims 19 to 21 selected from the group consisting of polypeptides of the formula:
 LeuLeuValAlaLeuLeuValLeuSerCysLysSerSerCysSerValGlyCysAsp
 LeuProGlnThrHisSerLeuGlySerArgArgThrLeuMetLeuLeuAlaGlnMet
 ArgArgIleSerLeuPheSerCysLeuLysAspArgHisAspPheGlyPheProGln
 GluGluPheGlyAsnGlnPheGlnLysAlaGluThrIleProValLeuHisGluMet
 IleGlnGlnIlePheAsnLeuPheSerThrLysAspSerSerAlaAlaTrpAspGlu

ThrLeuLeuAspLysPheTyrThrGluLeuTyrGlnGlnLeuAsnAspLeuGluAla
 CysValIleGlnGlyValGlyValThrGluThrProLeuMetLysGluAspSerIle
 LeuAlaValArgLysTyrPheGlnArgIleThrLeuTyrLeuLysGluLysLysTyr
 SerProCysAlaTrpGluValValArgAlaGluIleMetArgSerPheSerLeuSer
 ThrAsnLeuGlnGluSerLeuArgSerLysGlu, CysAspLeuProGlnThrHis
 SerLeuGlySerArgArgThrLeuMetLeuLeuAlaGlnMetArgArgIleSerLeu
 PheSerCysLeuLysAspArgHisAspPheGlyPheProGlnGluGluPheGlyAsn
 GlnPheGlnLysAlaGluThrIleProValLeuHisGluMetIleGlnGlnIlePhe
 AsnLeuPheSerThrLysAspSerSerAlaAlaTrpAspGluThrLeuLeuAspLys
 PheTyrThrGluLeuTyrGlnGlnLeuAsnAspLeuGluAlaCysValIleGlnGly
 ValGlyValThrGluThrProLeuMetLysGluAspSerIleLeuAlaValArgLys
 TyrPheGlnArgIleThrLeuTyrLeuLysGluLysLysTyrSerProCysAlaTrp
 GluValValArgAlaGluIleMetArgSerPheSerLeuSerThrAsnLeuGlnGlu
 SerLeuArgSerLysGlu, and polypeptides from whatever source
 obtained related to any of the foregoing polypeptides by
 mutation, including single or multiple, base substitutions,
 deletions, insertions and inversions of the DNA sequences
 which code for them.

24. A polypeptide or fragments and derivatives
 thereof according to any one of claims 19 to 21 selected
 from the group consisting of polypeptides of the formula:
 METALALEUSERPHERSERLEULEUMETALAVALLEUVALLEUSERTYRLYSSERILE
 CYSSERLEUGLYCYSASPLEUPROGLNTHRRHISERLEUGLYASNARGARGTHR
 LEUILEULEULEUGLNGLNMETGLYARGILESERHISPHERCYSLEULYSASPARG
 HISASPPHEGLYPHEPROGLUGLUGLUPHEASPGLYHISGLNPHEGLNLYSTHRLN
 ALAILESERVALLEUHSGLUMETILEGLNGLNTHRPHEASNLEUPHERSERTHRGLU
 ASPSERSERALAALATRPGLUGLNSERLEULEUGLULYSPHERSERTHRGLULEUTYR
 GLNGLNLEUASNASPLEUGLUALACYSVALILEGLNGLUVALGLYVALGLUGLUTHR
 PROLEUMETASNVALASPSERILELEUALAVALARGLYSTYRPHGLNARGILETHR
 LEUTYRLEUTHRGLULYSLYSTYRSEPROCYALATRPGLUVALVALARGALAGLU
 ILEMETARGSERLEUSERPHERSERTHRASNLEUGLNLYSARGLEUARGARGLYSASP,
 CYSASPLEUPROGLNTHRRHISERLEUGLYASNARGARGTHRLEUILEULEULEUGLN
 GLNMETGLYARGILESERHISPHERCYSLEULYSASPARGHISASPPHEGLYPHE
 PROGLUGLUGLUPHEASPGLYHISGLNPHEGLNLYSTHRLNALAILESERVALLEU
 HISGLUMETILEGLNGLNTHRPHEASNSEUPHERSERTHRGLUASPSESERALAALA
 TRPGLUGLNSERLEULEUGLULYSPHERSERTHRGLULEUTYRGLNGLNLEUASNAS
 LEUGLUALACYSVALILEGLNGLUVALGLYVALGLUGLUTHRPROLEUMETASNVAL

ASPSERILELEUALAVALARGLYSTYRPHLEGLNARGILETHRLEUTYRLEUTHERGLU
LYSLYSTYRSERPROCYSALATRPGLUVALVALARGALAGLUILEMETARGSERLEU
SERPHESERTHRASNLEUGLNLYSARGLEUARGARGLYSASP, and polypeptides
from whatever source obtained related to any of the
foregoing polypeptides by mutation, including single or
multiple, base substitutions, deletions, insertions and
inversions of the DNA sequences which code for them.

25. A method for producing a recombinant DNA molecule comprising the step of introducing into a cloning vehicle a DNA sequence according to any one of claims 1-7.

26. The method according to claim 25 comprising the additional step of introducing into said cloning vehicle an expression control sequence according to claim 10, said expression control sequence being introduced into said cloning vehicle so as to control and to regulate the expression of said DNA sequence.

27. A method for transforming a host comprising the step of introducing into a host a recombinant DNA molecule according to any one of claims 8 to 11.

28. A method for producing a polypeptide displaying an immunological or biological activity of human leukocyte interferon, comprising the steps of transforming an appropriate host with a recombinant DNA molecule according to claim 10 or 11; culturing said host; and collecting said polypeptide.

29. The method according to claim 28, wherein the host is selected from the group consisting of strains of E. coli, Pseudomonas, Bacillus subtilis, Bacillus stearothermophilus, other bacilli, yeasts, fungi, animal or plant hosts, and human tissue cells.

30. A method for producing a polypeptide displaying an immunological or biological activity of human leukocyte interferon comprising the steps of culturing a host transformed by a recombinant DNA molecule according to claim 10 or 11 and collecting said polypeptide.

31. A process for selecting a DNA sequence coding for a polypeptide displaying an immunological or biological activity of HuIFN α from a group of DNA sequences

comprising the step of determining which of said DNA sequences hybridize to the DNA sequences according to any one of claims 1-7.

32. The process of claim 31 wherein the DNA sequence screened is selected from the group consisting of DNA sequences from natural sources, synthetic DNA sequences, DNA sequences from recombinant DNA molecules and DNA sequences which are a combination of any of the foregoing DNA sequences.

33. A composition for treating human viruses or treating human cancers or tumors which comprises at least one polypeptide selected from the group consisting of the polypeptides according to any one of claims 19 to 24.

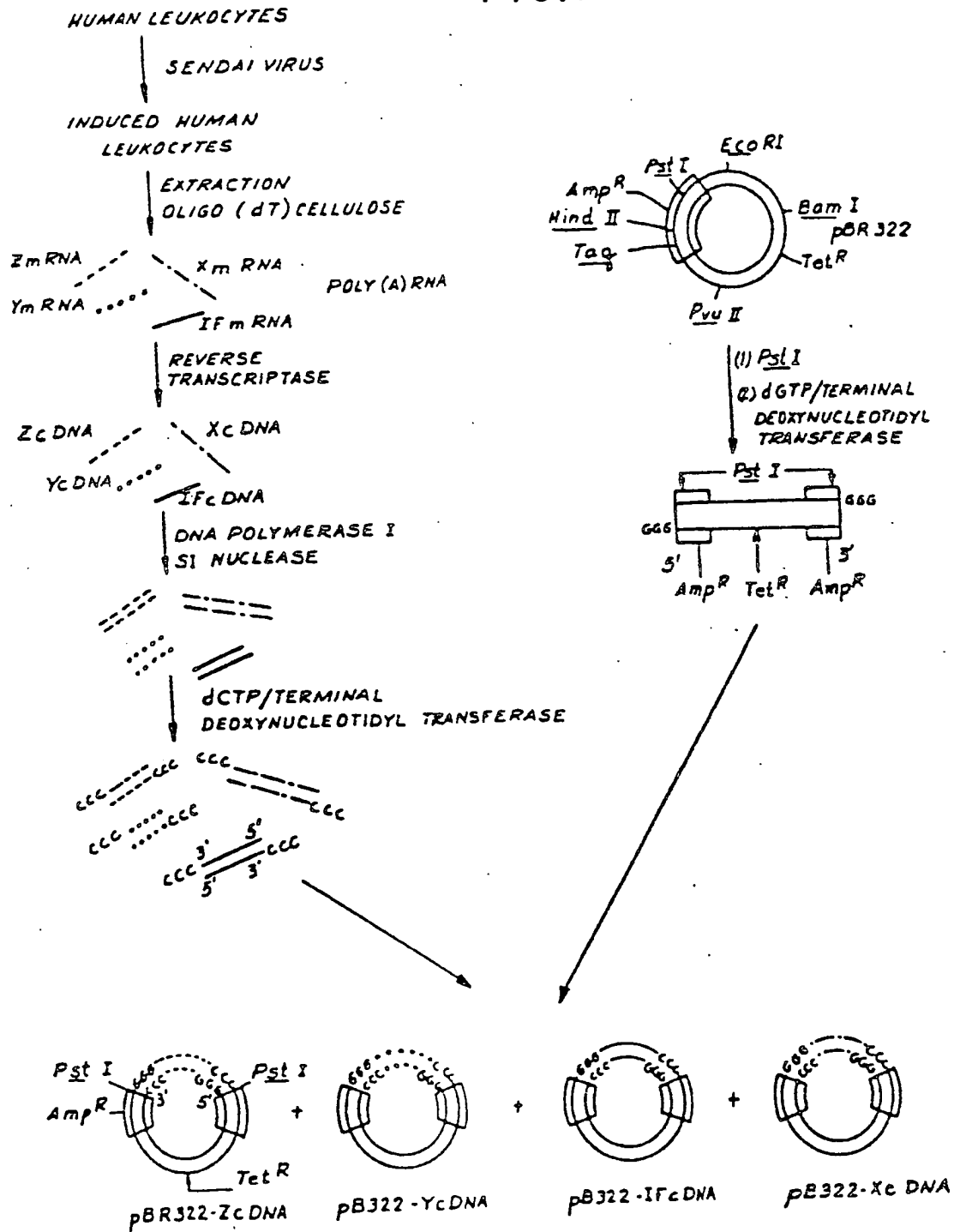
34. A composition for treating bovine viral infections which comprises at least one polypeptide selected from the group consisting of the polypeptides according to any one of claims 19 to 24.

35. A method for treating human viruses or treating human cancers or tumors which comprises administering to said humans in a pharmaceutically acceptable manner an effective amount of a composition according to claim 33.

36. A method for treating bovine viral infections which comprises administering to said animals in a pharmaceutically acceptable manner an effective amount of a composition according to claim 34.

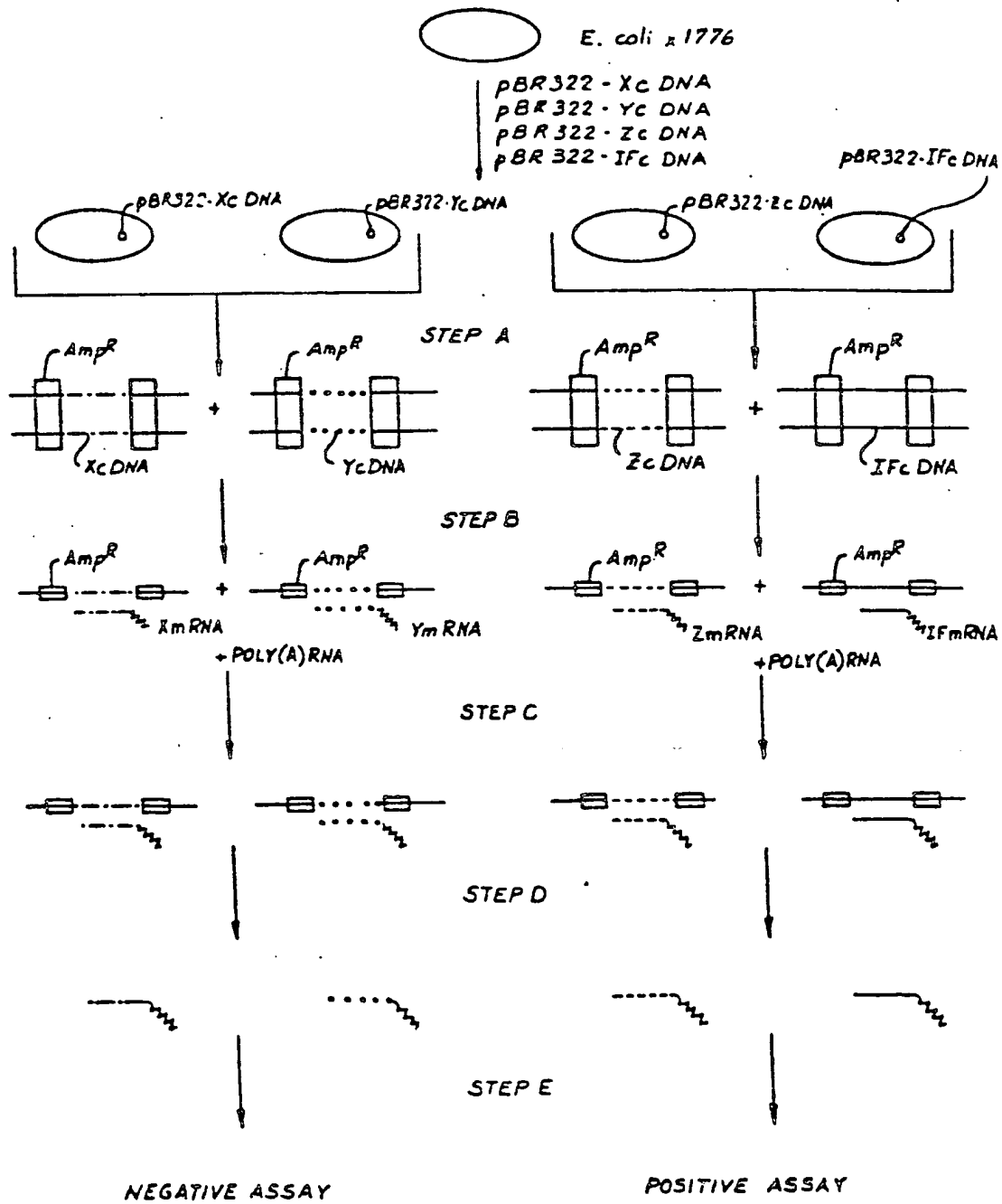
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FIG. 1



2 / 3 1

FIG. 2



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FIG. 3

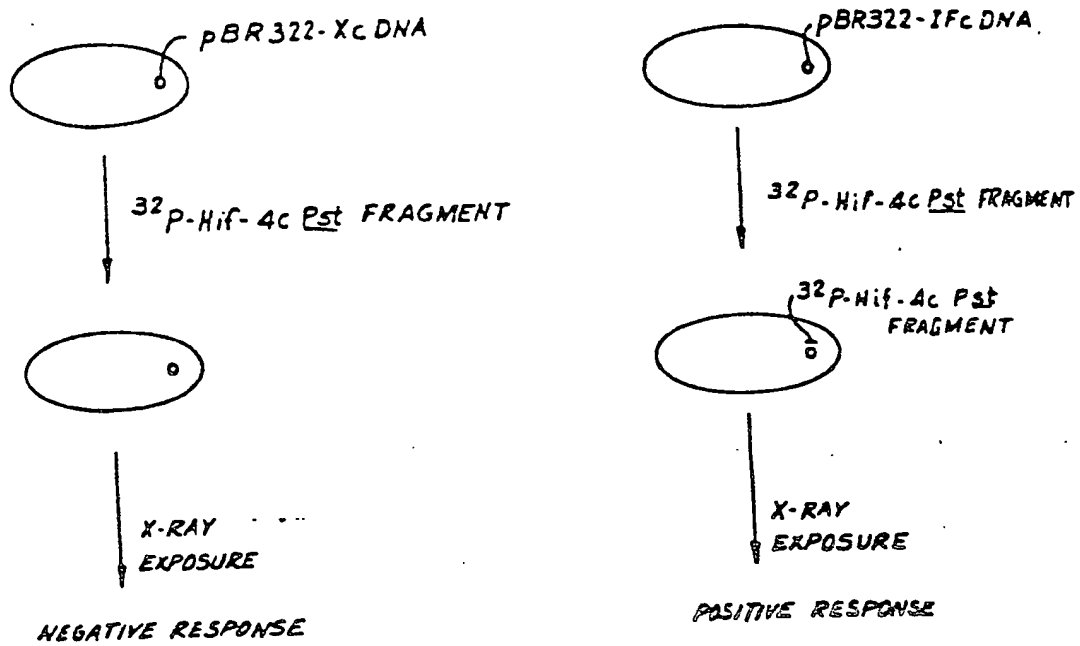
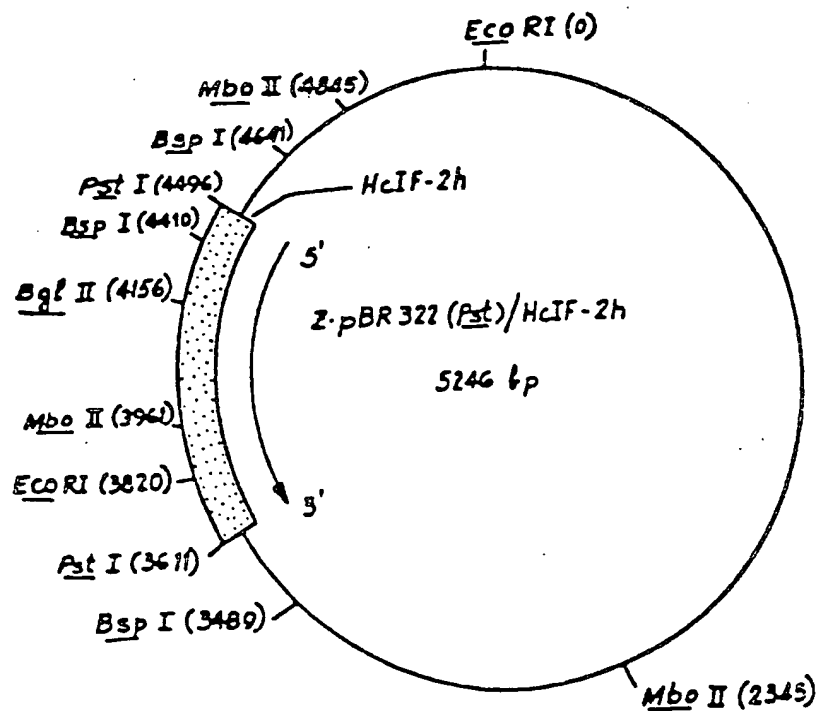
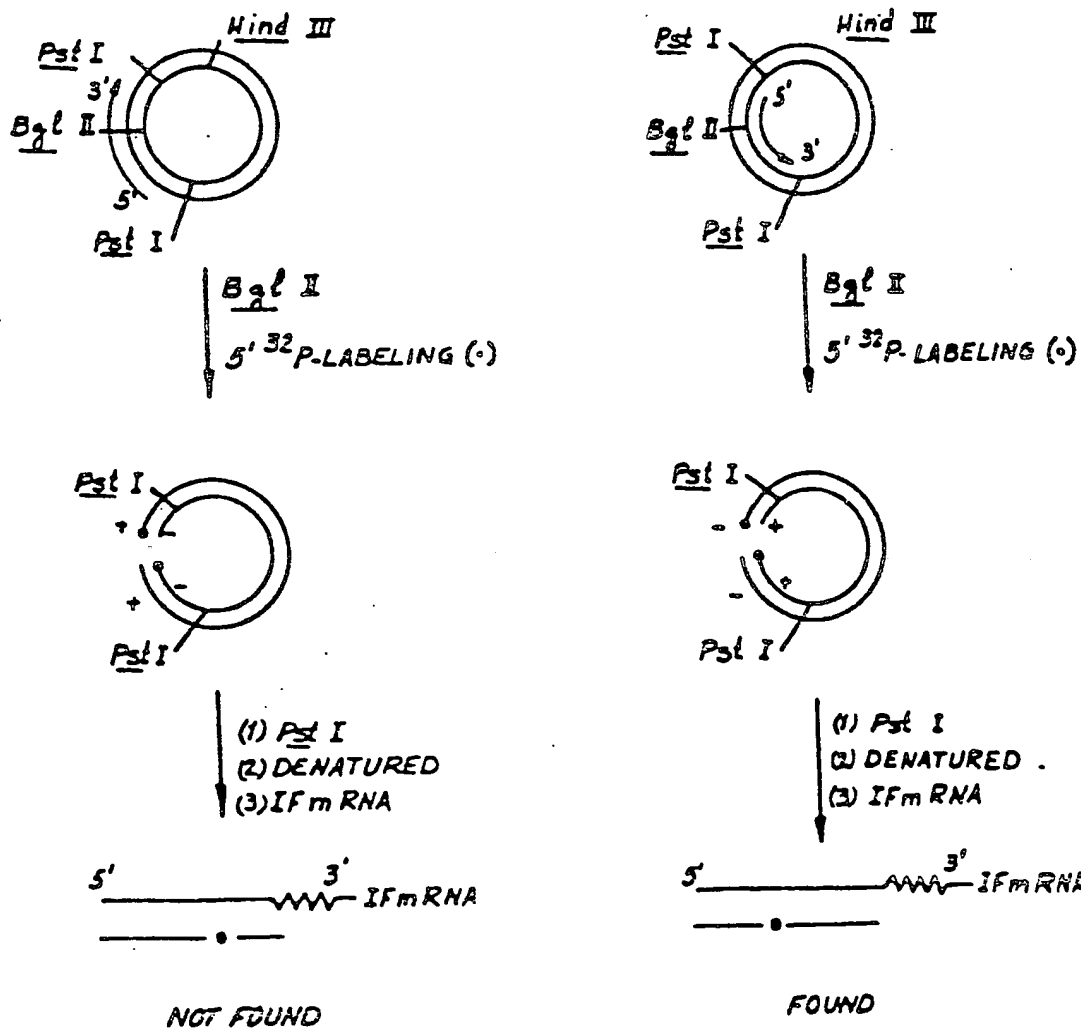


FIG. 4



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FIG. 5



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10
 20
 MetSerIleGlnHisPheArgValAlaLeuIleProPhePheAlaAlaPheCysLeuProValPheAlaHisProGluThr
 pBR322 ATGAGTATTCACACATTTCCGTGTCGCCCTTATCCCTTTTTTGGGGCATTTTGCCTTCCIGTTTTTGTCTACCCAGAAACG
 181
 29
 LeuVal ProAlaAlaMet
 CTGGTG...CTGCAGCAATG...
Pat

24
 MetSerIleGlnHisPheArgValAlaLeuIleProPhePheAlaAlaPheCysLeuProValPheAlaHisArgCysSerAsn
 pKT279 ATGAGTATTCACACATTTCCGTGTCGCCCTTATCCCTTTTTTGGGGCATTTTGCCTTCCIGTTTTTGTCTACCCGCTGCAGCAATG...
Pat

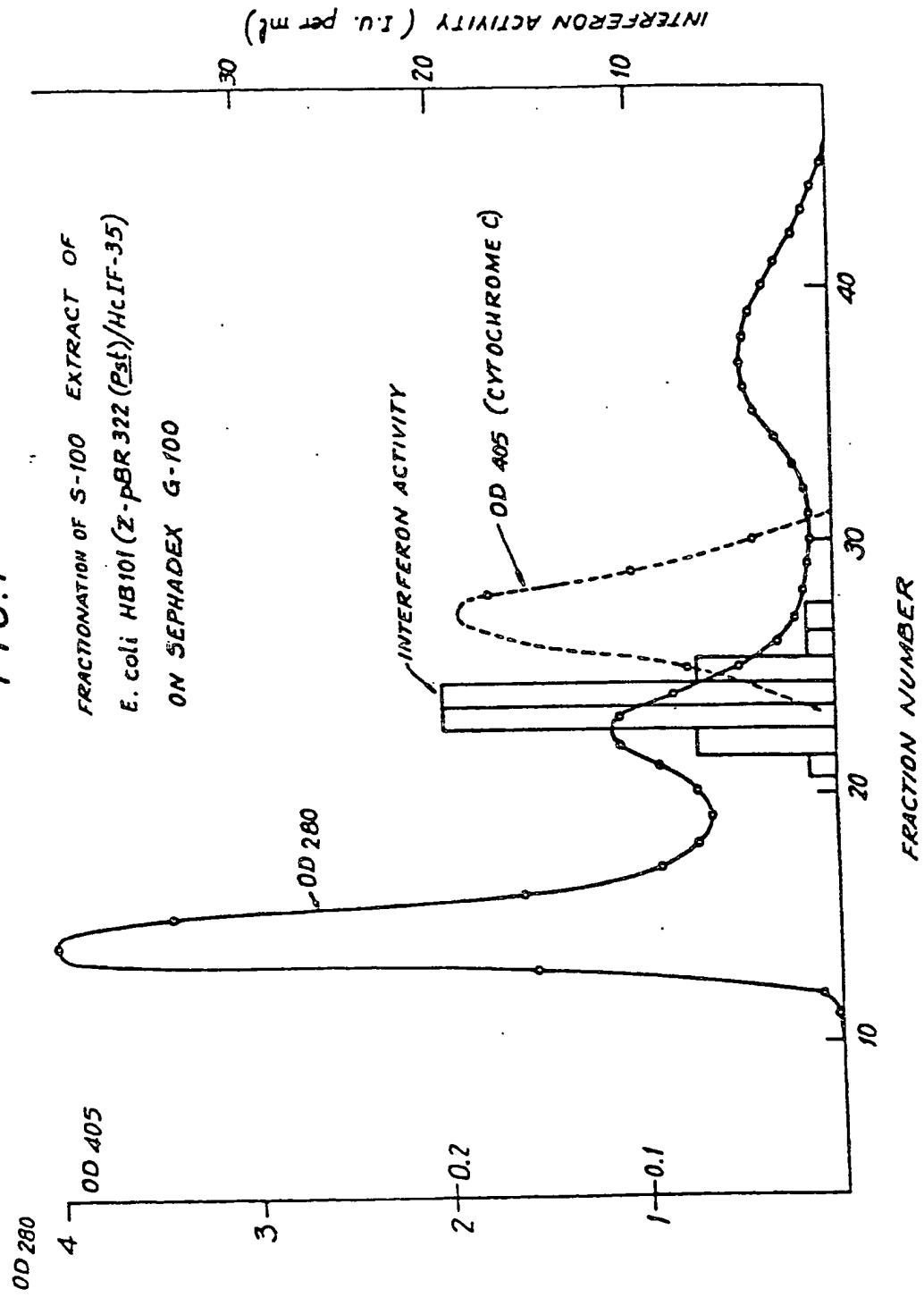
25
 MetSerIleGlnHisPheArgValAlaLeuIleProPhePheAlaAlaPheCysLeuProValPheAlaHisProLeuGlnGln
 pKT280 ATGAGTATTCACACATTTCCGTGTCGCCCTTATCCCTTTTTTGGGGCATTTTGCCTTCCIGTTTTTGTCTACCCGCTGCAGCAATG...
Pat

27
 MetSerIleGlnHisPheArgValAlaLeuIleProPhePheAlaAlaPheCysLeuProValPheAlaHisProGluThr
 pKT287 ATGAGTATTCACACATTTCCGTGTCGCCCTTATCCCTTTTTTGGGGCATTTTGCCTTCCIGTTTTTGTCTACCCAGAAACG...
 AlaAlaAlaMet
 GCTGCAGCAATG...
Pat

FIG 6

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FIG. 7



1 10 20 30 40 50 60 70 80
 S1 S10
 G₂₃CTCTAGGTTCAGAGTCACCCATCTCAGCAAGCCCAGAGTATCTGCAATATCTACGATGGCCTGCCCCTTGCCTTACTGATGGTCCCTG
 METALASERPROPHEALALEULEUMETVALLEU
 BspI
 EcoRII
 AValI

90 100 110 120 130 140 150 160 170
 S20 1 10
 VALVALLEUSERCYSLYSSERSEYSSERLEUGLYCYSASPLEUPROGLUTHRHISSELEUASPASNARGARGTHRLEUMETLEULEU
 GGGTGCTCAGCTGCAAGTCAAGCTGCTCTCTGGGCTGTGATCTCCCTGAGACCCACAGCCTGGATAACAGGAGGACCTTGATGCTCCCTG
 DdeI PvuII ALuI MboI DdeI EcoPI EcoRII AValI SfaNI EcoRII
 ALuI

20 30 40
 180 190 200 210 220 230 240 250 260
 ALAGLNMETSERARGILESERPROSEYSSERLEUMETASPARGHISASPPHEGLYPHEPROGLNGLUPHEASPGLYASNGLNPHE
 GCACAANTGAGCAGAATCTCTCCCTCCCTGCTGATGGACAGACATGACTTTGGATTTCCCCAGGAGGAGTTTGATGGCAACCAGTTC
 HINF
 EcoRII

FIG. 8

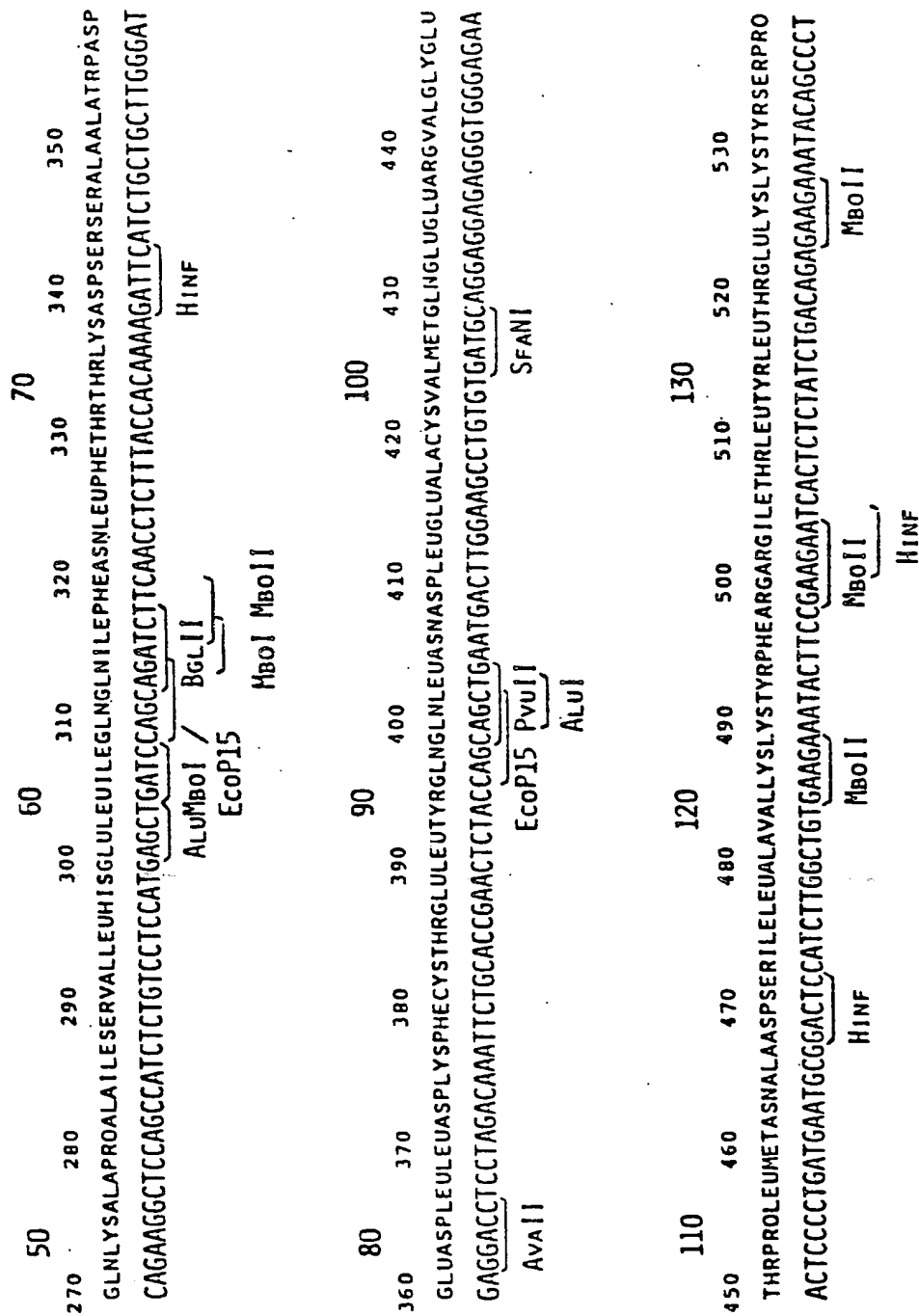


FIG. 9

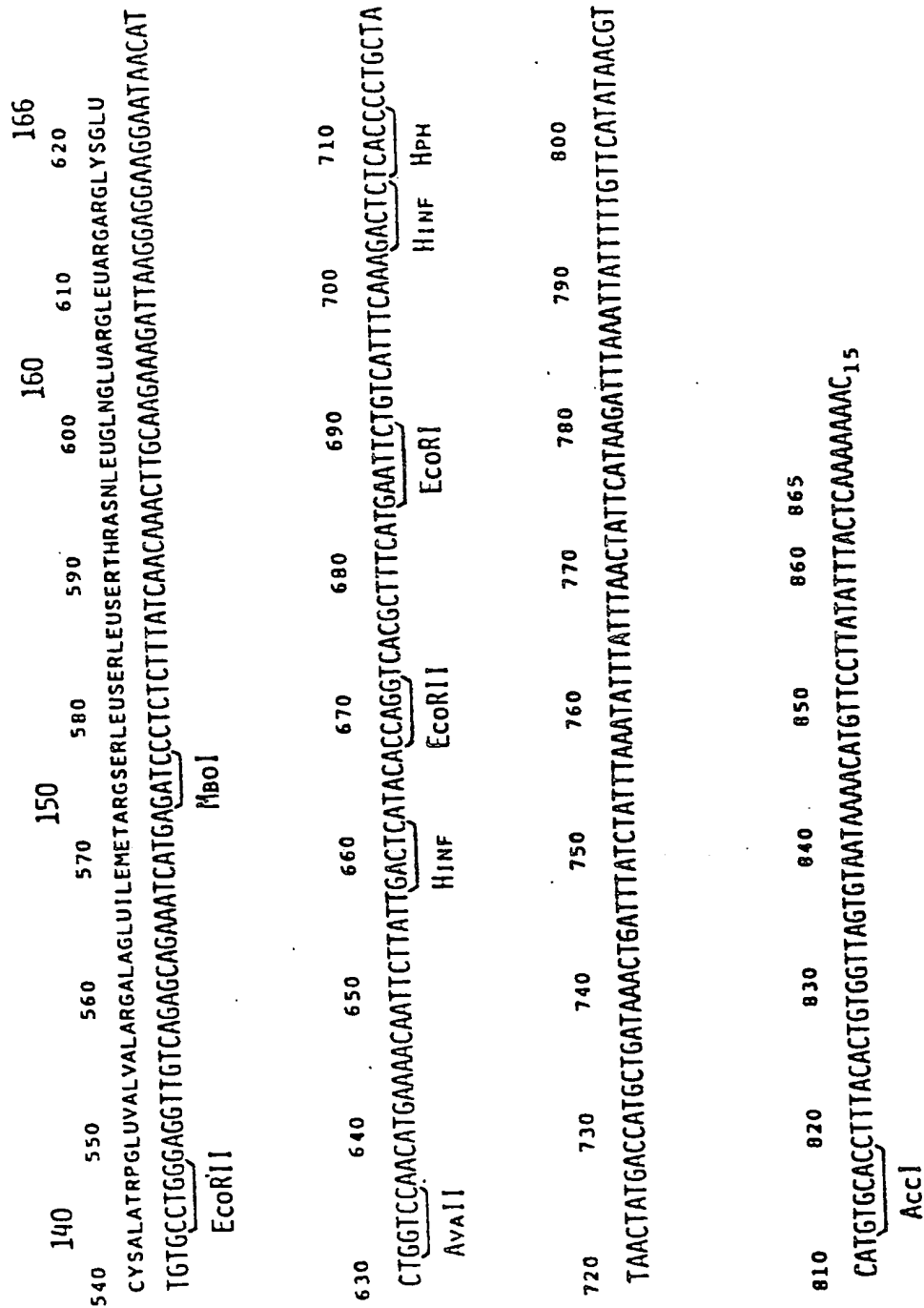
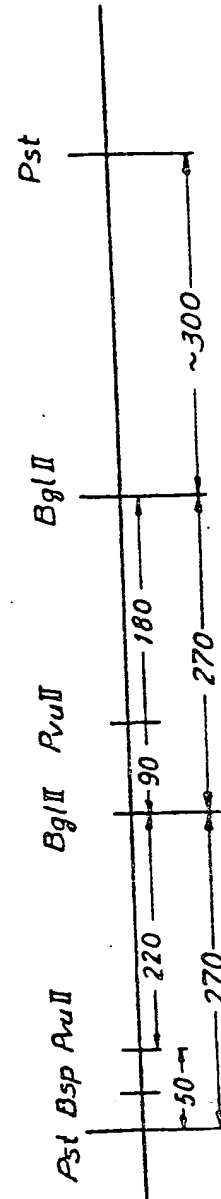
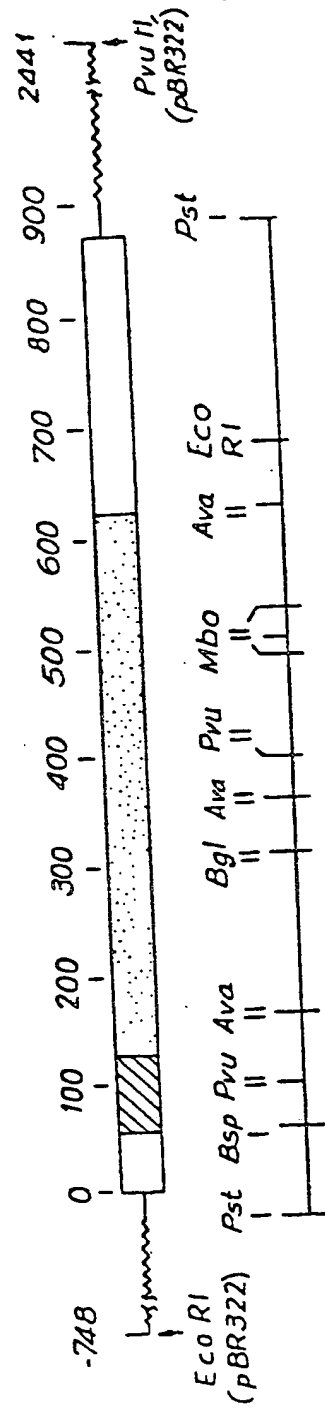


FIG.10

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FIG. 11



-50 -40
 6₁₃TTACTGGTGGCCCTG
 leu leu val ala leu

206(11)

-20

met ala ser pro phe ala leu leu met val leu
 6₂₃CTCTAGGTTACAGAGTCACCCATCTCAGCAAGCCAGAAAGTATCTGCAATATCTACGATGGCCTCGCCCTTTCCTTTACTGATGGTCCCTG
 -120 -110 -100 -90 -80 -70 -60 -50 -40

2H(1)

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-30 -20 -10 -1 1 10 20 30 40 50
 CTGGTGCCTCAGCTGCAAGTCAAGCTGCTCTGTGGGCTGTGATCTGCTCAAAACCCACAGGCTGGGTAGCAGGAGGACCTTGTGATGCTCCCTG
 leu val leu ser cys lys ser ser cys ser val gly cys asp leu pro gln thr his ser leu gly ser arg thr leu met leu leu

206(11)

10

-1 1

-10

val val leu ser cys lys ser ser cys ser leu gly cys asp leu pro glu thr his ser leu asp asn arg thr leu met leu leu
 GTGGTGCCTCAGCTGCAAGTCAAGCTGCTCTGTGGGCTGTGATCTGCTCAAAACCCACAGGCTGGGTAGCAGGAGGACCTTGTGATGCTCCCTG
 -30 -20 -10 -1 1 10 20 30 40 50

2H(1)

FIG. 12

GCACAGATGAGGAGAATCTCTTTTCCTCCCTGAGGACAGACAATGACATTGGATTCCCGAGGAGGAGTTT---GGCAACCAGATTTC
60 70 80 90 100 110 120 130 140
ALA GLN MET ARG ARG ILE SER LEU PHE SER CYS LEU LYS ASP ARG HIS ASP PHE GLY PHE PRO GLN GLU GLU PHE - GLY ASN GLN PHE
40
30
20
ALA GLN MET SER ARG ILE SER PRO SER SER CYS LEU MET ASP ARG HIS ASP PHE GLY PHE PRO GLN GLU GLU PHE ASP GLY ASN GLN PHE
GCACAAATGAGCAGAGAATCTCTCTTCTCCCTCCCTGATGGACAGACAATGACATTGGATTCCCGAGGAGGAGTTTGGTGGCAACCAGATTTC
60 70 80 90 100 110 120 130 140

206(11)

 $2\text{H}(\text{I})$

CAAAGGCTGAAACCATCCCTGTCCTCCATGAGATGATCCAGCAGATCTTCAATCTCTTCAGCACAAAGGACATCATCTGCTGCTGGGAT.
Gln Lys Ala Glu Thr Ile Pro Val Leu His Glu Met Ile Cln Cln Ile Phe Asn Leu Phe Ser Thr Lys Asp Ser Ser Ala Ala Trp Asp
50 60 70
Gln Lys Ala Pro Ala Ile Ser Val Leu His Glu Leu Ile Cln Cln Ile Phe Asn Leu Phe Thr Thr Lys Asp Ser Ser Ala Ala Trp Asp
50 60 70
CAGAAAGGCTCCAGCCATCTCTGTCCTCCATGAGCTGATCCAGCAGATCTTCAACCTCTTTACCCACAAAGATTCATCTGCTGCTGGGAT.
150 160 170 180 190 200 210 220 230

206(11)

 $2\text{H}^+(\text{I})$

FIG. 13

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205(11) 240 250 260 270 280 290 300 310 320
 GAGACCTTCCTAGACAAAATTCACACAGACCTACCAAGCTGAAAGCTGTTGATACAGGGGTGGGGTGACAGAG
 GCU THR LEU LEU ASP LYS PHE TYR THR GCU LEU TYR GUN GUN LEU ASN ASP LEU GCU ALA CYS VAL ILE GUN GLY VAL GLY VAL THR GCU
 80 90 100
 80 240 250 260 270 280 290 300 310 320
 GCU ASP LEU LEU ASP LYS PHE CYS THR GCU LEU TYR GUN GUN LEU ASN ASP LEU GCU ALA CYS VAL MET GUN GCU ARG VAL GLY GCU
 GAGACCTTCCTAGACAAAATTCACACAGACCTACCAAGCTGAAAGCTGTTGATGACAGGGGTGGGGTGACAGAA
 240 250 260 270 280 290 300 310 320

2H(1)

205(11) 330 340 350 360 370 380 390 400 410
 ACTCCCCGTGATGAAGGAGGACATCCATTCTGGCTGTGAGGAAATACATCCAAAGATCCTCTCTGAAAGAGAGAAATACAGCCCT
 THR PRO LEU MET LYS CYS ASP SER ILE LEU ALA VAL ARG LYS TYR PHE GUN ARG ILE THR LEU TYR LEU LYS GCU LYS TYR SER PRO
 110 120 130
 110 330 340 350 360 370 380 390 400 410
 THR PRO LEU MET ASN ALA ASP SER ILE LEU ALA VAL LYS LYS TYR PHE ARG ARG ILE THR LEU TYR LEU THR GCU LYS TYR SER PRO
 ACTCCCCGTGATGAAGGAGGACATCCATTCTGGCTGTGAGGAAATACATCCAAAGATCCTCTCTGAAAGAGAGAAATACAGCCCT
 330 340 350 360 370 380 390 400 410

2H(1)

FIG. 14

206(11)

CTGGTTCAACATGGAAATGATTTTCATTGATCGTATGCCAGCTCACCTTTTATGA--TC TGCCATTTC AAAGACTCATGTTTTCIGCTA

510 520 530 540 550 560 570 580

2H(1)

CTGGTCCAACATGAAAACAATTCCTTATTGACTCATACCAAGGTACGGCTTTCATGAATTC TGTCATTTC AAAGACTCTCACCCCTGCTA

510 520 530 540 550 560 570 580 590

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590 600 610 620 630 640 650 660 670 680
TGACCATGACACGATTAAATCTTTTCAMAIGTTTTAGGAGTATTAAATCAACATTGTATTGAGCTCTTAAGGCACCTAGTCCCTTACAGAG

206(11)

600 610 620 630 640 650 660 670 680
TAACTATGACCATGCTGATAAACTGATTATCTATTTAAATATTTTAACTATTTCATAAGATTAAATTATTTTGTTCATATAACGTC

2H(1)

690
GACCATGCTGAC₂₉

206(11)

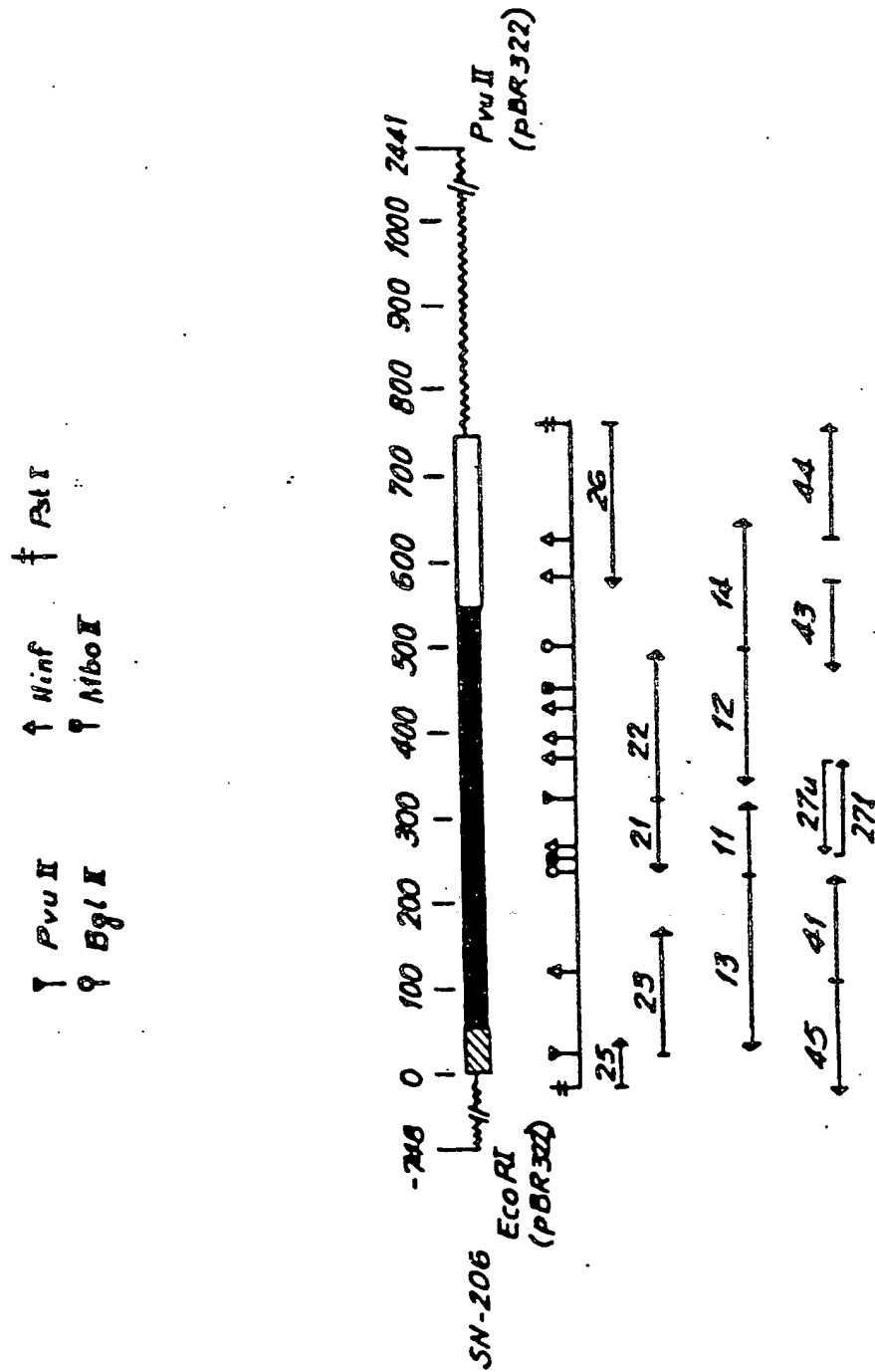
690 700 710 720 730 740
ATGTGCACCTTACACTGTGGTTAGTGTAAATAAACATGTTCCCTTATATTACTCAAAAAAAC₁₅

2H(1)

FIG.16

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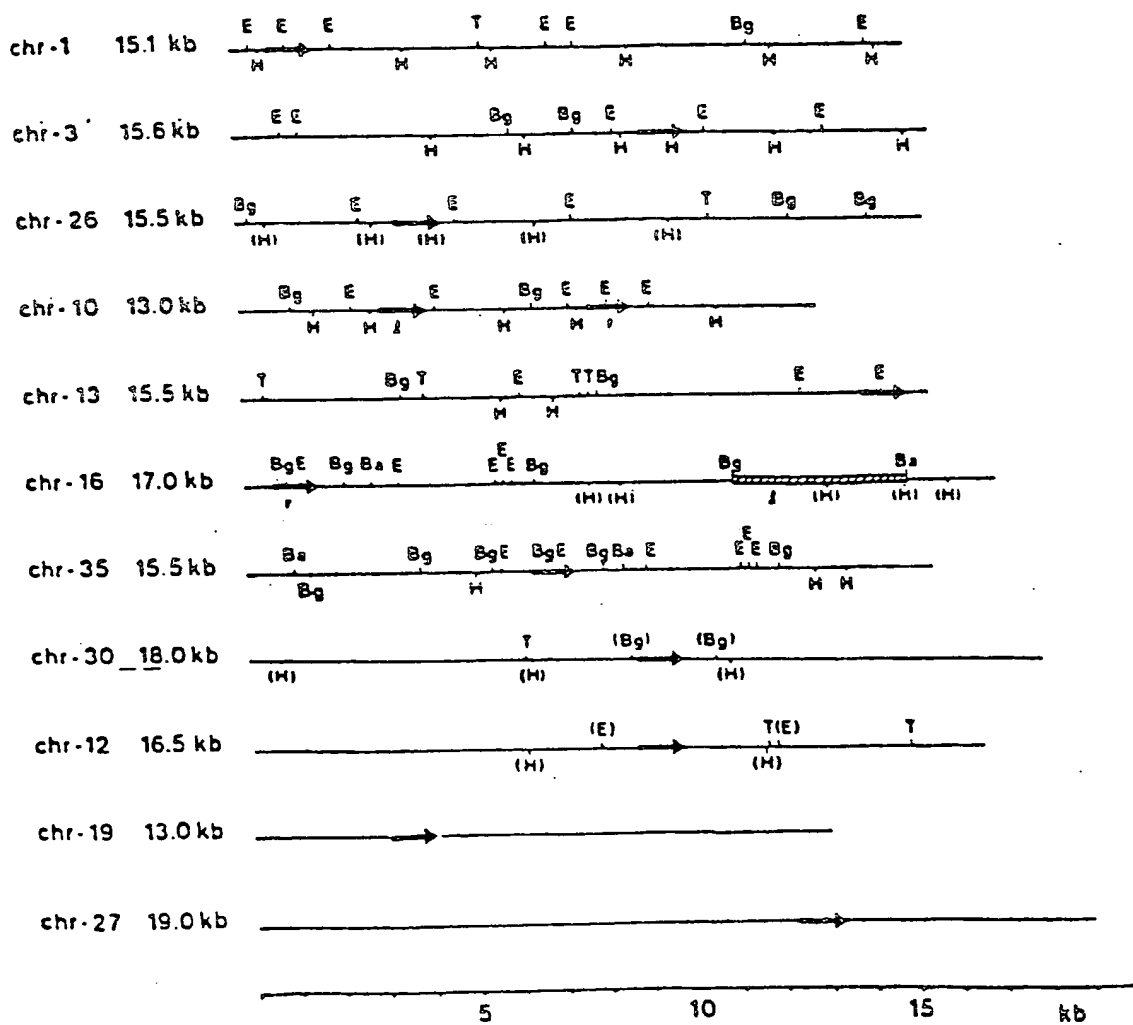
FIG. 17



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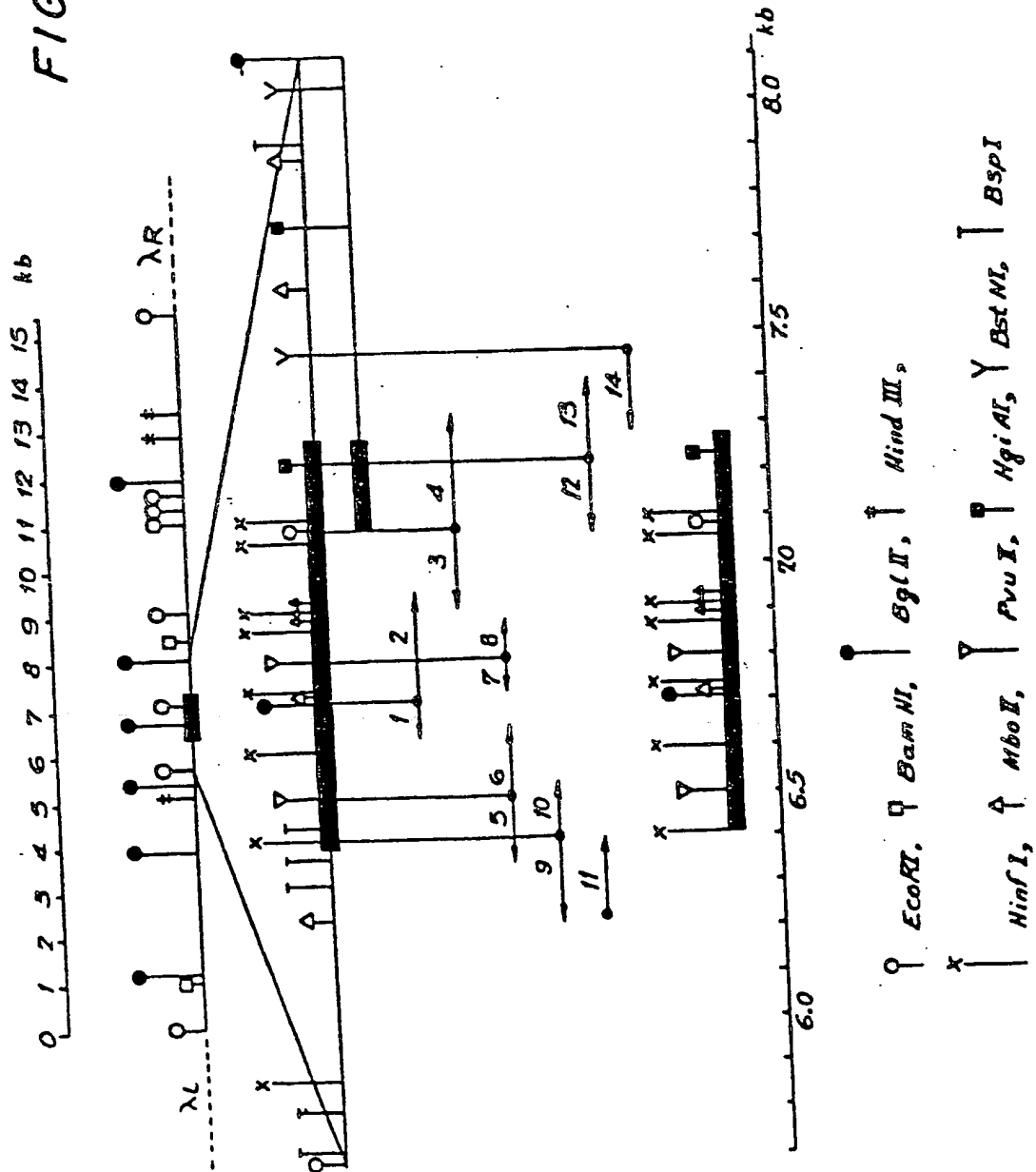
FIG. 18

PARTIAL RESTRICTION MAPS OF CLONED, IFN- α RELATED
CHROMOSOMAL DNA SEGMENTS



E : EcoRI, Ba : BamHI, Bg : BglIII, H : HincII, T : Tacl

FIG. 19



AAACAAACATTGAGAACACGGCTCTAACTCATGTAAGAGTGCATGAAGGAAAGCAAAACAGAAATG

GAAAGTGGCCCAAGCATTAAAGAAAGTGGAATCAGTATGTTCCCCTAATTAAAGGCATTTCAGGAAACCAAGGCCCTTCAGAGAACCTAGA
-140 -120 -80 -60
BspI BspI

S10
 S1
 +1
 20
 metalaseerpropheala leuleumetvalleu
 -20
 -40
 HINF
 G C C C A A G G T T C A G A G T C A C C C A T C T C A G C A A G C C C A G A G T A T C T G C A A T A T C T A C G A T G G C C T T G C C C T T G C T T A C T G A T G G T C C T G
 EcoRI
 BsPI
 AvalI

valvalleuueerovelyssereerovesserleuglvCYSASPLEUPROGLUTHRRHISSEERLEUASPASNARGARGTHRLEUMETLEULEU
GTGGTGCTCAGCTGCAAGTCAAGCTGCTCTCTGGGCTGTGATCTCCCTGAGACCCACAGCCTGGATACAGGAGGACCTTGATGCTCTG
S20 1 10 100 120
40 60 80
DDEI PVUI ALUI MBOI DDEI ECOPI ECORII AVAII SFANI ECORII
ALUI

FIG. 20

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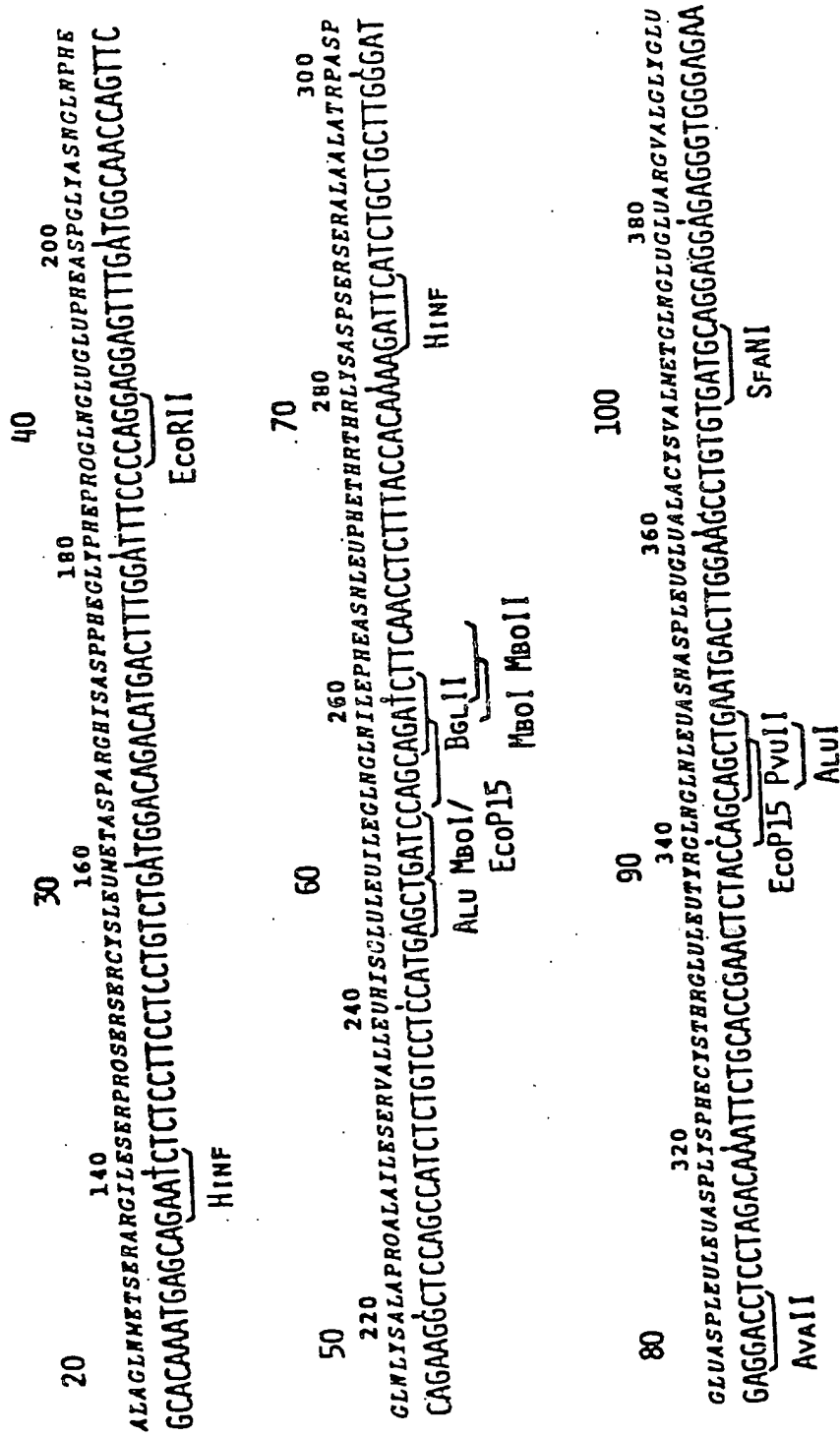


FIG. 21

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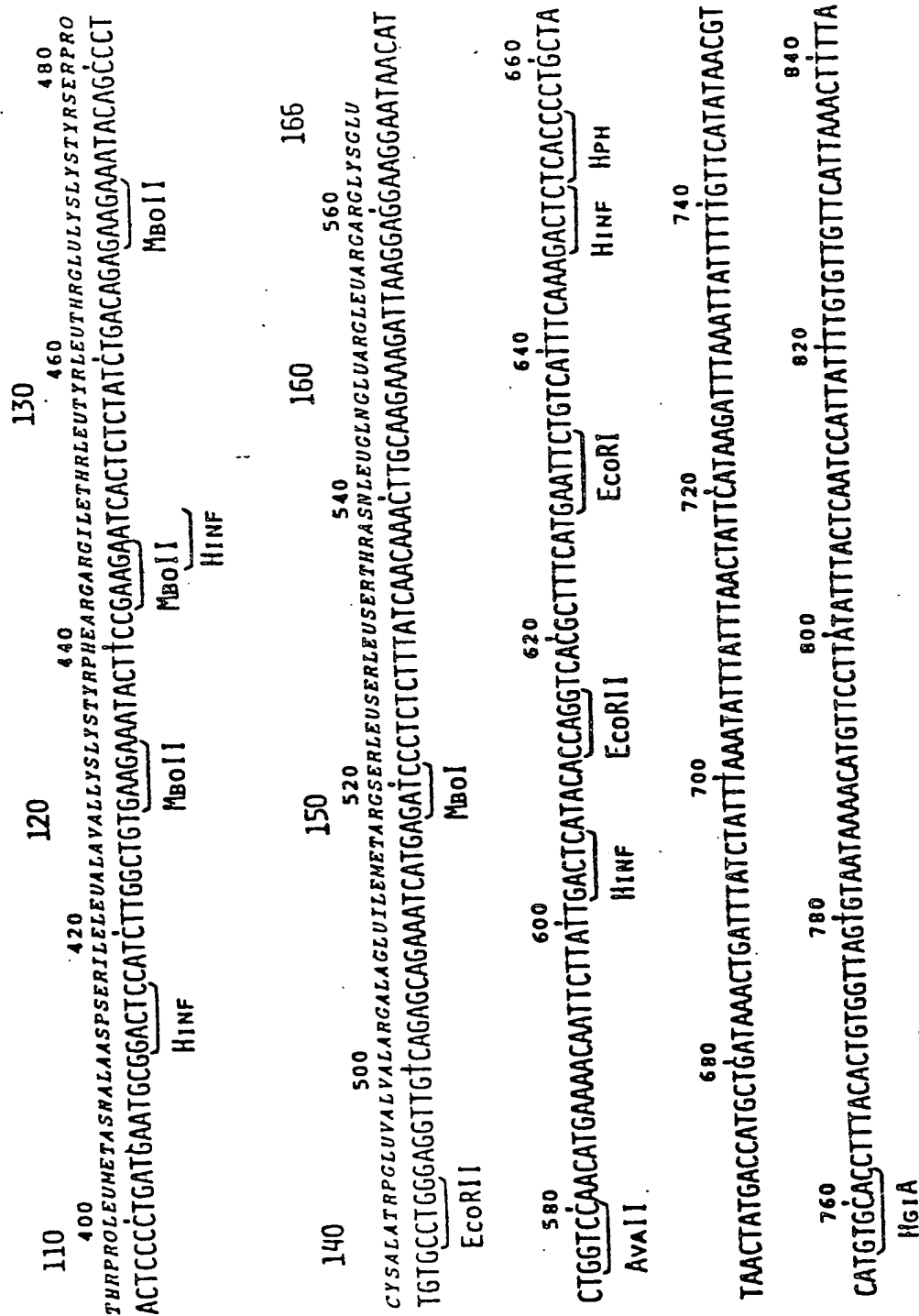


FIG. 22

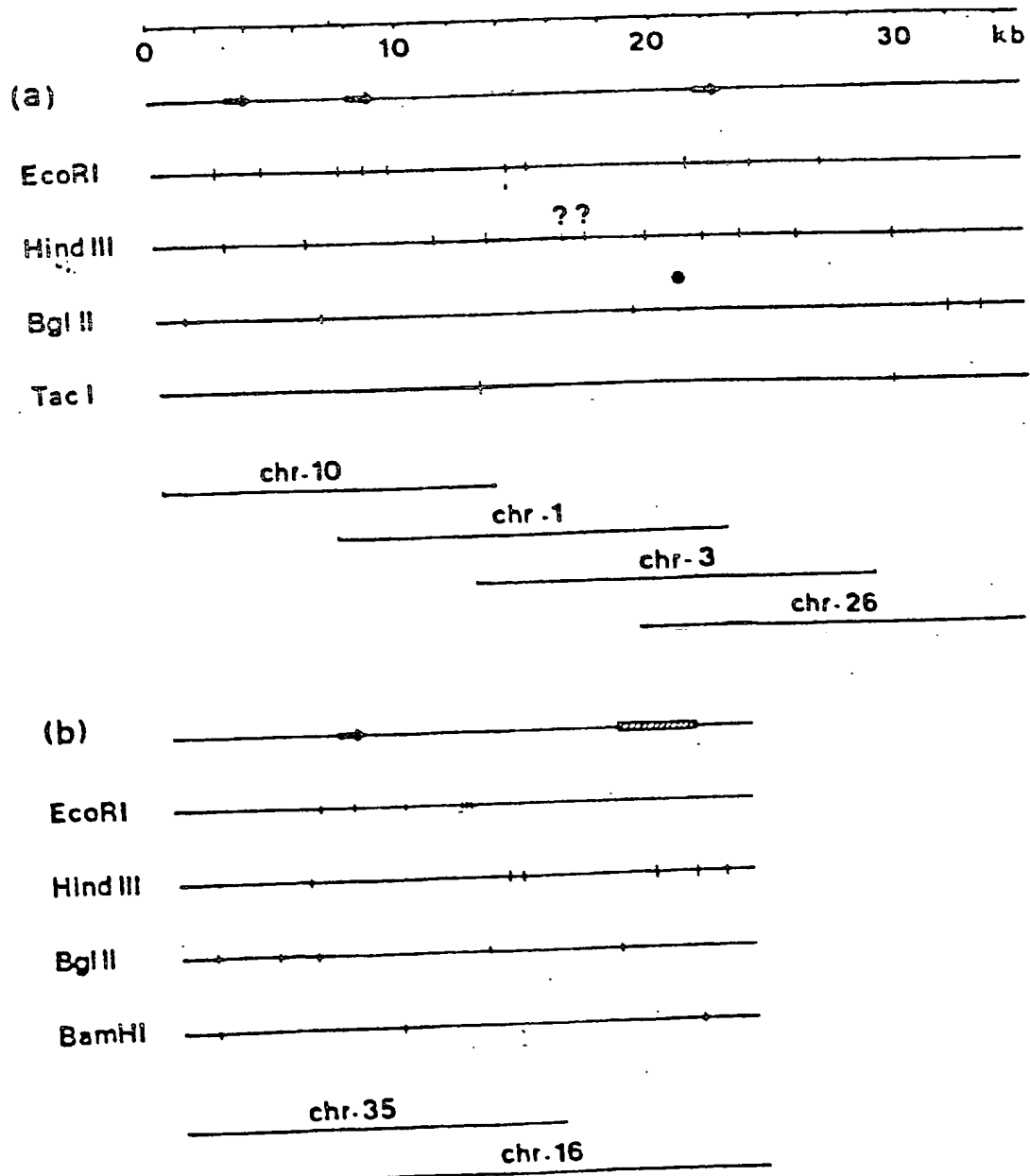
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860 CTATAGGAACTTCCTGATGIGTTCATTCTTTAATAIGAATTCCTAGCCTGACTGTGCAACCTGATTAGAGAATAAAGGGTATATTTTA 920
880
940 TTGCTTATCATTATTATAIGTAAGA 959

FIG. 23

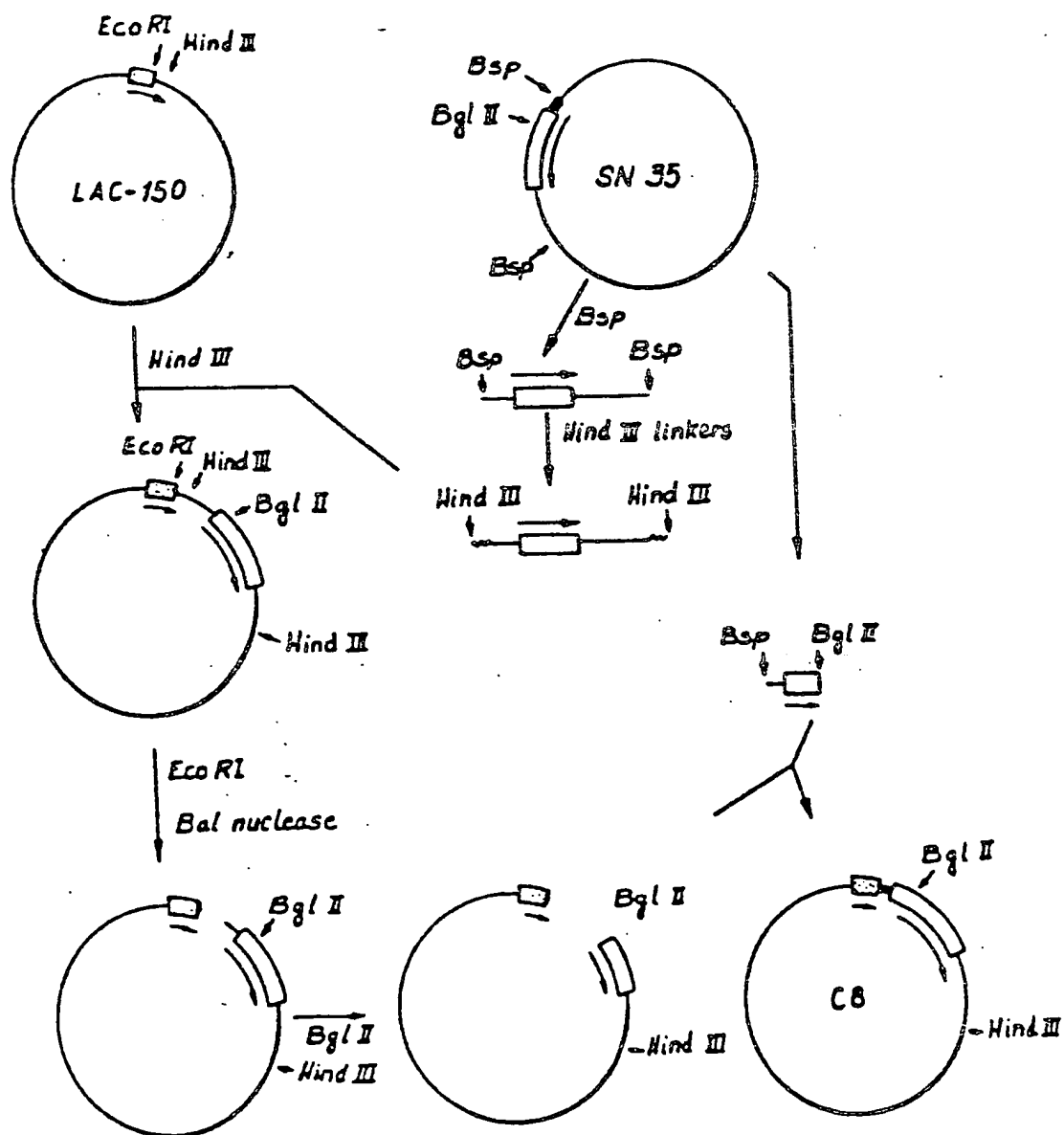
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FIG. 24

LINKAGE OF IFN- α RELATED GENES

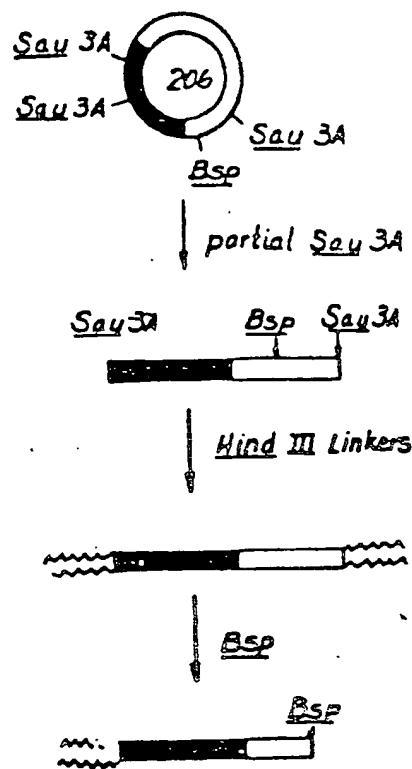
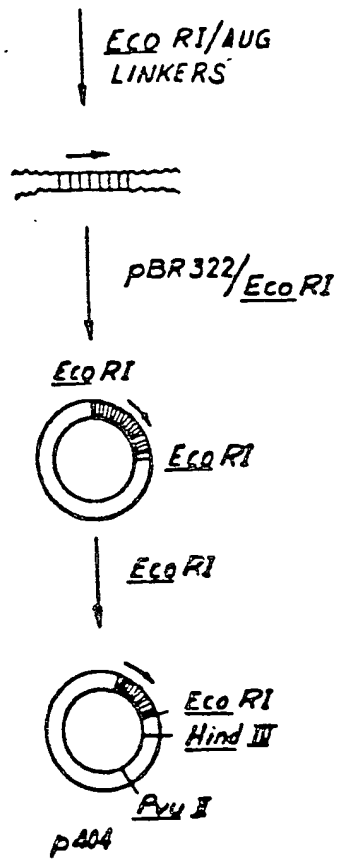
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FIG. 25

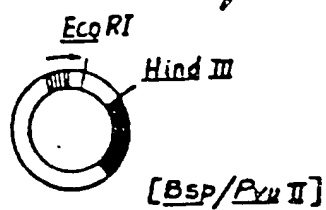


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FIG. 26

LAC-Alu Fragment



Hind III
Pvu II



*Eco*RI/*Hind*III
S1 Exonuclease



LAC-AUG (α2)

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FIG. 27

CONSTRUCTION OF PLASMID LAC-AUG (α -2)

LAC PROMOTER

MET
 ...ACAGCTATG...
 ...TGTCGATAC...

ALUI

Eco LINKER

...AAACAG + CTATGAATTCATAG
 ...TTTCTC + GATACTTAAGTATC

LIGASE

...ACAGCTATGAATTCATAG
 ...TGTGATACTTAAGTATC

EcoRI
 S₁ NUCLEASE

IFN- α CDNA

LEU GLY CYS ASP
 CTGGGCTGTGAT...
 GACCCGACACTA...

Sau 3A

ACAAGCTTGT + GAT...
 TGTTCGAACA + CTA...

LIGASE

CYS ASP
 ACAAGCTTGTGAT...
 TGTTCGAACACTA...

Hind III
 S₁ NUCLEASE

... ACAGCTATG TGTGAT ...
 ... TGTGATAC AACTA ...

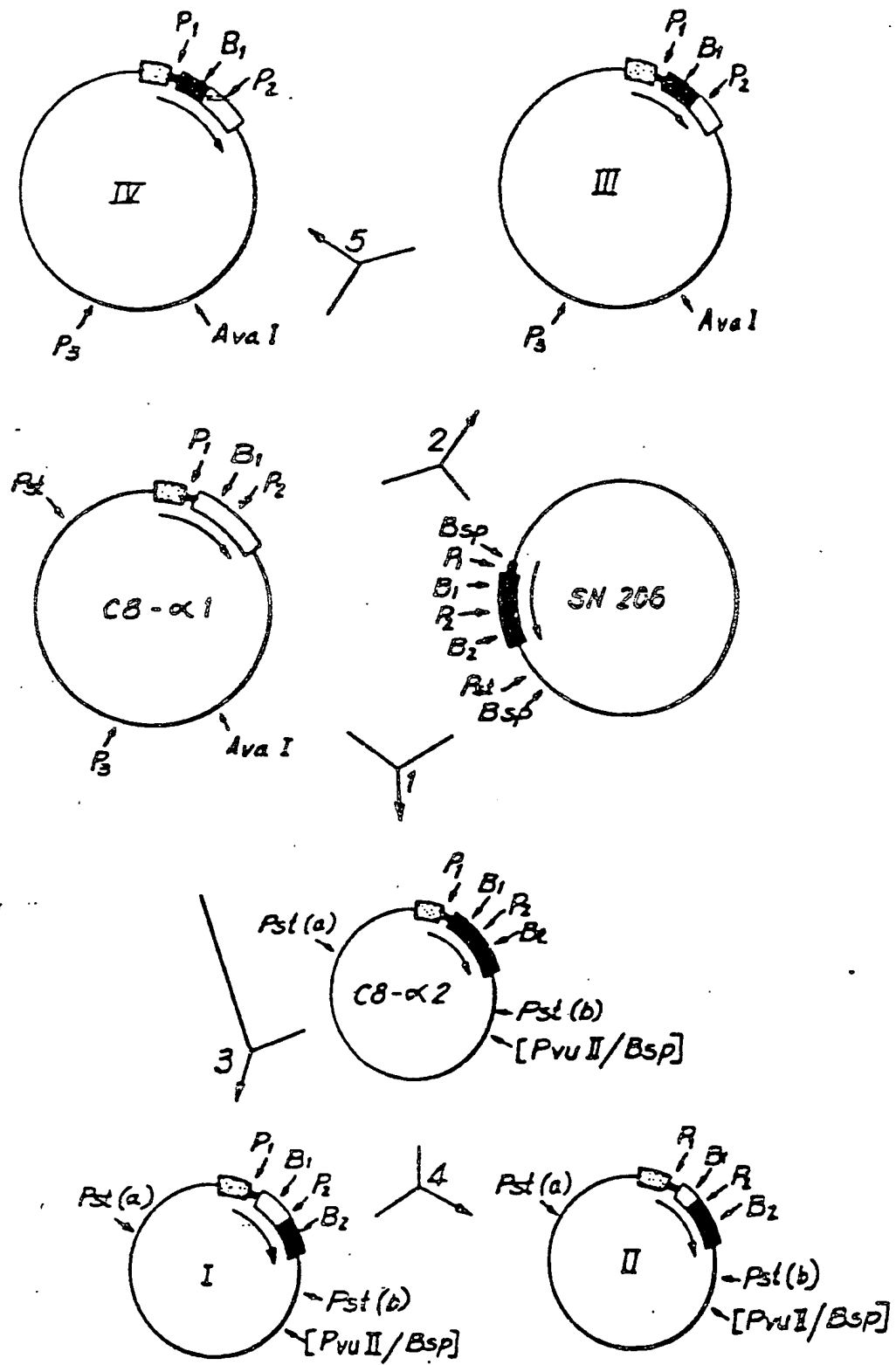
LIGASE

Met CYS ASP

... ACAGCTATGTGTGAT...
 ... TGTGATACAACTA...

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FIG. 28



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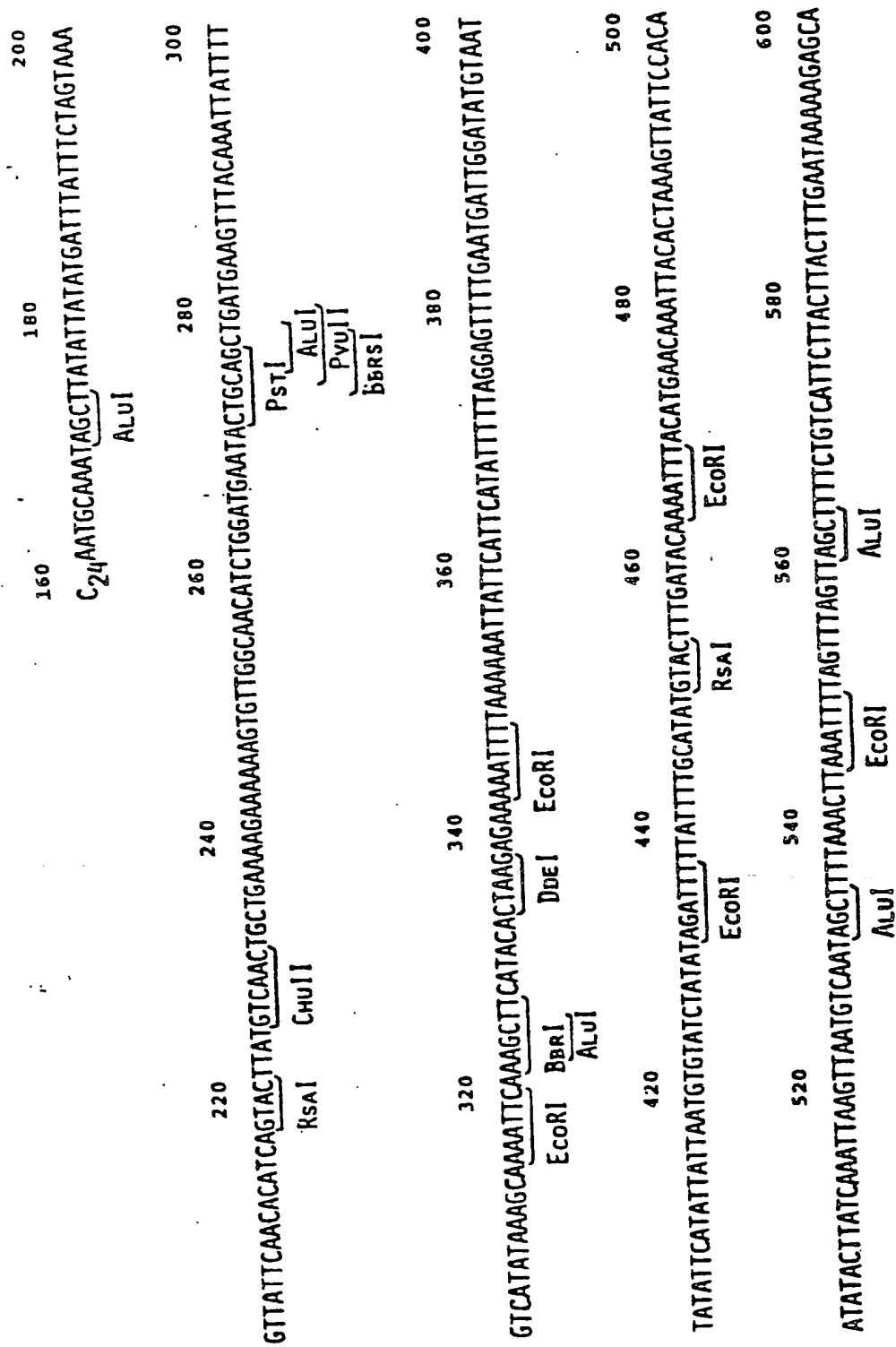


FIG. 29

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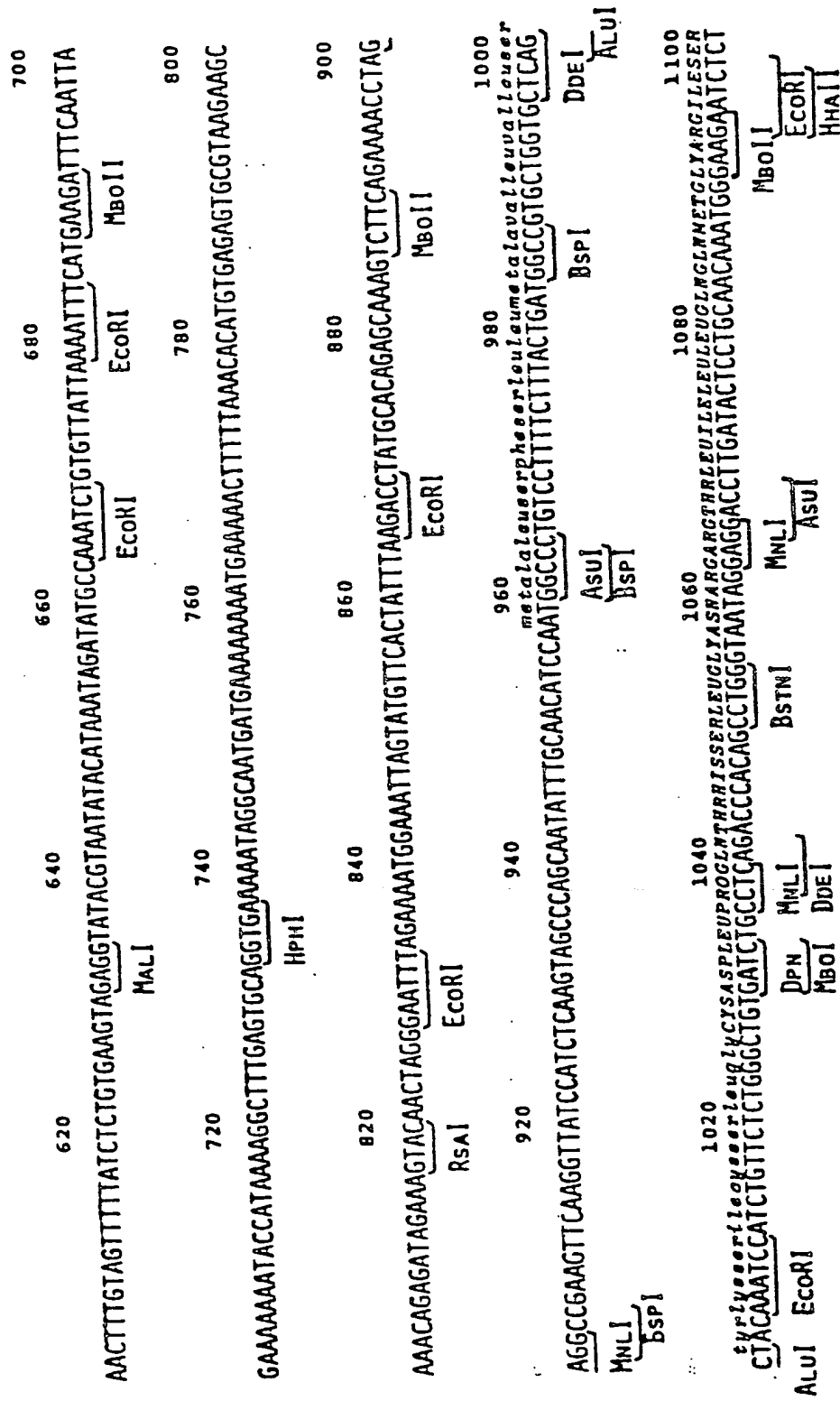


FIG. 30

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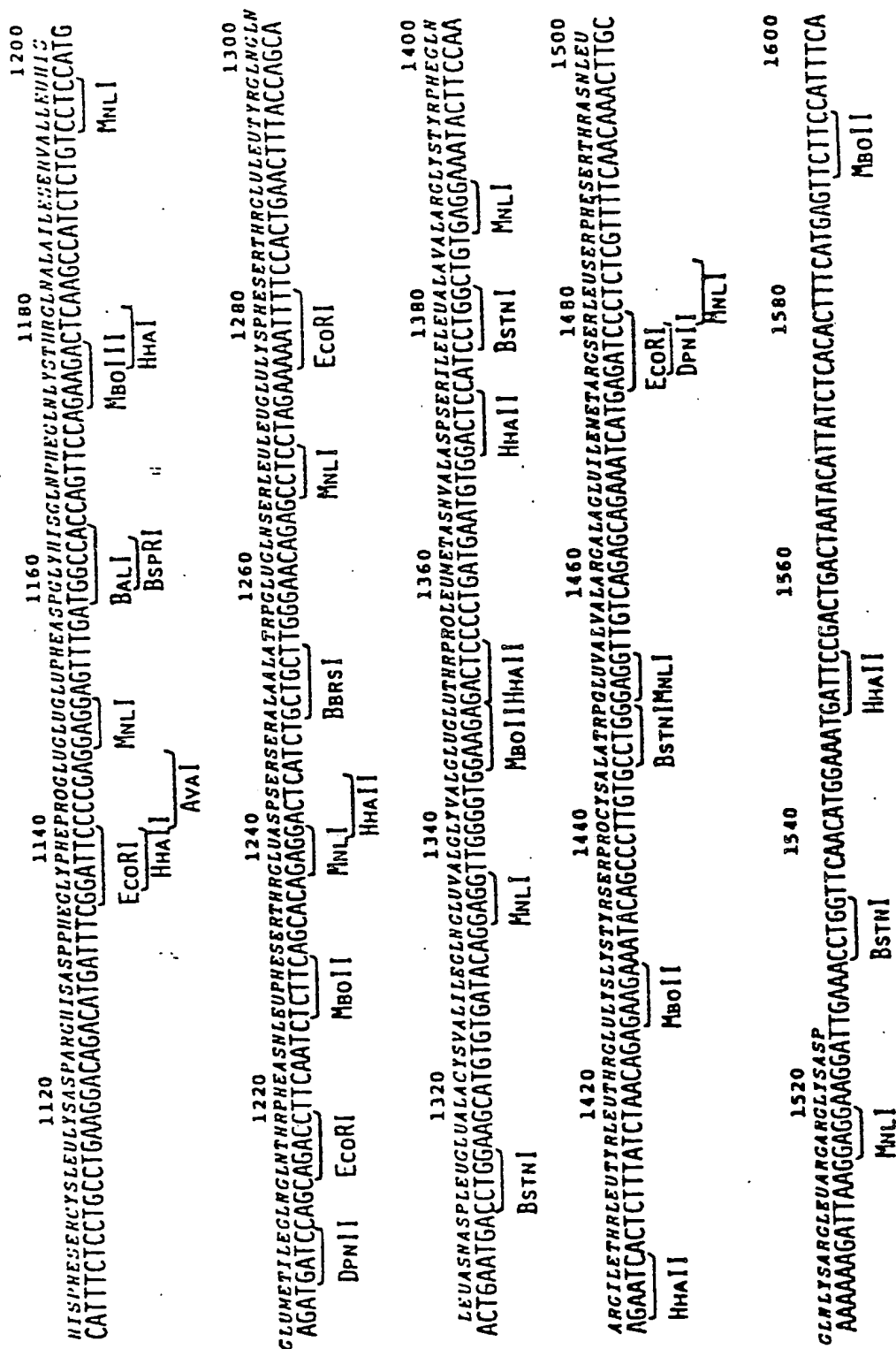


FIG. 31

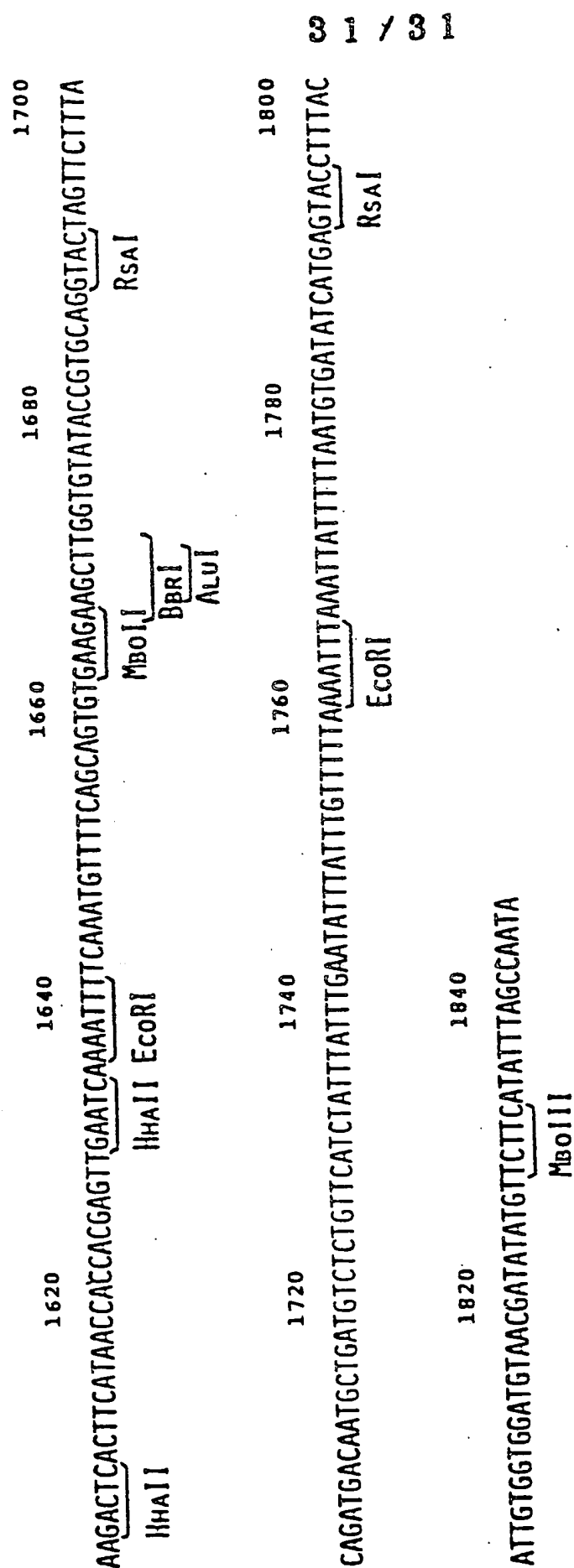


FIG 32

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